

# Sensitive Detection of 2-Methoxy-3-Isobutylpyrazine (IBMP) and Trichloroanisole (TCA) in Wine Using Triple Quadrupole GC/MS

Application Chemists: Stephan Baumann, Anna Hjelmeland

# General recommendations for wine analysis

## Extraction:

- Headspace-solid phase microextraction (HS-SPME) is an inexpensive and simple extraction procedure
- Other options are stirbar sorptive extraction (SBSE) or headspace stirbar sorptive extraction (HSSE)

## Increasing headspace vapor pressure improves response:

- Saturating the solution with a salt like sodium chloride increases the concentration for some analytes in the headspace depending on the sample matrix.
- Increasing the temperature but beware of degradation/artifacts
- Use agitation, if possible

## Optimizing the stationary phase selection

- PDMS, PDMS/DVB, PDMS/DVB/CAR

## HS-SPME advice

- Maximize extraction/equilibration time for less volatile compounds
- Use Internal Standards that mimic the physicochemical properties of analytes to normalize for variations in extraction efficiency
- Try to use a single fiber for an experiment. This is especially important if there are no internal standards.

## 3-Alkyl-2-Methoxypyrazines

Sauvignon Blanc, Sémillon and Cabernet Sauvignon  
and other Bordeaux varietals

### Types:

- IBMP: 2-Isobutyl-3-methoxypyrazine; green bell pepper aroma
- IPMP: 2-Isopropyl-3-methoxypyrazine; green pea earthy nutty
- SBMP: 2-Sec-butyl-3-methoxypyrazine; musty vegetative
- EMP: 2-Ethyl-3-methoxypyrazine; raw potato earthy bell pepper nutty

### Sources:

- Natural biosynthesis in plants and microbes
- Ladybugs (IBMP)
- Grapevine Stems (SBMP)



MALB

# Methoxypyrazines

## Concentrations need to be managed to make good wines

Canopy management steps practiced to reduce presence of methoxypyrazines:

- Expose grape clusters to sunlight before the onset of ripening
- Reduce the number of leaves between grape clusters
- Vines with excessive growth will generate more methoxypyrazines
- Grapes picked before sugar maturity will have higher concentrations of methoxypyrazines

### Reference:

Justin J. Scheiner, Gavin L. Sacks, Justine E. Vanden Heuvel, “How Viticulture Factors Affect Methoxypyrazines”, *Wines & Vines*, 117-120 (2009).

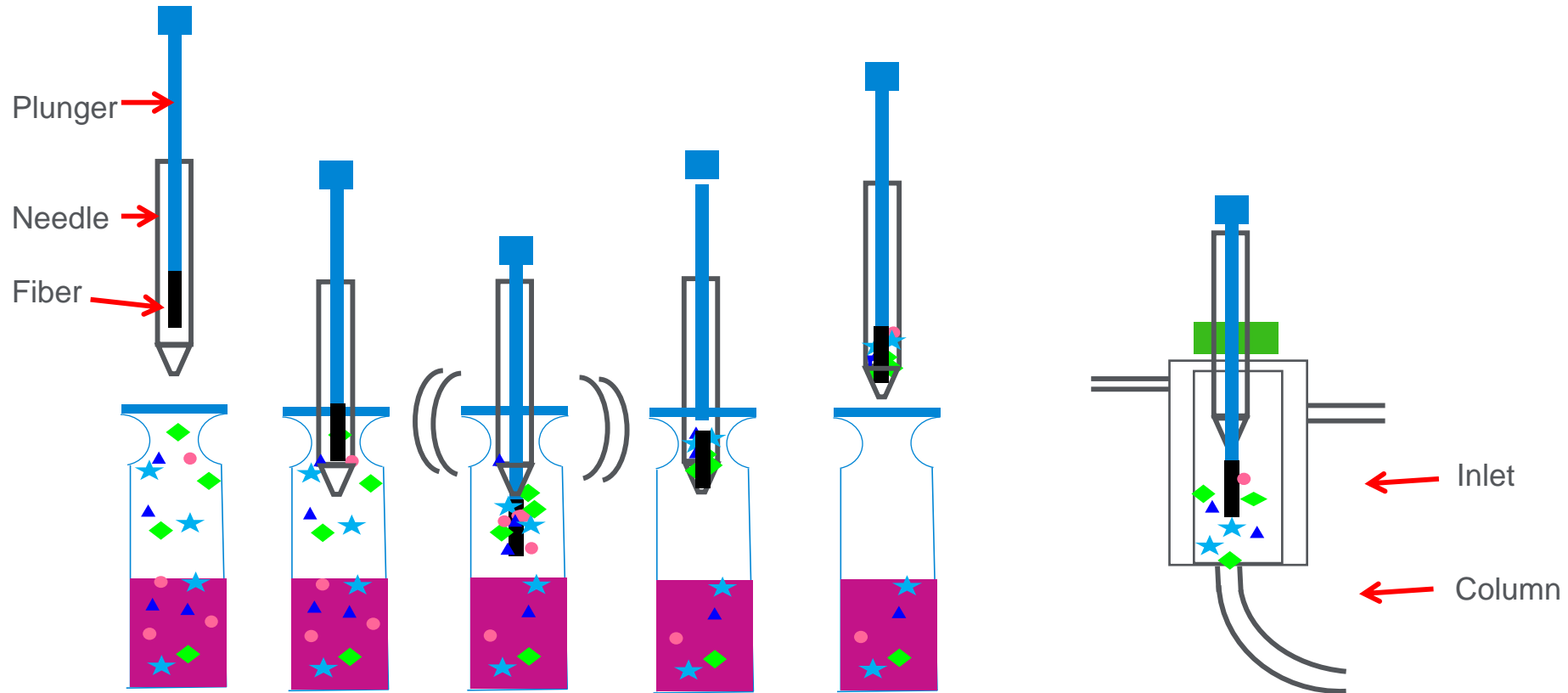
# IBMP Sample Preparation

- 20 mL headspace vials
- IBMP calibration standards are made in model wine (0.5 % w/v tartaric acid in 12 % v/v ethanol).
- Sauvignon Blanc and Cabernet Sauvignon wine were used to spiked samples
- 5 mL Aliquots taken
- Standards and spikes made at 0, 5, 20, and 100 ng/L in MIBP, and 80 ng/L in isotopically labeled IBMP
- 2 g NaCl added

## Reference:

R. Godelman, S. Limmert, T. Kuballa, "Implementation of headspace solid-phase-micro-extraction-GC-MS/MS methodology for determination of 3-alkyl-2-methoxypyrazines in wine", Eur Food Res Technol 227, 449-461 (2008).

# Solid Phase Microextraction (SPME) Steps



The 1 cm 50/30  $\mu\text{m}$  DVB/Carboxen/PDMS fiber SPME samples were equilibrated at ambient temperature  $\sim 25^\circ\text{C}$  for 30 min. The fiber was then lowered into the headspace and the samples were extracted for 30 min before being desorbed in the inlet for two minutes. At that time the inlet was purged. The fiber was left in the inlet to clean for an additional nine minutes.

# IBMP GC Run Conditions

| GC Run Conditions         |   |
|---------------------------|---|
| Analytical Column         | Two 15 m x 0.25 mm x 0.25 µm HP-5msUI columns (P/N 190915-433UI)                                  |
| Inlet temperature         | 250°C   |
| Inlet pressure            | 9.5 psi   |
| Carrier gas               | Helium, constant flow mode, 1.2 mL/min  |
| Splitless                 | Purge 50 mL/min @ 2 min   |
| Oven program              | 45°C (2.25min hold), 8°C/min to 130 °C  |
| Column velocity           | 39.8 cm/s   |
| Injection                 | SPME (PDMS/DVB/CAR) ; 2 min.; 250°C   |
| Transfer line temperature | 250 °C  |
| GC Post-Run Conditions    |   |
| Backflush device          | Purged Ultimate Union (P/N G3186-60580) controlled by a Pressure Control Module (P/N G3476-60501) |
| Backflush conditions      | -1.2 mL/min@200 °C for 2 min  |

Note the SPME fiber was equilibrated for 30 min at room temperature since I didn't have any fancy equipment to heat, agitate (automate) sample adsorption.

# IBMP MS Conditions

| MS Conditions          |   |
|------------------------|---|
| Tune                   | PCI Autotune                                  |
| Delta EMV              | 800V  |
| Acquisition parameters | PCI; selected reaction monitoring             |
| Reagent Gas Flow       | 20 % Methane                                  |
| Collision Gas Flows    | Nitrogen at 1.5 mL/min, Helium at 2.35 mL/min |
| Solvent delay          | 3.75 minutes                                  |
| MS temperatures        | Source 300 °C; Quadrupoles 150 °C             |

| Compound  | Triple Quadrupole GC/MS |         |                 |                       |
|---|-------------------------|---------|-----------------|-----------------------|
|   | RT (min)                | SRM     | Dwell Time (ms) | Collision Energy (EV) |
| IBMP  | 11.5                    | 167→94  | 60              | 35                    |
|   |                         | 195→124 | 60              | 30                    |
|   |                         | 195→106 | 60              | 35                    |
| <sup>2</sup> H <sub>3</sub> -IBMP (Internal Standard) | 11.5                    | 170→127 | 20              | 30                    |
|   |                         | 170→128 | 20              | 30                    |
|   |                         | 170→100 | 20              | 30                    |

Note that Anna also identified a 167→125 (15 EV) qualifier transition for IBMP.



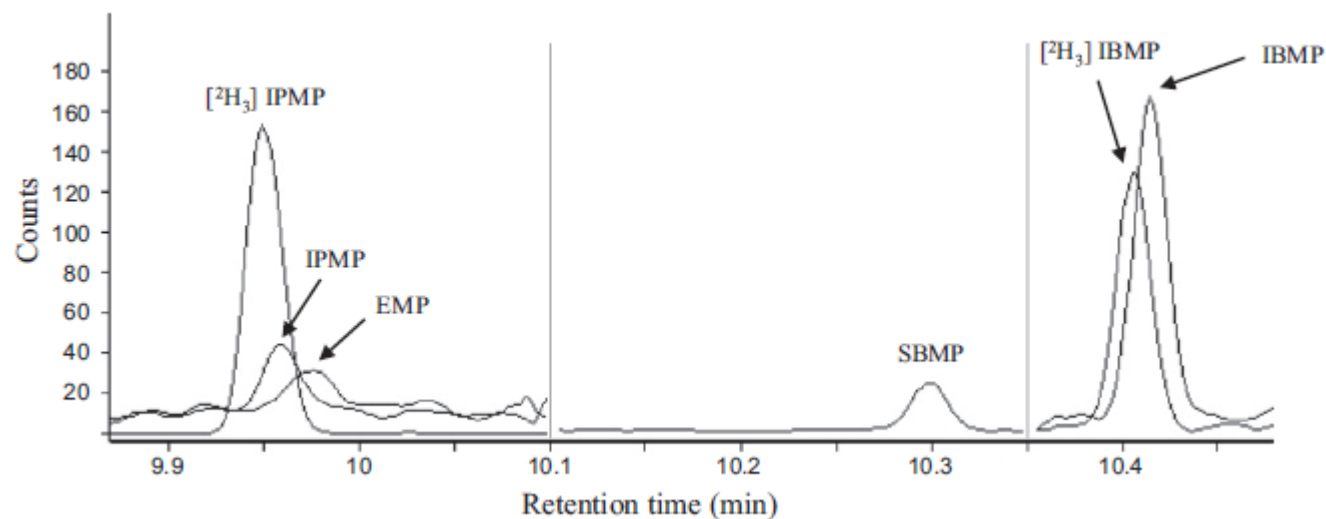
# IPMP, SBMP, EMP MS Conditions

Anna Hjelmeland: Talanta 148 (2016) 336-345

| Compound                          | Triple Quadrupole GC/MS |         |                     |                            |
|-----------------------------------|-------------------------|---------|---------------------|----------------------------|
|                                   | RT (min)                | SRM     | Collision Energy EV | Quantifier/Qualifier Ratio |
| <sup>2</sup> H <sub>3</sub> -IPMP | 9.95                    | 156→121 | 15                  | 1.17                       |
|                                   |                         | 156→123 | 30                  |                            |
| IPMP                              | 9.96                    | 153→121 | 15                  | 1.04                       |
|                                   |                         | 153→123 | 25                  |                            |
| EMP                               | 9.98                    | 139→107 | 15                  | 0.9                        |
|                                   |                         | 139→124 | 20                  |                            |
| SBMP                              | 10.1                    | 167→138 | 20                  | 1.23                       |
|                                   |                         | 167→123 | 30                  |                            |

In addition to IBMP, three other methoxypyrazines IPMP (2-iso-propyl-3-methoxypyrazine), SBMP (2-sec-butyl-3-methoxypyrazine), and EMP (2-ethyl-3-methoxypyrazine) have been identified in grapes and/or wine and can impact aroma quality. Anna used a 30 m x 0.32 mm i.d. x 1.0 µm film thickness DB-WAXetr capillary column for optimal separation of all four methoxypyrazines.

# Chromatographic Separation and Results from Talanta article

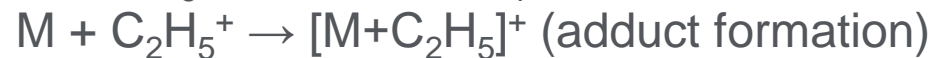


| Compound | Sensory Threshold In Wine (ng/L) | Standard Curve Range (ng/L) | Correlation Coefficient R | LOQ (ng/L) | LOD (ng/L) |
|----------|----------------------------------|-----------------------------|---------------------------|------------|------------|
| IBMP     | 8 - 15                           | 0.5 - 50                    | 0.993                     | 0.5        | 0.25       |
| IPMP     | 0.3 - 1.6                        | 0.5 - 50                    | 0.986                     | 1          | 0.5        |
| SBMP     | -                                | 0.5 - 50                    | 0.992                     | 0.5        | 0.25       |
| EMP      | -                                | 0.5 - 50                    | 0.99                      | 1          | 0.5        |

# What is Positive Chemical Ionization?

- Reagent ions are formed from a "reagent gas" by bombardment with electrons
- Reagent gas ions undergo subsequent reactions with sample molecules to form sample ions ("Brönsted acids/bases")

Example:

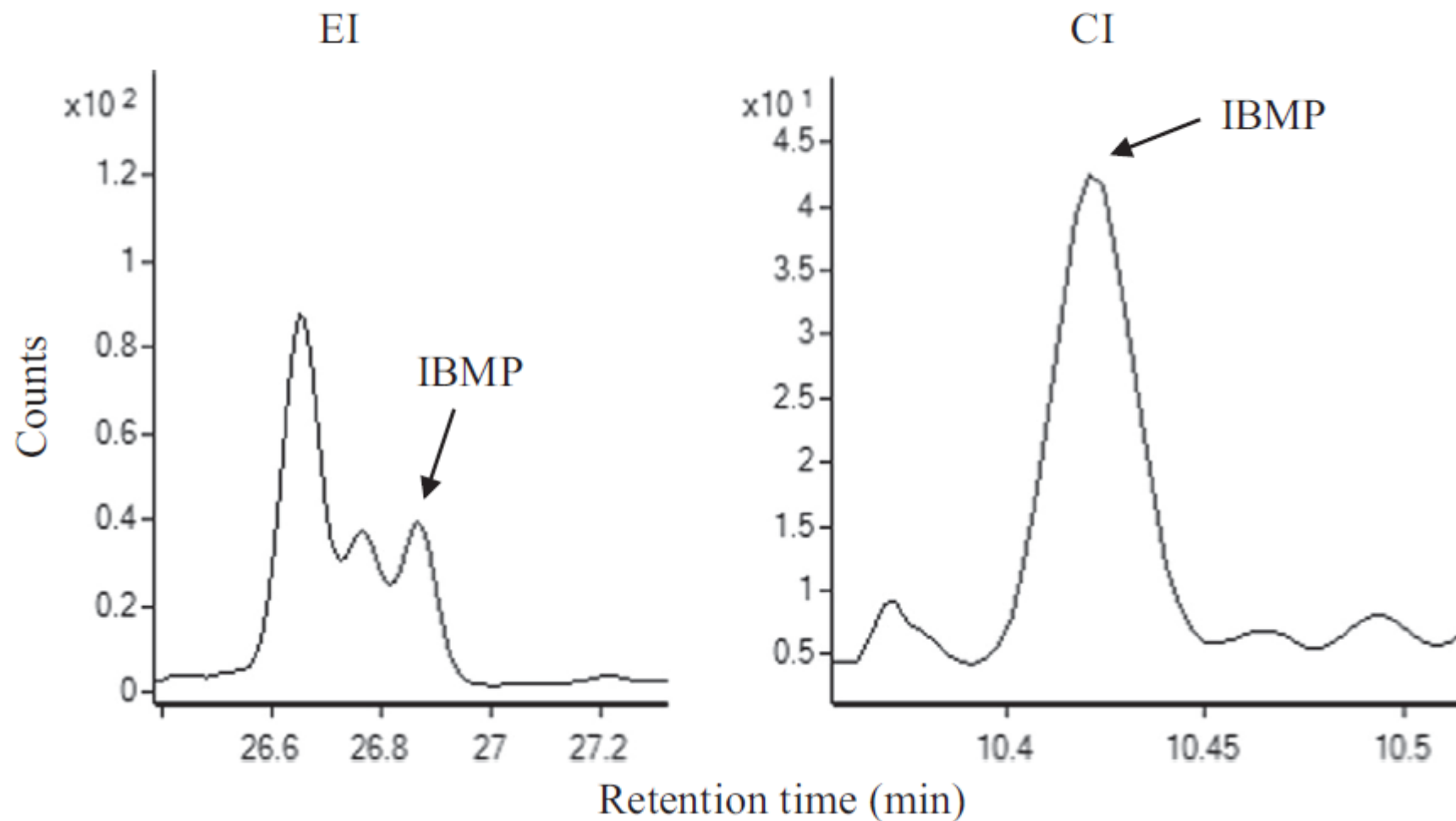


## Chemical Ionization (CI) Attributes

- CI ion formation is much more "gentle" than electron ionization (EI) (less fragmentation) and primarily produces the protonated molecule
- Chemical control of ionization allows selectivity through choosing different reagents gases; methane, isobutane, ammonia, etc.

Note that the proton affinity, in kcal/mole, of  $\text{CH}_5^+$  is 131.6 while the  $\text{C}_2\text{H}_5^+$  adduct is 162.6. Note that the proton affinity of ammonia is 204. There is much more energy left over for fragmentation when using methane relative to ammonia.

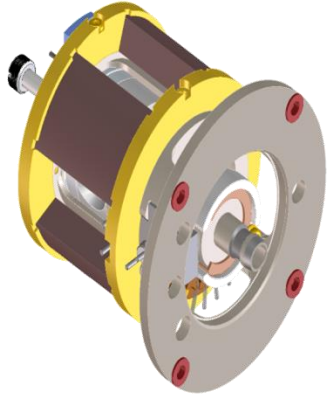
# Why not use traditional EI?



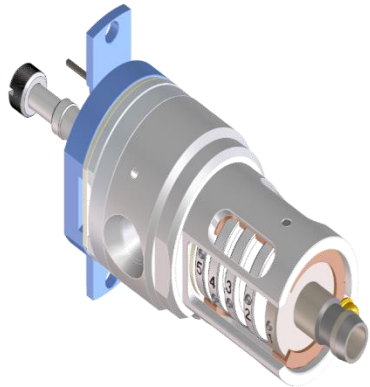
Note that even with a painfully slow chromatographic separation, IBMP is not separated from interferences using the EI source.

# Could Low Energy EI be applied on the 7010?

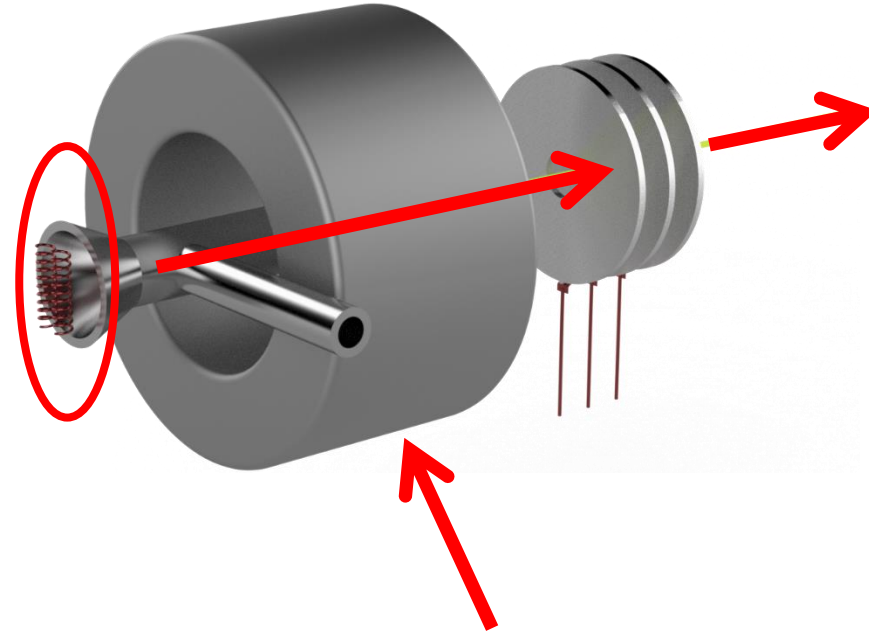
## A definite maybe!



High Efficiency Source



High Efficiency Source with  
Magnet Removed



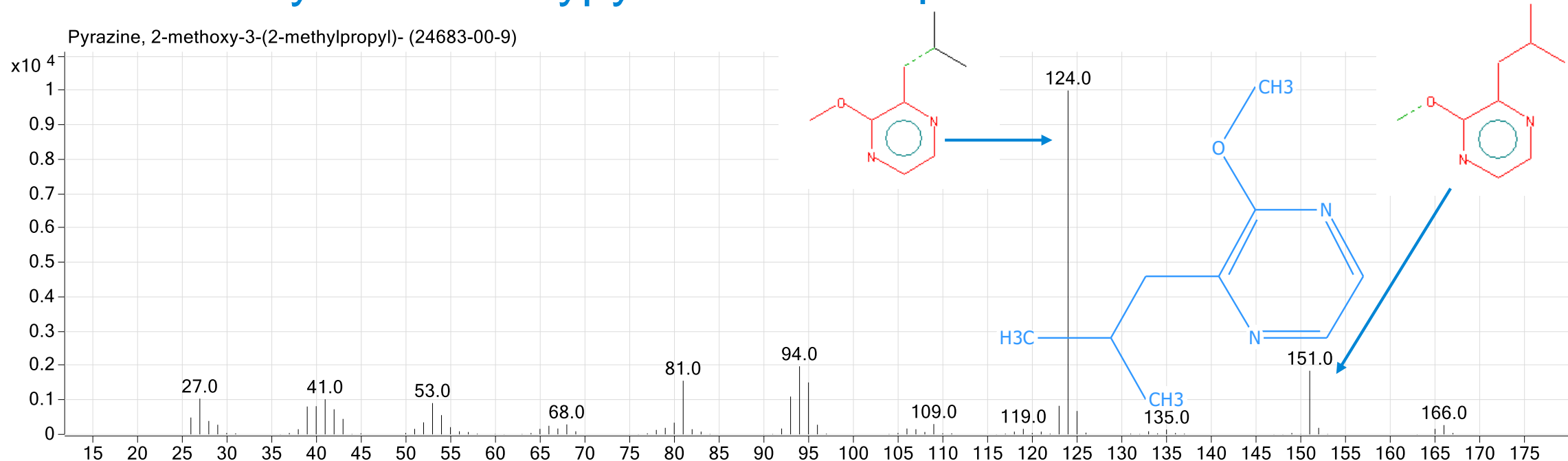
High emission filaments for more electron current

Powerful cylindrical magnet collimates electrons

Long path length for ionization

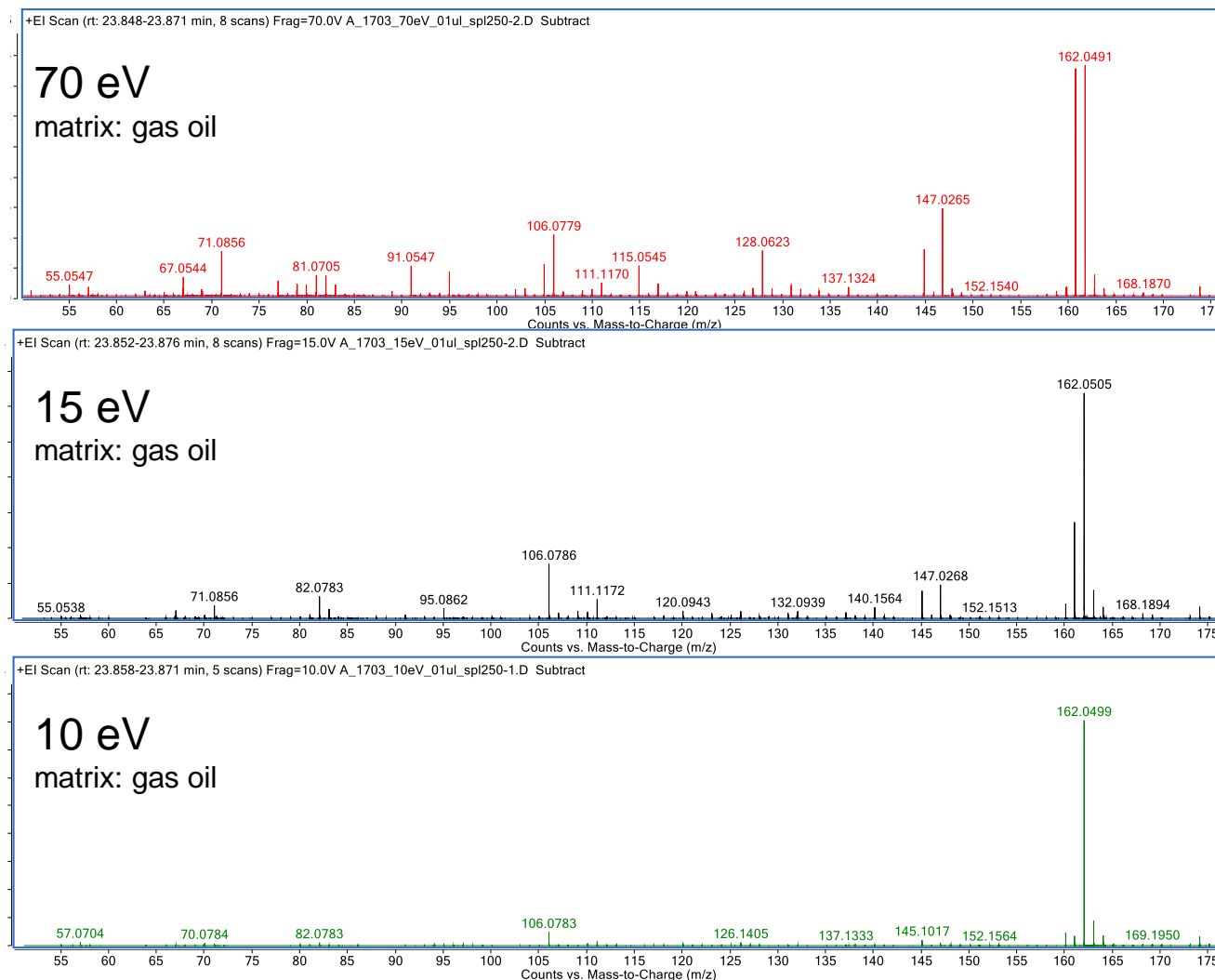
# Could Low Energy EI be applied on the 7010?

## 2-Isobutyl-3-methoxypyrazine example



Typically in EI ionization, high-energy electrons are created through thermionic emission at 70 eV. The most abundant EI fragments found at 70 eV are at 151 and 124 Da. These fragments should be greatly reduced in abundance relative to the parent ion if we reduce the ionization energy to 30, 20 or 15 eV. This should simplify the fragmentation pattern for both IBMP and the numerous interferences found in wine matrix.

# HES Low Energy EI example from the new 7250 GC/QTOF



Note that the response goes down with source voltages but the chemical interferences go down by more, often enhancing sensitivity. So please try this with methoxypyrazines and publish the results!

# Haloanisoles TCA and TBA implicated in Cork Taint

## TeCA and PCA are associated with musty aromas in beverages



### Types:

- TCA: 2,4,6-trichloroanisole
- TBA: 2,4,6-tribromoanisole
- TeCA: Tetrachloroanisole
- PCA: Pentachloroanisole

### Sources:

- Biomethylation of halophenols
- Halophenols are often anthropogenic although there are some that come from natural sources – cork bleaching, industrial pollution.

### Typical Odor:

- musty, moldy (wet newspaper) aroma



# Haloanisoles Contamination typically starts with a Halophenol

## TCP and TBP Uses

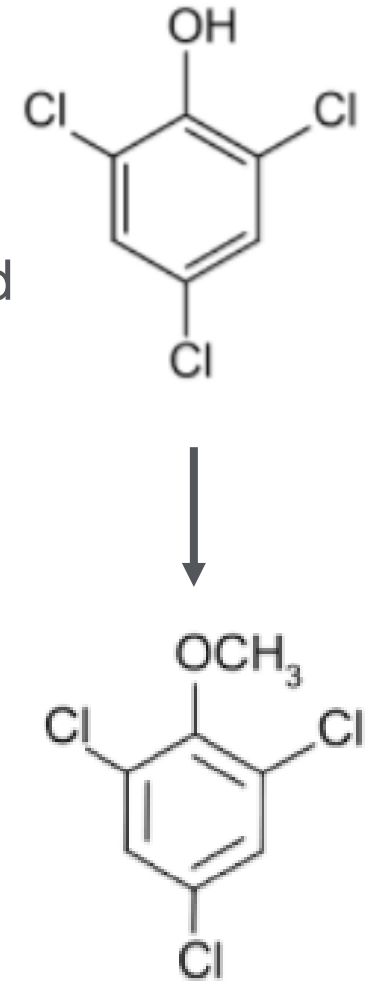
### 2,4,6-Trichlorophenol

- Also known as TCP, phenaclor, Dowicide 2S, Dowcide 2S, omal.
- It has been used as a fungicide, herbicide, insecticide, antiseptic, defoliant, and glue preservative.

### 2,4,6-Trichlorophenol

- Also known as Tribromophenol, 2,4,6-TBP, TBP.
- It is often used as a wood preservative and a flame retardant.

These ubiquitous halophenols are toxic to molds and fungi so they methylate the phenols to make them biologically harmless. This also makes the phenols more volatile!



Reference:

Robert Tracy, "2,4,6-TBA - Next 2,4,6-TCA in U.S. Wine Industry", Practical Winery & Vineyard Journal, November-December (2008).

# TCA Sample Preparation

- 20 mL headspace vials
- TCA calibration standards are made in 13 % alcohol Cabernet Sauvignon
- 5 mL Aliquots taken
- Standards and spikes made at 1, 5, 10, 15, and 20 ng/L in TCA, and 10 ng/L in 2,3,6-TCA as an internal standard.
- 2 g NaCl added

# TCA GC Run Conditions

| GC Run Conditions         |   |
|---------------------------|---|
| Analytical Column         | Two 15 m x 0.25 mm x 0.25 µm HP-5msUI columns (P/N 190915-433UI)                                  |
| Inlet temperature         | 250 °C  |
| Carrier gas               | Helium, constant flow mode, 3 mL/min  |
| Splitless                 | Purge 50 mL/min @ 2 min   |
| Oven program              | 45°C (2 min hold), 25 °C/min to 215 °C  |
| Injection                 | SPME (PDMS/DVB/CAR) ; 2 min.; 250°C   |
| Transfer line temperature | 280 °C  |
| GC Post-Run Conditions    |   |
| Backflush device          | Purged Ultimate Union (P/N G3186-60580) controlled by a Pressure Control Module (P/N G3476-60501) |
| Backflush conditions      | -5 mL/min @ 250 °C for 2 min  |

# TCA MS Conditions

| MS Conditions          |   |
|------------------------|---|
| Tune                   | Autotune                                      |
| Gain                   | 20  |
| Acquisition parameters | El; selected reaction monitoring              |
| Collision Gas Flows    | Nitrogen at 1.5 mL/min, Helium at 2.35 mL/min |
| Solvent delay          | 6.5 minutes                                   |
| MS temperatures        | Source 300 °C; Quadrupoles 150 °C             |

| Compound                    | Triple Quadrupole GC/MS |         |                 |                       |
|-----------------------------|-------------------------|---------|-----------------|-----------------------|
|                             | RT (min)                | SRM     | Dwell Time (ms) | Collision Energy (EV) |
| 2,4,6-Trichloroanisole      | 7.594                   | 210→195 | 25              | 15                    |
|                             |                         | 167→83  | 25              | 20                    |
| 2,3,6-Trichloroanisole (IS) | 7.867                   | 210→195 | 25              | 10                    |
|                             |                         | 210→167 | 25              | 20                    |
|                             |                         | 167→83  | 25              | 20                    |

Note that Anna also identified a 212→197 (10 EV) qualifier transition for TCA.

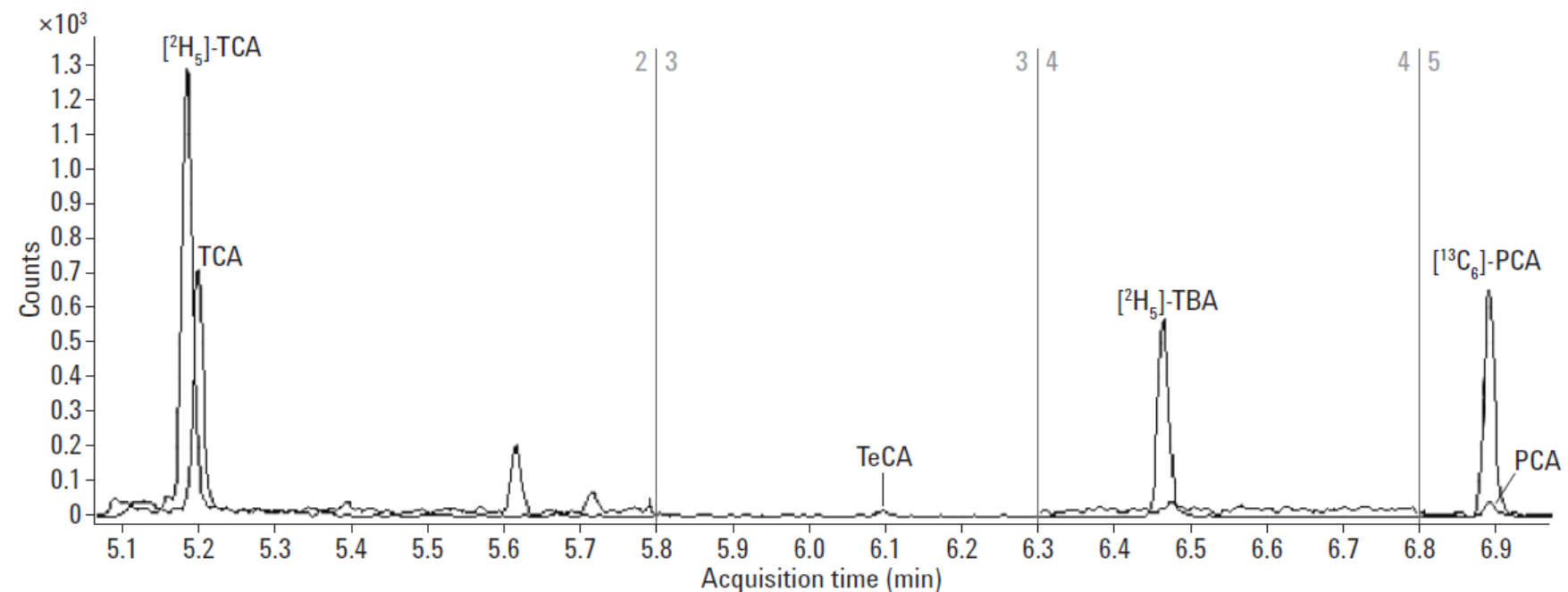
# Haloanisole MS Conditions

See Agilent Application note 5991-3812EN for details

| Compound                         | Triple Quadrupole GC/MS |         |                       |
|----------------------------------|-------------------------|---------|-----------------------|
|                                  | RT (min)                | SRM     | Collision Energy (EV) |
| <sup>2</sup> H <sub>5</sub> TCA  | 5.20                    | 215→197 | 10                    |
|                                  |                         | 217→199 | 10                    |
| TeCA                             | 6.10                    | 246→203 | 25                    |
|                                  |                         | 231→203 | 15                    |
| TBA                              | 6.50                    | 344→329 | 10                    |
|                                  |                         | 346→331 | 10                    |
| <sup>2</sup> H <sub>5</sub> TBA  | 6.48                    | 351→333 | 15                    |
|                                  |                         | 349→331 | 15                    |
| PCA                              | 6.91                    | 265→237 | 10                    |
|                                  |                         | 280→237 | 25                    |
| <sup>13</sup> C <sub>6</sub> PCA | 6.91                    | 286→242 | 25                    |
|                                  |                         | 286→271 | 10                    |

A 30 m DB-5 column was used for this study, like the TCA methodology. The flow rate was dropped from 3 to 1.2 mL/min. The oven parameters were also similar but without an initial hold time.

# Chromatographic Separation and Results



| Compound | Sensory Threshold in Wine (ng/L) | Standard Curve Range (ng/L) | Correlation Coefficient (R) | LOQ (ng/L) | LOD (ng/L) |
|----------|----------------------------------|-----------------------------|-----------------------------|------------|------------|
| TCA      | 3                                | 0.1 - 50                    | 0.999                       | 0.5        | 0.1        |
| TeCA     | 15                               | 0.1 - 50                    | 0.999                       | 0.1        | <0.1       |
| PCA      | 3                                | 0.1 - 50                    | 0.999                       | 0.25       | 0.1        |
| TBA      | 10,000                           | 0.5 - 50                    | 0.999                       | 1          | 0.5        |

# In Summary

- Headspace-solid phase microextraction (HS-SPME) is an inexpensive and sensitive extraction procedure
- Canopy management steps can reduce presence of methoxypyrazines
- 2-Alkyl-3-Methoxypyrazines work well by methane PCI
- 2-Alkyl-3-Methoxypyrazines don't work well by electron ionization (EI) without derivatization
- It is probable that Low Energy EI with the High Efficiency Source on the 7010 GC/QQQ would also work
- Most haloanisole contamination is anthropogenic
- TCA is the most prevalent haloanisole contaminant but TBA is also becoming an issue in wines
- With a sensitive extraction procedure, haloanisoles can be detected well below their odor threshold