

Agilent HS RNA Kit (15 nt) 5200 Fragment Analyzer System

Quick Guide

For Research Use Only.

Not for use in diagnostic procedures.

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Kit Specifications

| Specifications | Description | |
|---|---|--|
| RNA sizing range | 200 nt - 6,000 nt | |
| RNA sizing accuracy ¹ | ± 20% (Total RNA and mRNA) | |
| RNA sizing precision ¹ | 20% CV (Total RNA and mRNA) | |
| Limit of detection | 50 pg/μL (Total RNA); 250 pg/μL (mRNA) | |
| Quantitative range (per smear) | 50 pg/μL to 5,000 pg/μL (Total RNA); 250 pg/μL to 5,000 pg/μL (mRNA) | |
| RNA quantification accuracy ¹ | ± 30% | |
| RNA quantification precision ¹ | 20% CV | |

¹ Using RNA Ladder as sample.

Kit Components

Agilent HS RNA kit, 15 nt (DNF-472)

| Part Number | Name | |
|--------------|---|--|
| DNF-265 | RNA Separation Gel | |
| DNF-600-U030 | Intercalating Dye | |
| DNF-355 | 5x 930 dsDNA Inlet Buffer (Dilute to 1x) | |
| DNF-475 | 5x Capillary Conditioning Solution (Dilute to 1x) | |
| DNF-497 | 0.25x TE Rinse Buffer | |
| DNF-370 | HS RNA Diluent Marker (15 nt) | |
| DNF-386 | HS RNA Ladder | |
| DNF-301 | BF-1 Blank Solution | |
| GP-440-0100 | Capillary Storage Solution (sold separately) | |

Analysis Protocol

Gel preparation

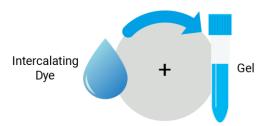
Prepare gel/dye mixture for 5200 Fragment Analyzer system:

| # of samples to be analyzed1 | Volume of Intercalating Dye | Volume of gel |
|------------------------------|-----------------------------|---------------|
| 12 | 1.0 μL | 10 mL |
| 24 | 1.5 µL | 15 mL |
| 36 | 2.0 μL | 20 mL |
| 48 | 2.5 μL | 25 mL |
| 96 | 4.5 µL | 45 mL |

¹ Typically one sample well per separation is dedicated to the ladder.

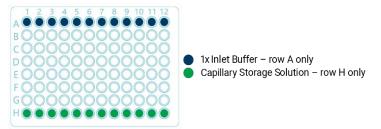
Instrument and sample preparation

1 Mix fresh gel and dye. Refill 1x Capillary Conditioning Solution as needed.1.0 μL dye / 10 mL gel

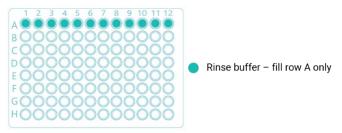


2 Place a fresh 1x 930 dsDNA Inlet Buffer tray on the 5200 Fragment Analyzer system. Fill row A (1.0 mL/well). Replace 1x 930 dsDNA Inlet Buffer daily.

3 Place Capillary Storage Solution in row H (1.0 mL/well). Replace Capillary Storage Solution every 2-4 weeks.



4 Place 0.25x TE Rinse Buffer plate in marker drawer location. Fill row A (200 μ L/well). Replace 0.25x TE Rinse Buffer daily.

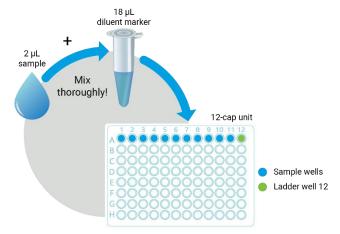


5 Dilute the as received HS RNA Ladder to a working concentration of 2 ng/µL*.

NOTE

*Initial step done when using as received HS RNA Ladder aliquot for the first time only. Proceed to Step 6 if already completed.

- **6** Heat denature samples and HS RNA Ladder at 70°C for 2 minutes, immediately cool to 4°C and keep on ice before use.
- 7 Mix samples or HS RNA Ladder with HS RNA Diluent Marker (15 nt) in sample plate. Add 20 μ L of BF-1 Blank Solution to any unused wells of the row to be analyzed.



WARNING

Working with Chemicals

The handling of reagents and chemicals might hold health risks.

- Refer to product material safety datasheets for further chemical and biological safety information.
- Follow the appropriate safety procedures such as wearing goggles, safety gloves and protective clothing

Fragment Analyzer software operating procedure

- 1 Select **Tray** and **Row** to run for 12-Cap.
- 2 Enter Sample ID and Tray ID (optional).
- 3 Select Add to Queue, select the DNF-472(M or T)-(22, 33 or 55) HS Total RNA (or mRNA) 15nt method from the Dropdown menu.
- 4 Enter Tray Name, Folder Prefix, and Notes (optional).
- **5** Select **OK** to add method to the queue.
- **6** Select **▶** to start the separation.

NOTE

Please refer to the Kit Guide for additional details.

Technical Support and Further Information

For technical support, please visit www.agilent.com.

It offers useful information, support and current developments about the products and technology.

www.agilent.com

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