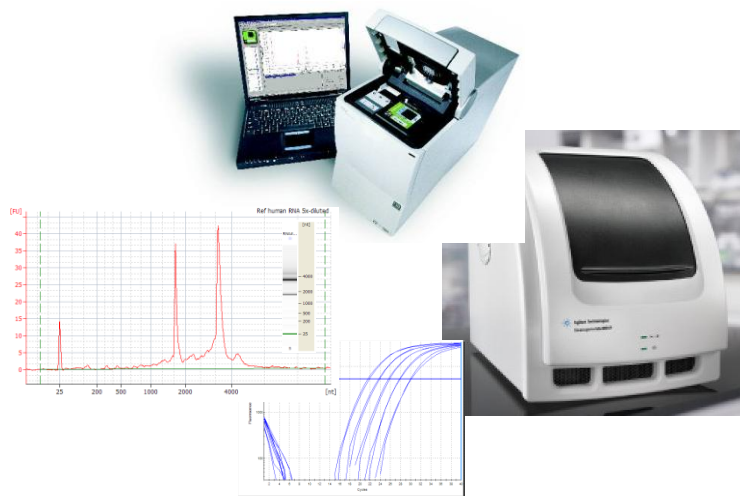


# Overview Agilent Micro fluidics



**Cathy Cutler**  
**Field Application Scientist**  
**Agilent-Genomics**

# The Agilent 2100 Bioanalyzer

**March 2010: >15,000 citations !**

First commercially available Lab-on-a-Chip product

Introduced 1999

Analysis of biomolecules:

DNA, RNA, Proteins and cells

More than 7000 of instruments sold  
WW

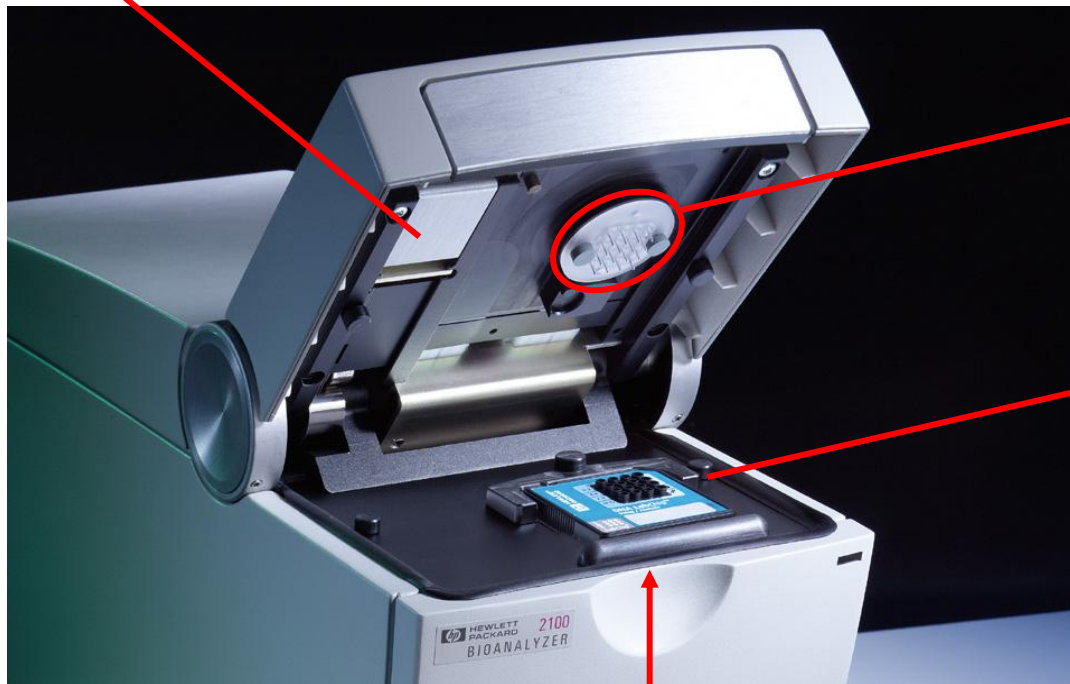
Industry-Standard for the analysis of  
RNA

Standard for sample QC in Next-  
Generation Sequencing workflows



# Agilent 2100 Bioanalyzer

**Exchangeable  
cartridge for different  
assays**



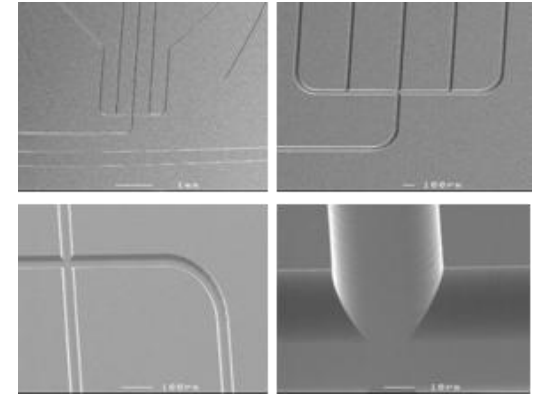
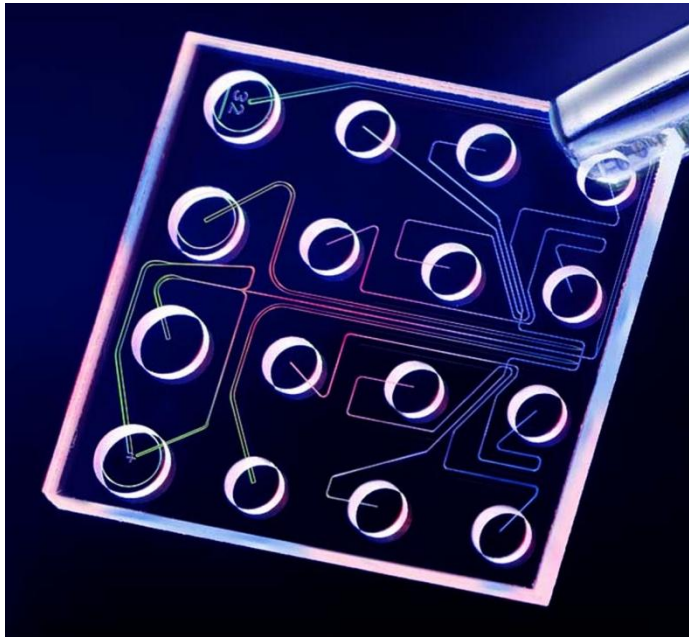
**16 pin electrodes  
connected to  
HV-sources**

**Chip holder  
with heater  
plate**

**Optics for detection**

# The Lab-on-a-Chip Approach

## Increasing quality and speed of gel electrophoresis



Sample volumes 1 - 5  $\mu$ l

10 -12 samples depending on Assay

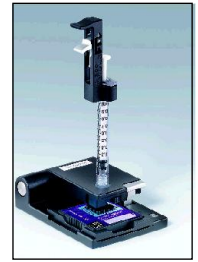
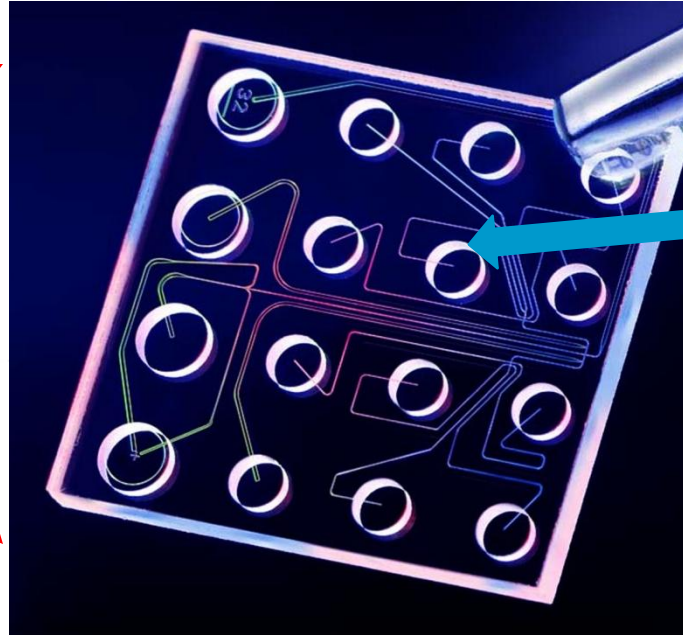
Separation, staining, detection of samples

Results in 5-30 minutes available

No extra waste removal needed

Disposable Chip, no crosscontamination

# Chip Architecture



Gel loading device

Conditioning channel and point of detection

- Chip accommodates **sample** wells, **gel** and **conditioning/destaining** wells, and a well for a **standard (ladder)**
- Sieving gel/dye matrix is forced into capillaries
- 16 pin electrodes apply voltage
- Capillary fluidics become “lanes”



# Principle of Electrodriven Flow





# Lab-on-a-Chip - General Features and Benefits

## Miniaturization (Scale)

- small sample volumes
- reduced reagent usage
- reduced bench space

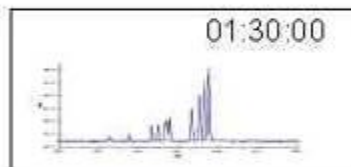


Handling

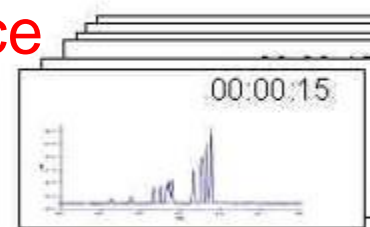


## Miniaturization (Speed)

- fast analysis



Surface



## Automation

- improved accuracy
- improved precision
- improved productivity

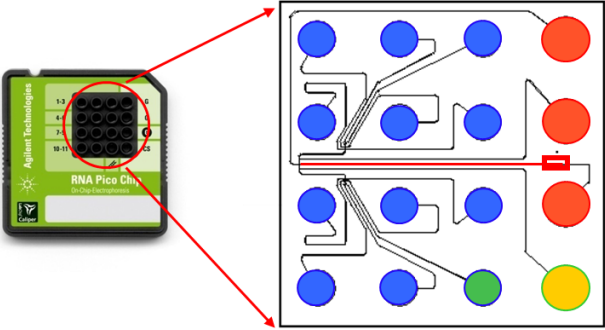
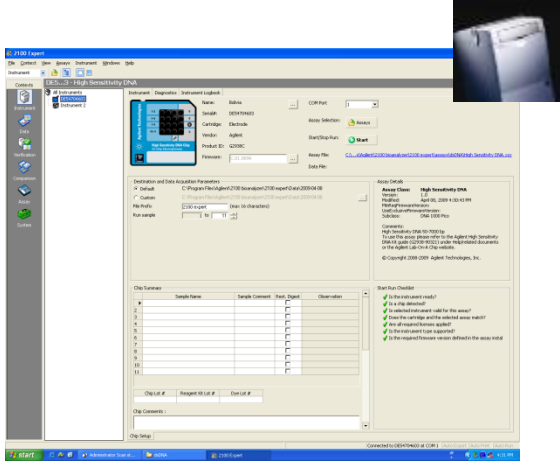
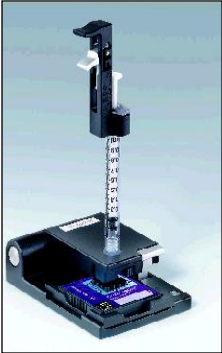


Software



*Smaller - Faster - Smarter*

# The Kits





# 2100 Expert Software

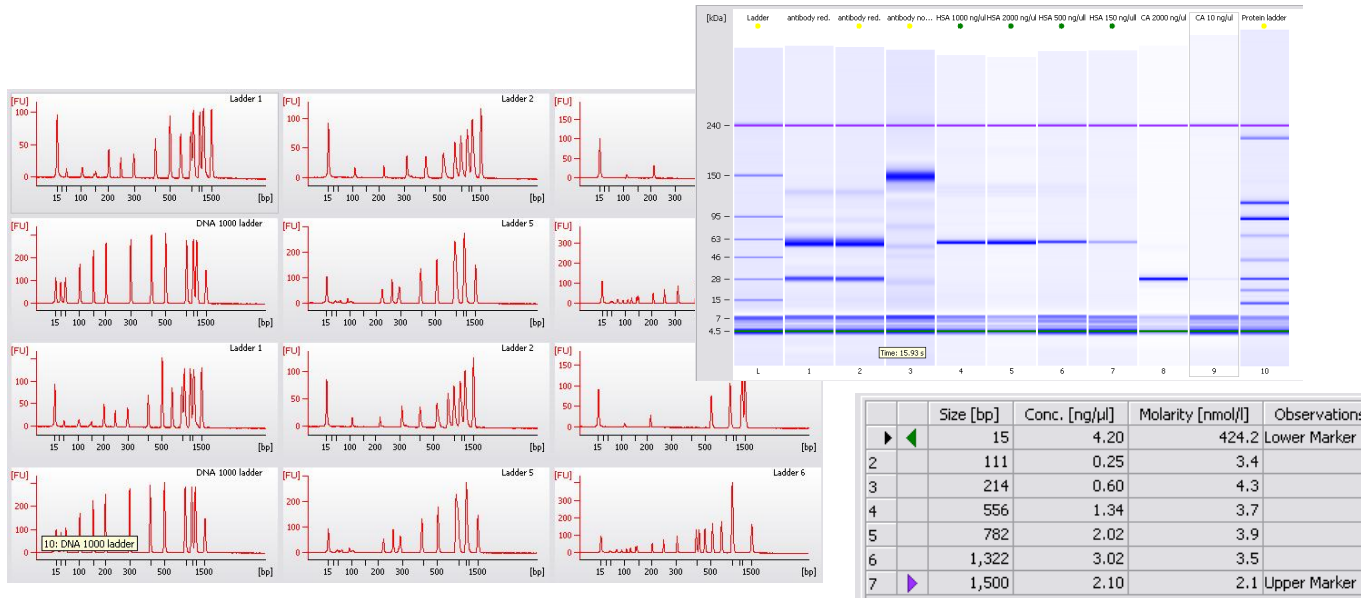
## Version B.02.08



- Easy to use for Instrument control, Data Analysis, Data comparison and Reporting
- Patented RIN (RNA Integrity Number)
- Color coded Result Flagging
- Easy comparison context (multi samples from various files)
- Customizable result tables for printing and reports
- Optional security pack software for 21 CFR part 11 compliance requirements

### NEW with B.02.08:

- High Sensitivity DNA Assay
- Plant RIN Application
- Windows 7 Compatibility



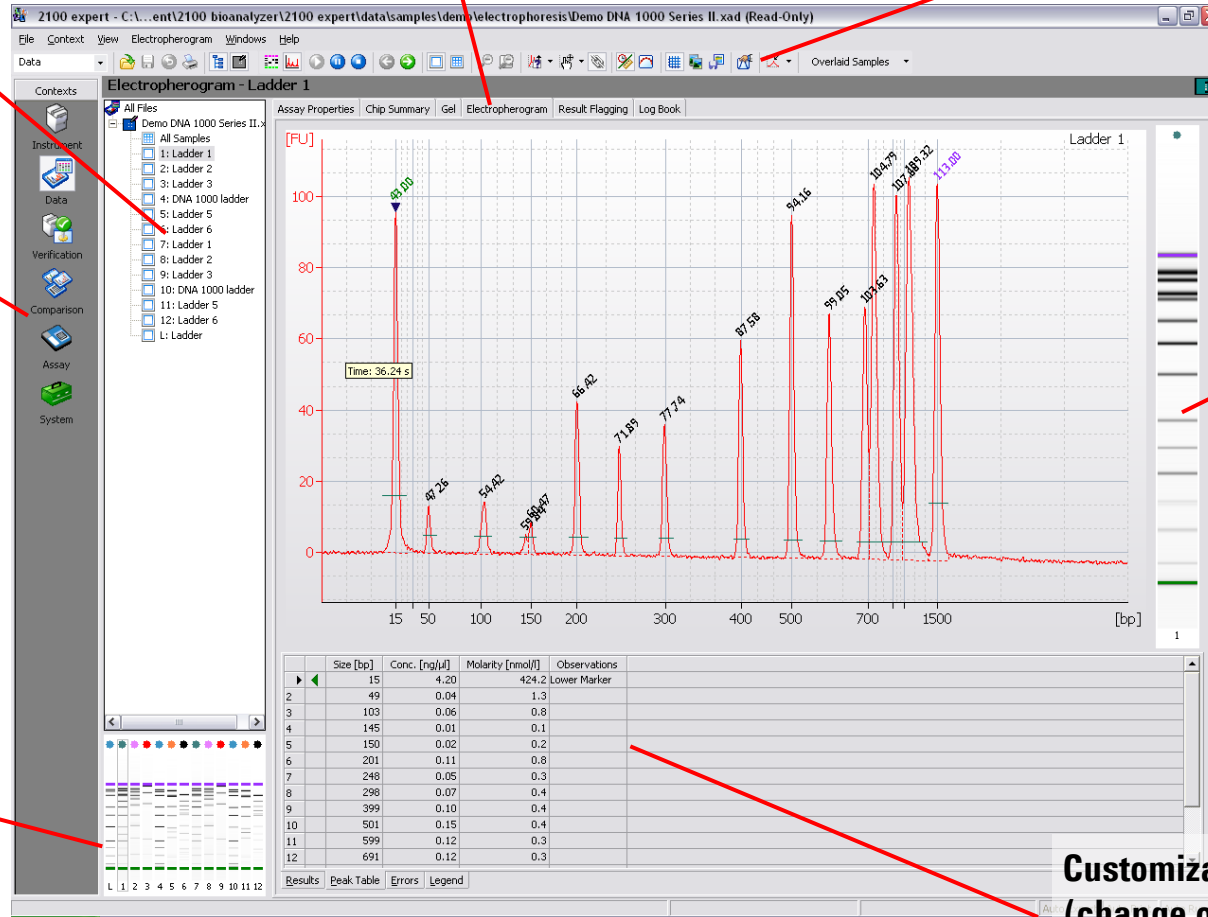
# Analyze Data (Peak based view)

Tree view for navigation between samples and files

Tabs for different data analysis functions

Task bar with context sensitive icons for different actions

Context menu bar

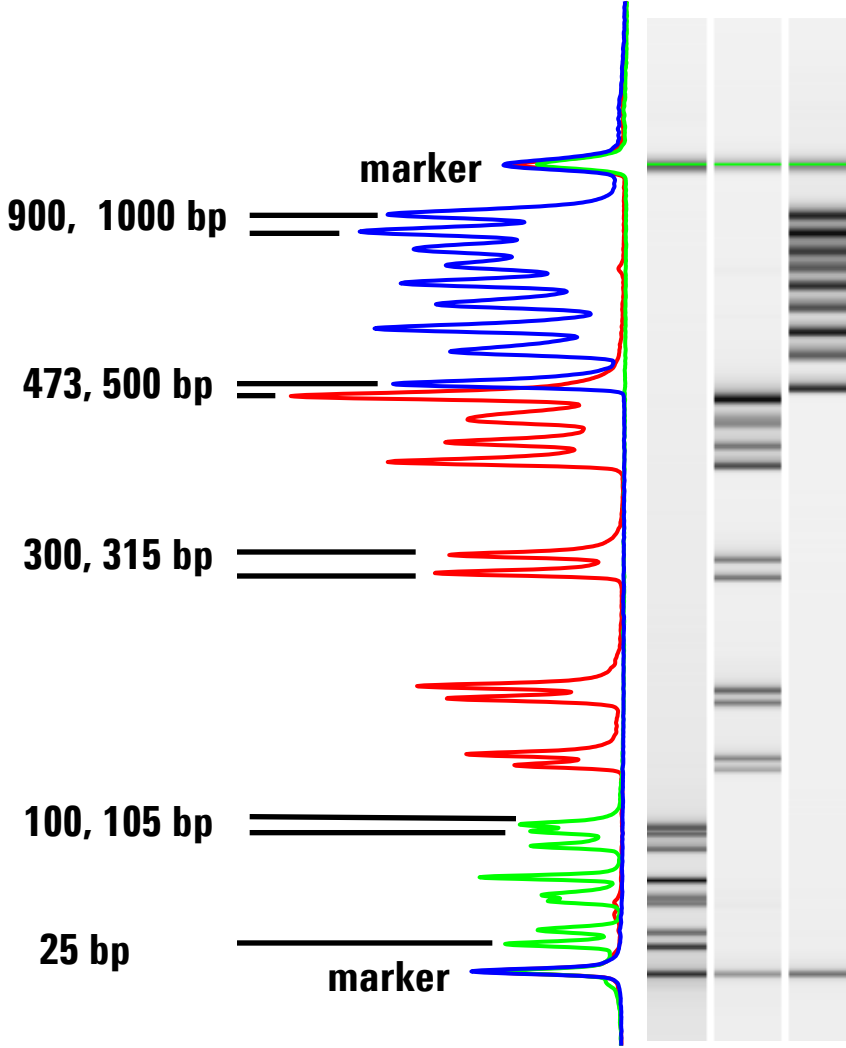


Single gel lane for selected E-gram

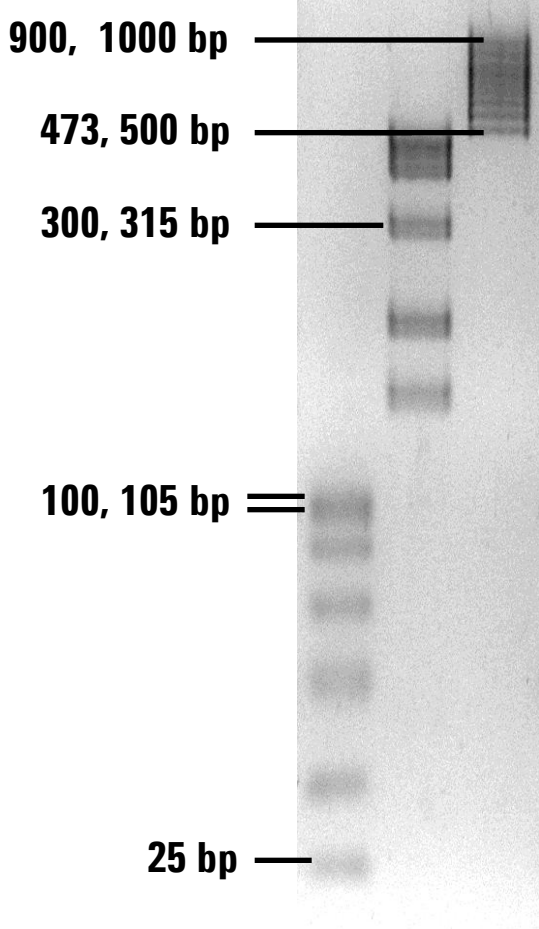
Customizable gel-like image (change order)

Customizable result table (change order and add additional columns)

# Data Format - Gel-Like Image c/w Agarose Gel



2100 bioanalyzer data  
Gel-like image



2 % agarose gel stained  
with Ethidiumbromide

# Current Assays (including anything required for analysis)



DNA1000

DNA7500

DNA12000



High Sensitivity  
DNA

## DNA Assays:

- Sizing
- Quantitation
- PCR products, digests, larger DNA fragments
- 12 samples in 30 min.



RNA Nano

RNA Pico

RNA Small

## RNA Assays:

- Quantitation (Sizing in Small RNA)
- total RNA, mRNA
- purity & integrity determination
- 10 samples in 30 min.



Flow Cytometry

## Cell Assays:

- Analysis of 6 samples
- Two color detection
- Analysis of protein expression in cells



P230

P80

HSP 250

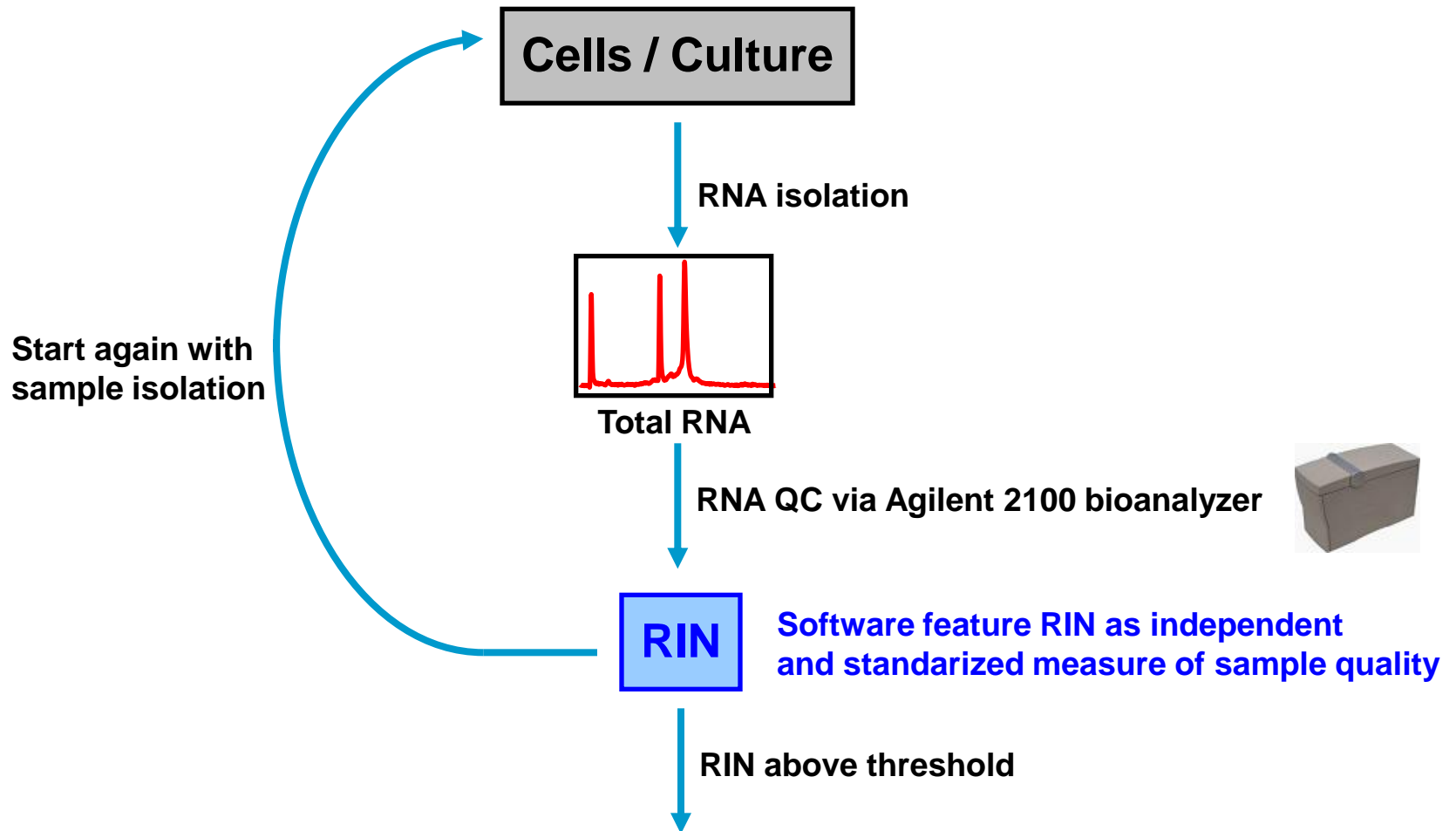
## Protein Assays:

- Sizing
- Quantitation
- cell lysates, column fractions, purified proteins, antibodies etc.
- 10 samples in 40 min.



# RNA and Small RNA Assays

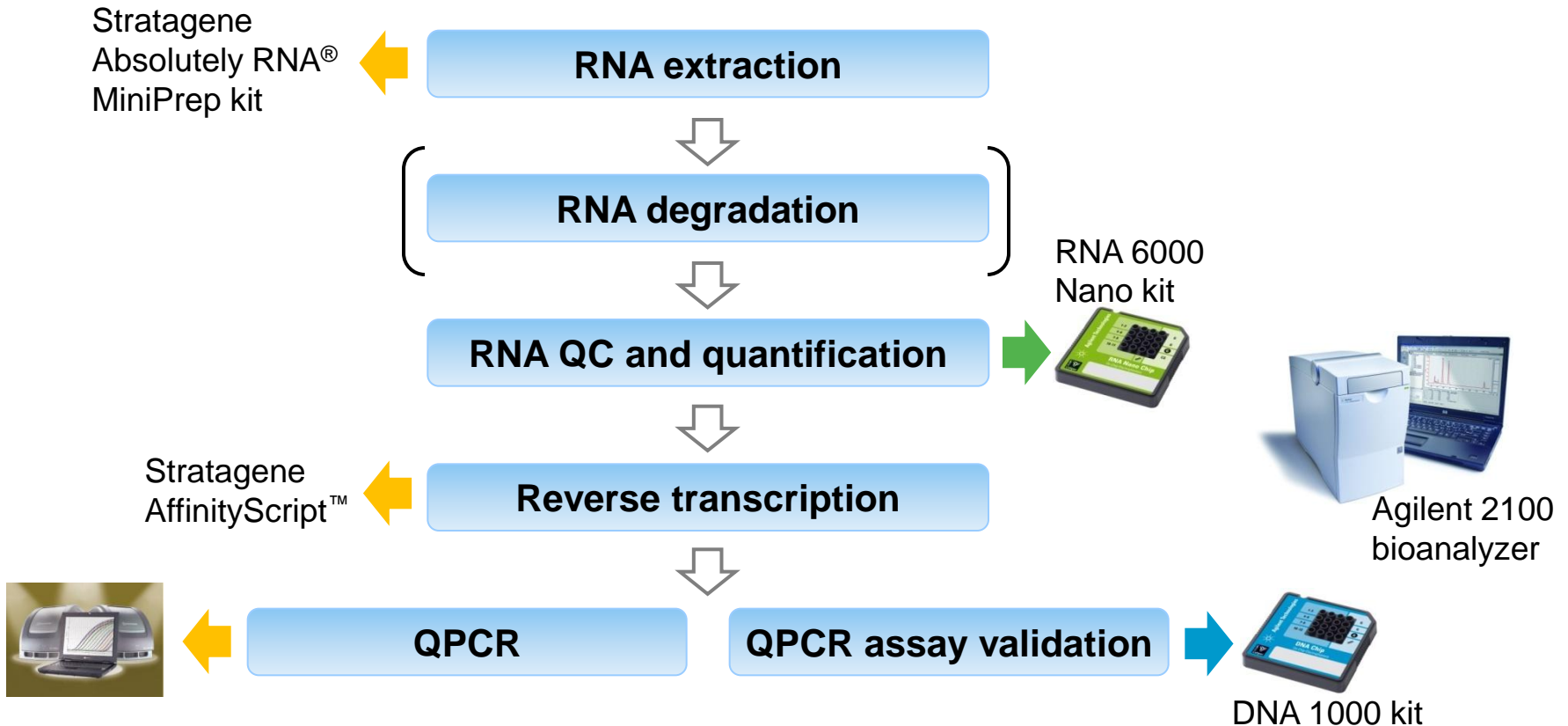
# RNA QC in Routine Gene Expression Workflow



**Continue with downstream Experiment (Microarray, real-time PCR, etc.)**



# Experimental workflow



# Successful QPCR with the Agilent 2100 bioanalyzer

## RNA quality - Effects of degraded RNA

Not knowing the extend of possible degradation might lead to false negative results or misinterpretation of the data if the amplicon falls into a degraded region

Knowing RNA quality allows to accommodate the design and set expectations avoiding wrong interpretation of results

## Assay validation - Limitations of SYBR Green melt curves

Restricted resolution, which can make it difficult to determine specificity

$T_m$  depends on dye/template ratio

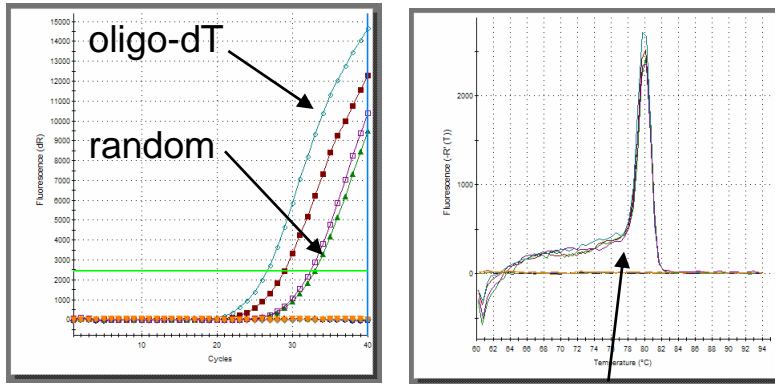
SYBR Green is a non-saturating dye with non-uniform distribution along the double-strand

Melt curve provides no info on the size of the generated amplicon

# QPCR assay validation

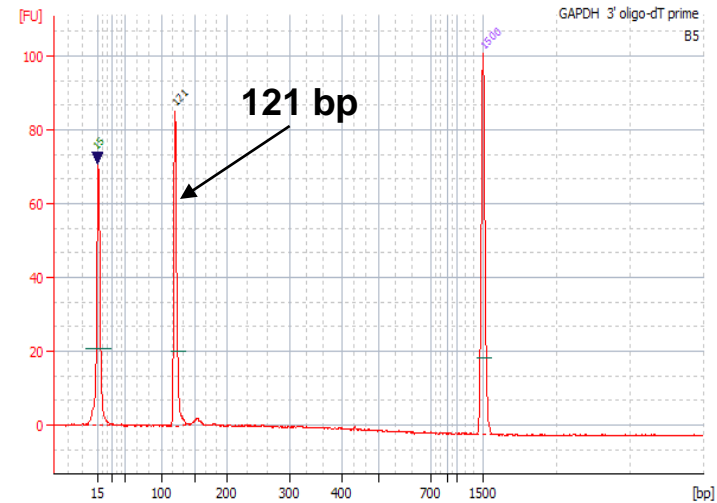
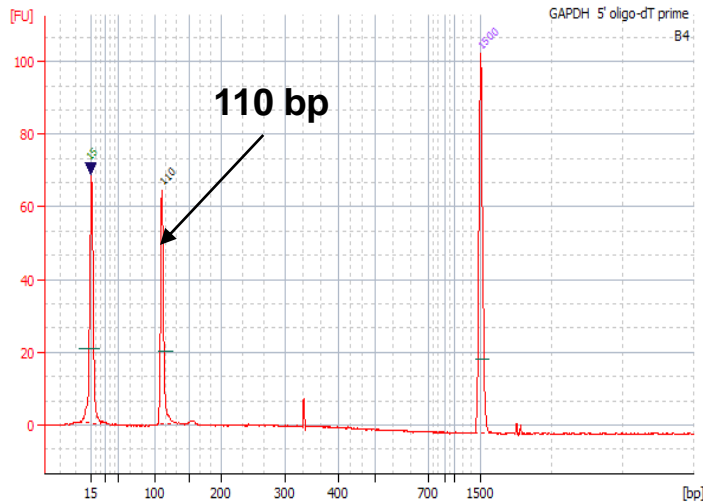
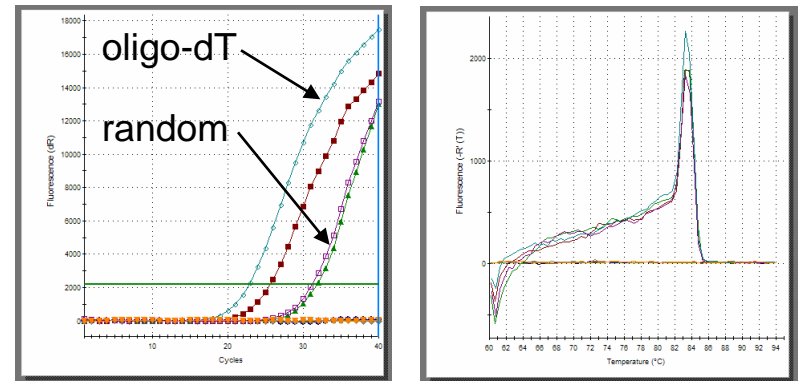
– amplification plots, melt curve and bioanalyzer analysis

**GAPDH 5' assay: Expected size 118 bp**



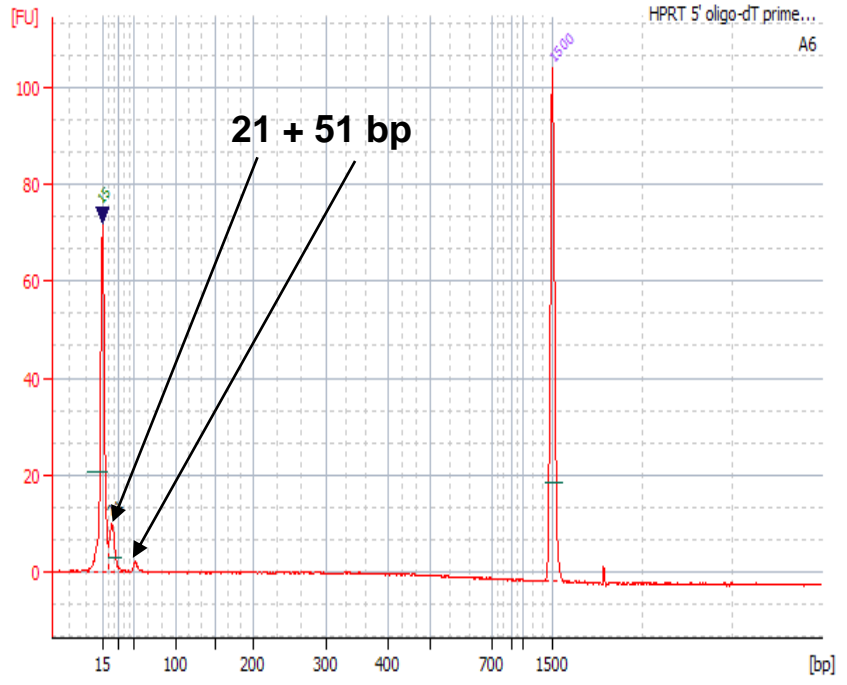
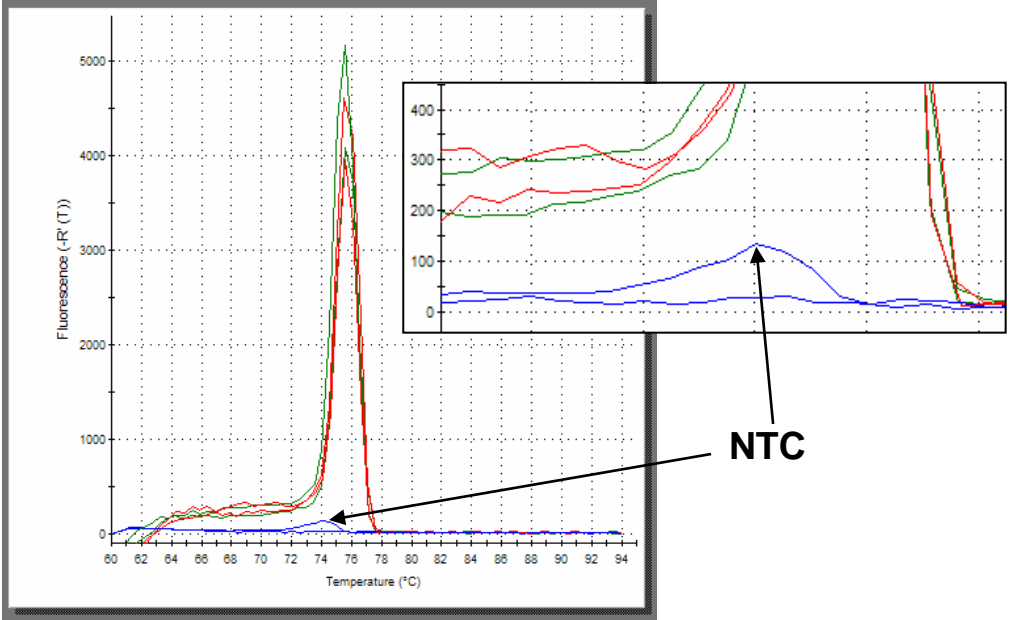
**NoRT**

**GAPDH 3' assay: Expected size 126 bp**



# QPCR assay validation

- No-template controls (NTC)



# Features of the RNA 6000 Assays

total RNA

**determine integrity and quality of total RNA**

**determination of RNA concentration**

**identify ribosomal peaks**

**calculate the ratio of ribosomal peaks (18S/28S or 16S/23S)**

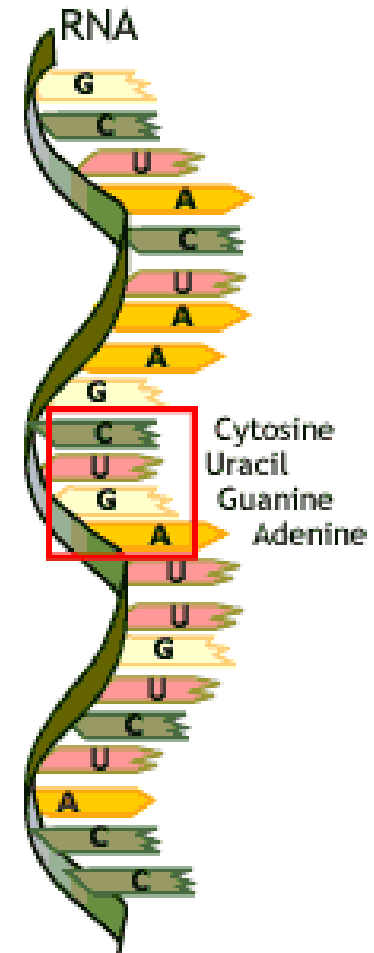
**RNA integrity number (RIN)**

mRNA

**determine integrity and quality of mRNA samples**

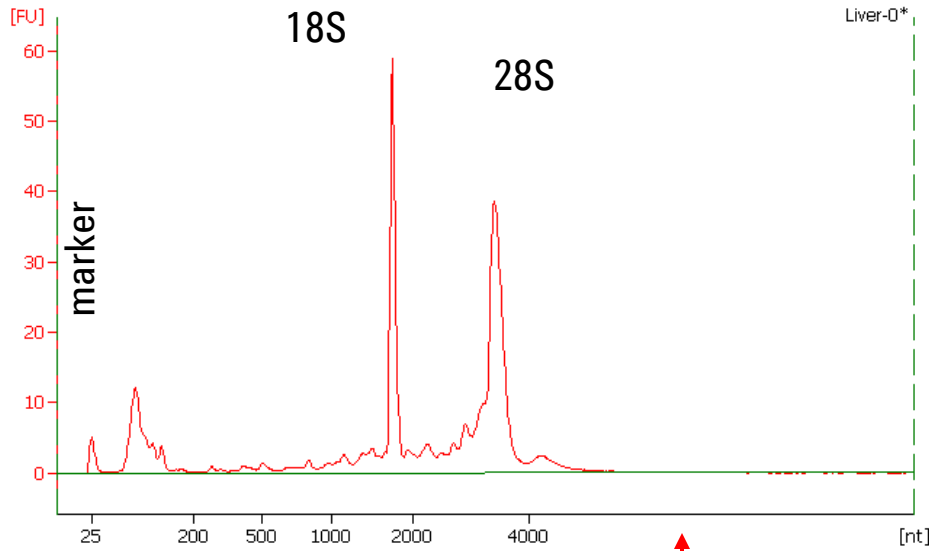
**Determination of mRNA concentration**

**calculate % ribosomal RNA in mRNA samples**



# Assesment of RNA Integrity

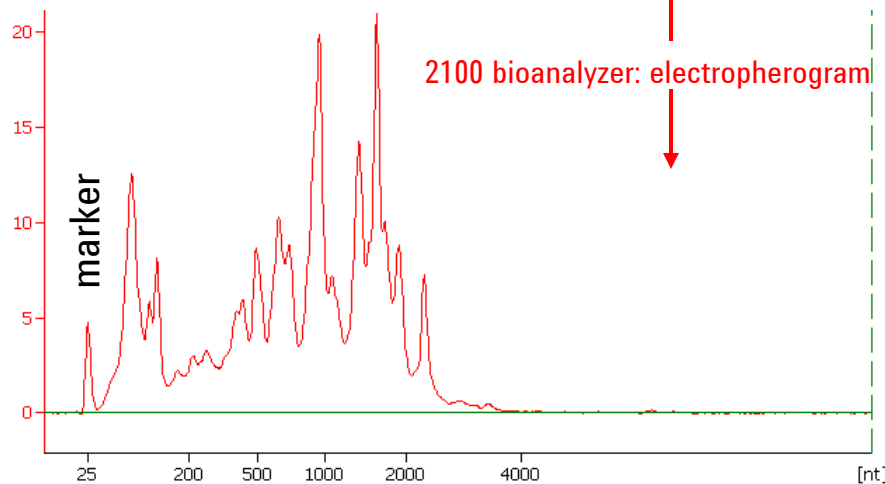
## RNA 6000 Nano LabChip kit



Typical first QC step during cDNA or cRNA sample prep for QPCR

High quality total RNA (RIN 8.8)

1



Partially degraded total RNA (RIN 3.7)

9





# DNA Applications

# Application Areas for the DNA Assays

**PCR product purity**

**Multiplex PCR Applications**

**Gene expression analysis via RT-PCR (target validation)**

**GMO testing**

**Pathogen detection (homeland defense, hospitals, environmental)**

**Genotyping applications**

- Duplications/ deletions
- Allele frequency
- Bacterial sub-typing
- Forensics

**Cancer diagnostics**



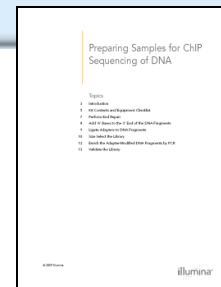
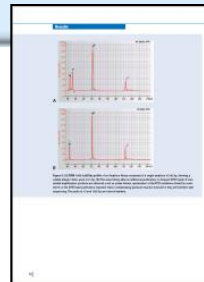
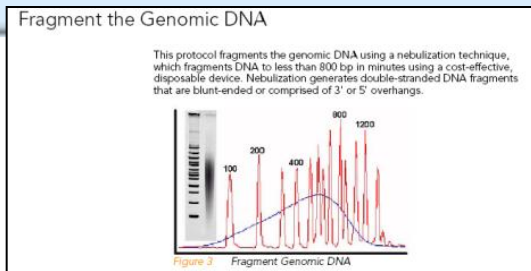
# DNA QC in Next-Gen Sequencing - Situation



- Next Generation Sequencing is an emerging technology.
- Quality of sequences is one of the chief technical concerns about next gen sequencing platforms in a recent customer survey by Insight pharma reports
- Leading next generation sequencing vendors (*Illumina, Roche, Life Technologies*) recommend existing Bioanalyzer DNA assays in their workflows

## References

- Manuals of all Next Gen Sequencing Vendors: Illumina, Roche, Life Technologies



Validate the Library

The amount of starting material is very low (10 ng), and after 18 cycles of PCR, the yield could still be too low to see on a regular gel, even though it is enough for cluster generation. Illumina recommends performing the following, more sensitive quality control analysis on your sample library using an Agilent Technologies 2100 Bioanalyzer.

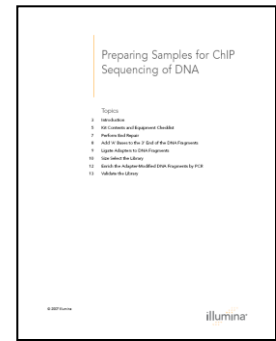
**Bioanalyzer Method**

1. Load 1  $\mu$ l of the resuspended construct and 1  $\mu$ l of the negative control on an Agilent Technologies 2100 Bioanalyzer.
2. Check the size, purity, and concentration of the sample.

**Alternative Methods**

For users who do not have access to an Agilent Technologies 2100 Bioanalyzer or similar instrument, you may try using a sensitive dsDNA measurement assay such as the Quant-iT dsDNA HS Assay Kit, 100 assays Q52107 for use with the Quin Fluorometer (Invitrogen). Note that this will not allow you to check the size and purity of your sample. Do not use an OD<sub>260</sub>/OD<sub>280</sub> ratio for concentration measurements, since this will not distinguish dsDNA from primers, and therefore cannot be used to validate the library.

# Bioanalyzer in Illumina/Solexa Workflow – Examples



Workflow

## ChIP-Seq

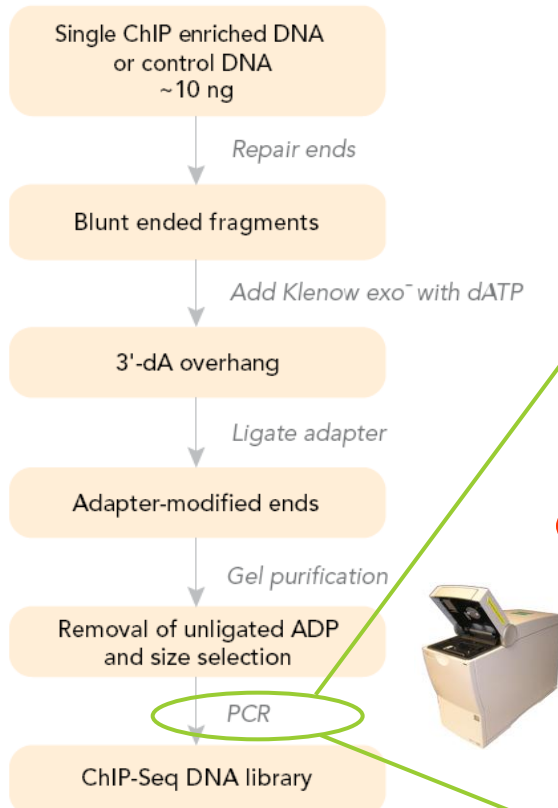


Figure 2 Sample Preparation Workflow

### Validate the Library

The amount of starting material is very low (10 ng), and after 18 cycles of pcr, the yield could still be too low to see on a regular gel, even though it is enough for cluster generation. Illumina recommends performing the following, more sensitive quality control analysis on your sample library using an Agilent Technologies 2100 Bioanalyzer.

### Bioanalyzer Method

1. Load 1  $\mu$ l of the resuspended construct and 1  $\mu$ l of the negative control on an Agilent Technologies 2100 Bioanalyzer.
2. Check the size, purity, and concentration of the sample.

### Alternative Methods

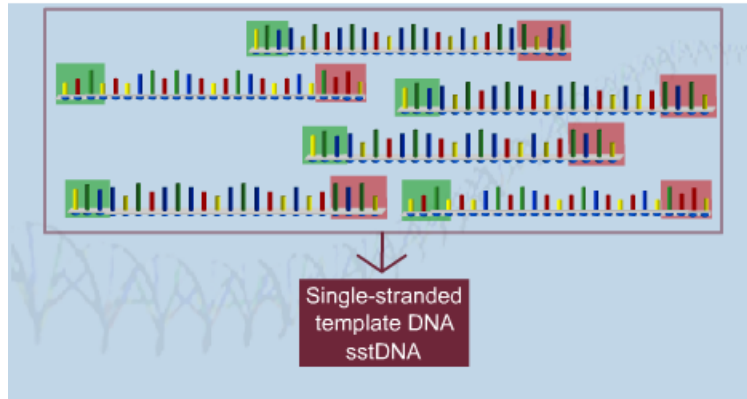
For users who do not have access to an Agilent Technologies 2100 Bioanalyzer or similar instrument, you may try using a sensitive dsDNA measurement assay such as the Quant-iT dsDNA HS Assay Kit, 100 assays 0.2-100 ng for use with the Qubit fluorometer (Invitrogen). Note that this will not allow you to check the size and purity of your sample. Do not use an OD260/280 ratio for concentration measurements, since this will not distinguish dsDNA from primers, and therefore cannot be used to validate the library.

# Bioanalyzer in Roche/454 GS FLX Workflow



**DNA Kit Family** Validate DNA fragmentation (< 800bp) with **DNA kit family**

DNA Library Preparation

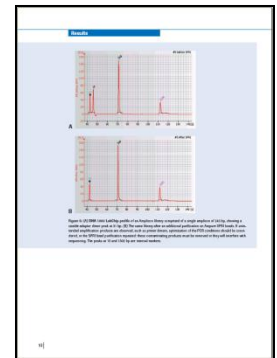


**RNA Kit Family**

Validate ssDNA library

RNA kit:

- estimate the size distribution
- to determine if fragments <300 bp successfully removed
- quantitation for correct dilution



454 recommends the use of Agilent Bioanalyzer in their manuals

[http://www.454.com/downloads/protocols/Guide\\_To\\_Amplicon\\_Sequencing.pdf](http://www.454.com/downloads/protocols/Guide_To_Amplicon_Sequencing.pdf)

<http://cage.unl.edu/3.%20GS%20FLX%20Shotgun%20DNA%20Library%20Preparation%20QuickGuide.pdf>

# Bioanalyzer in ABI Solid Small RNA Sequencing Workflow



Small RNA Kit

Check small RNA content in total RNA samples

If small RNA content >0.5% use total RNA

If small RNA content <0.5% enrich small RNA first

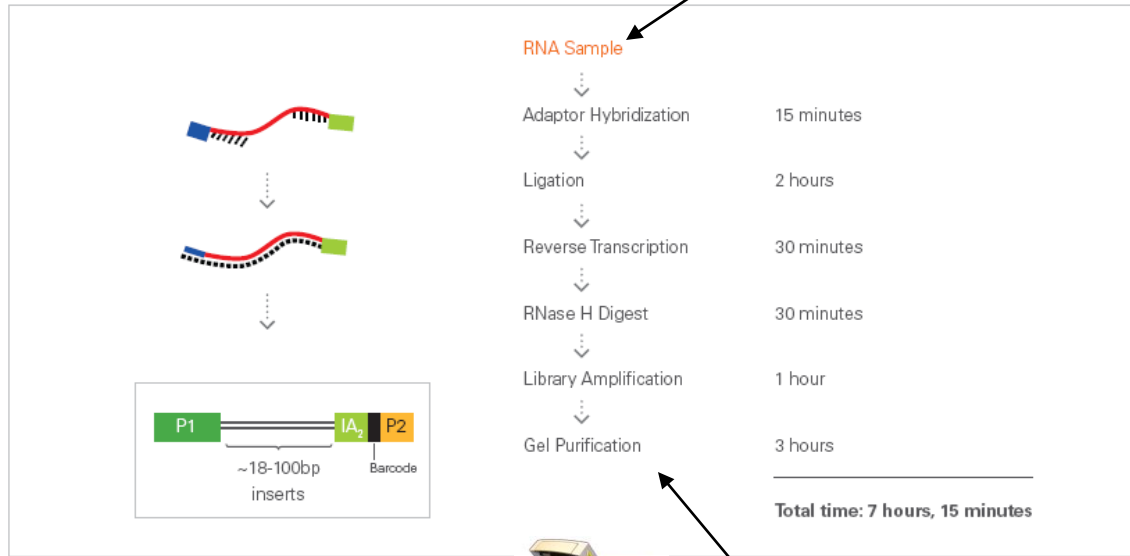


Figure 1: Workflow for SOLiD™ Small RNA Library Preparation

SOLiD™ Small RNA Expression Kit	
Part Number: 5067-1548	
Product	
I. Introduction	1
A. Subsequent and Product Overview	
B. Product Overview	
C. Reagent Overview and Use and Storage Conditions	
D. Materials Provided with the Kit	
E. Related Documents Available from Agilent Technologies	
II. Protocol	7
A. RNA Sample Type and Source	
B. Minimums Required	
C. Sample Preparation and Ethanol Digestion	
D. Small RNA Library Amplification	
E. Amplified Small RNA Library Cleanup	
F. The Addition of Amplified Small RNA Library to P1/E2	
III. Troubleshooting	10
A. System Configuration	
B. Troubleshooting the Agilent Bioanalyzer	
IV. Appendix	20
A. Sequence of the SOLiD™ P1/E2 Primers Included in the Kit	
B. Minimums of Gel Extraction	
C. References	
D. Quality Control	
E. Other Information	



DNA Kit Family

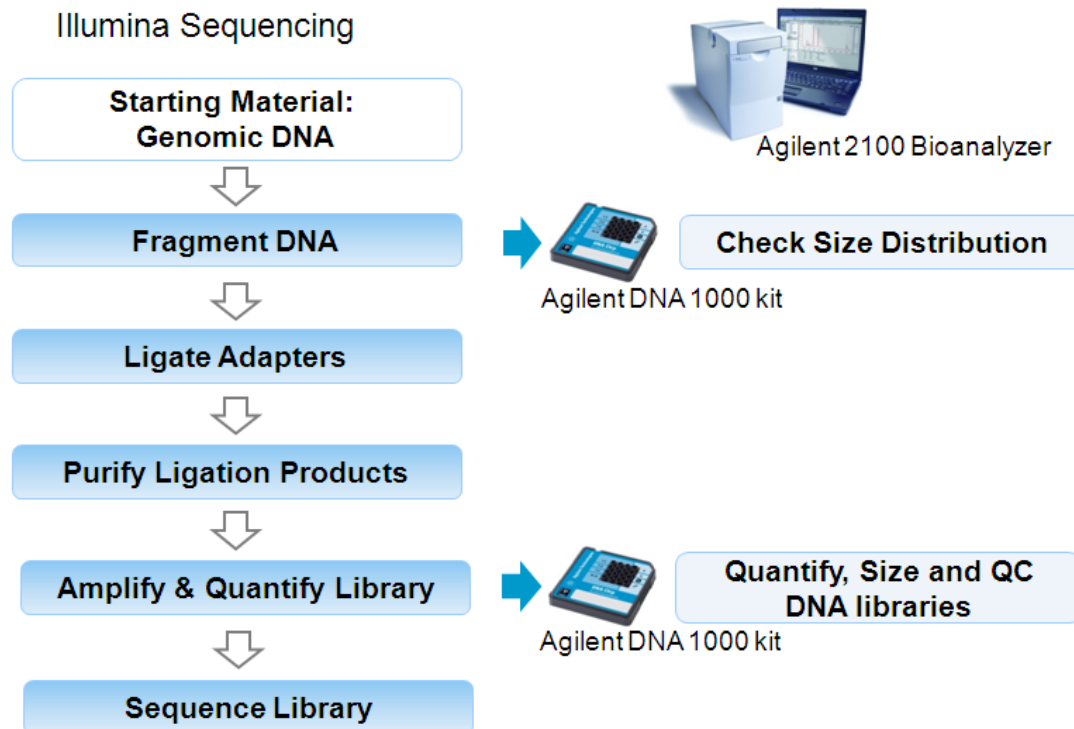
verify the size and quality using an Agilent bioanalyzer or 6% native PAGE.

“.....we recommend evaluating the small RNA content of samples to determine whether to use total RNA or size-selected RNA in your reactions. This can be done using an Agilent bioanalyzer with the Small RNA Chip (#5067-1548).”



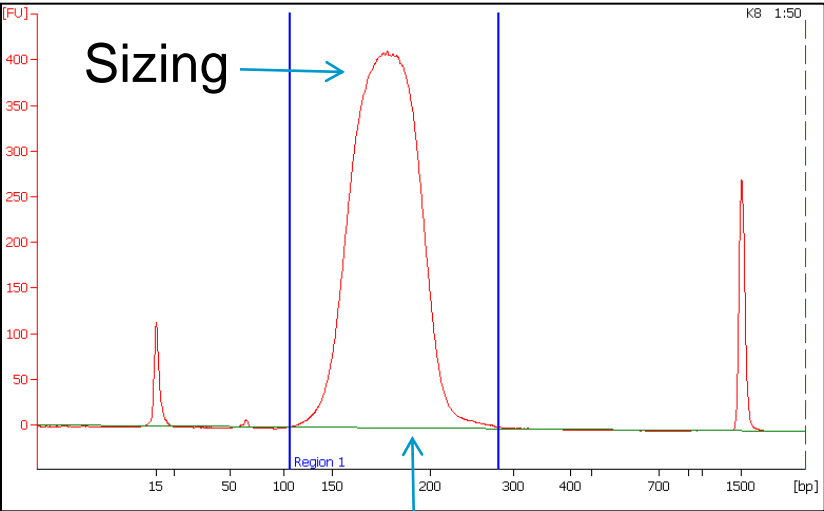
# Sneak Preview – High Sensitivity DNA Kit

## Simplified Illumina GAII Workflow



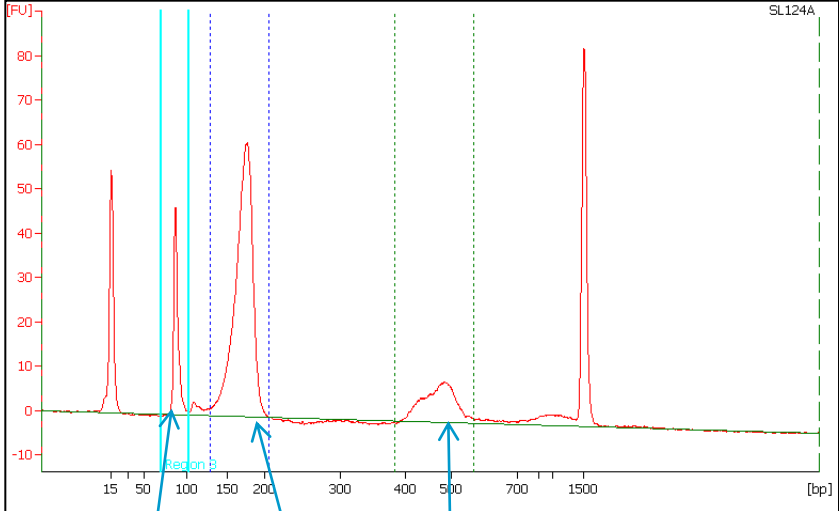
# Quantification, Sizing and QC of NGS Libraries

### High Quality DNA library



Quantification

### Identification and quantification of primer dimers and PCR artifacts

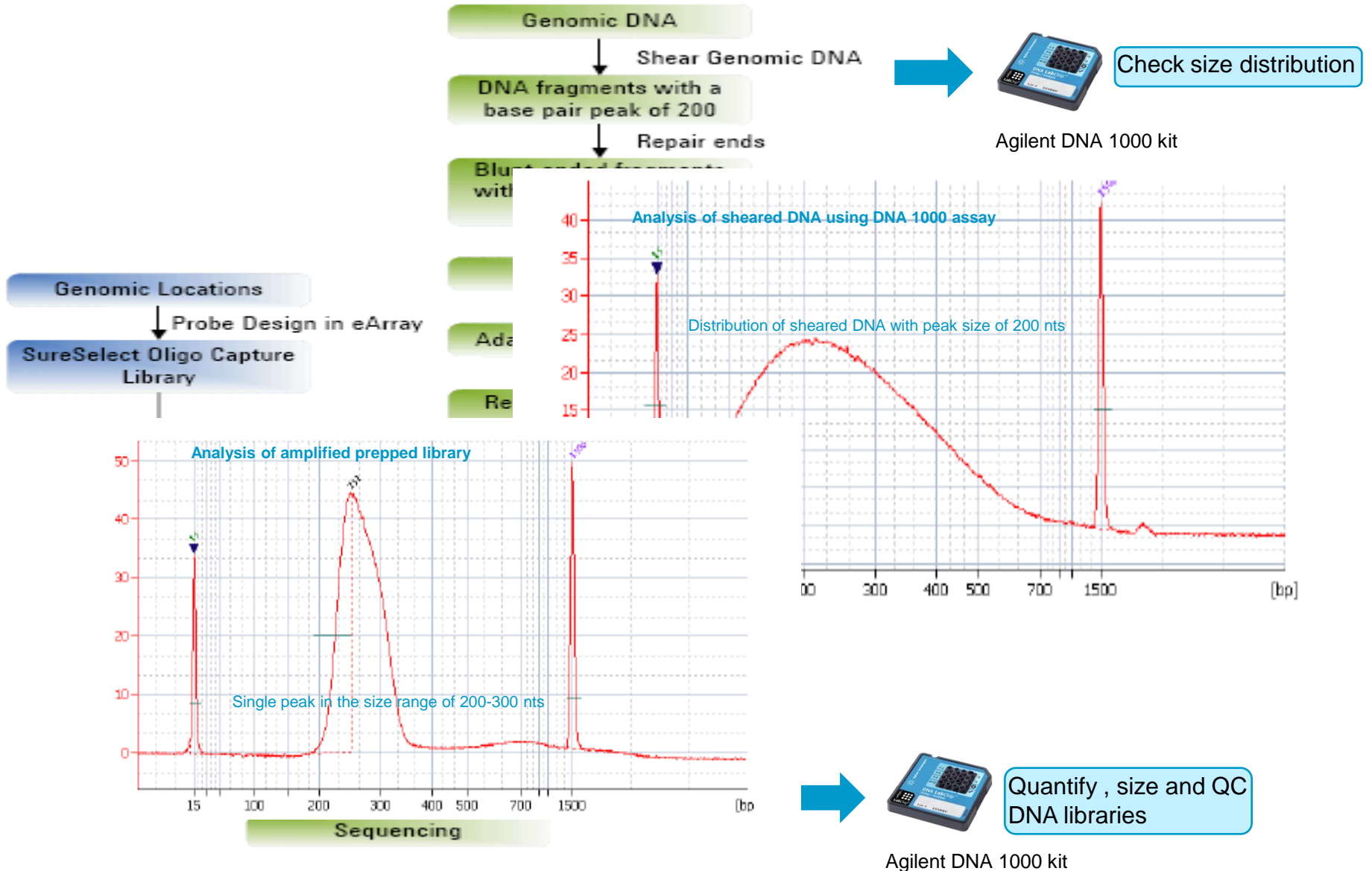


Primer Dimers/  
Adaptors

PCR artifact

DNA library

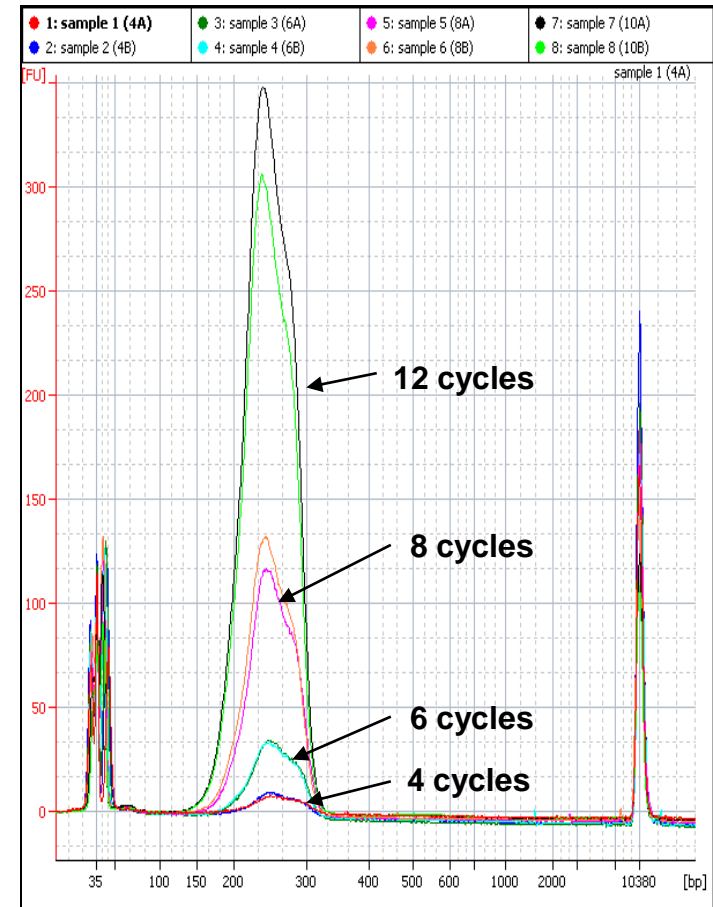
# SureSelect Overall Sequencing Sample Prep Workflow



# Improving the next-gen sequencing Sample Prep Workflow

**Sizing and quantification of a Illumina GAT library. The library was enriched with Agilent's SureSelect Target Enrichment platform and amplified with varying PCR cycles.**

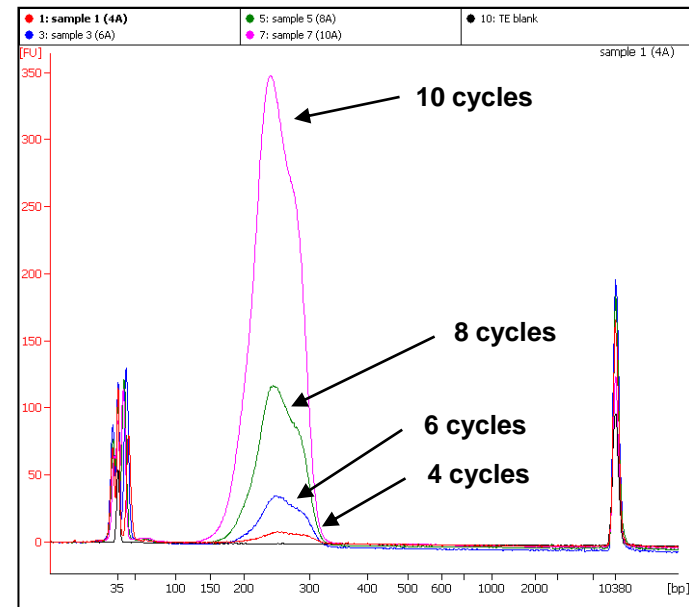
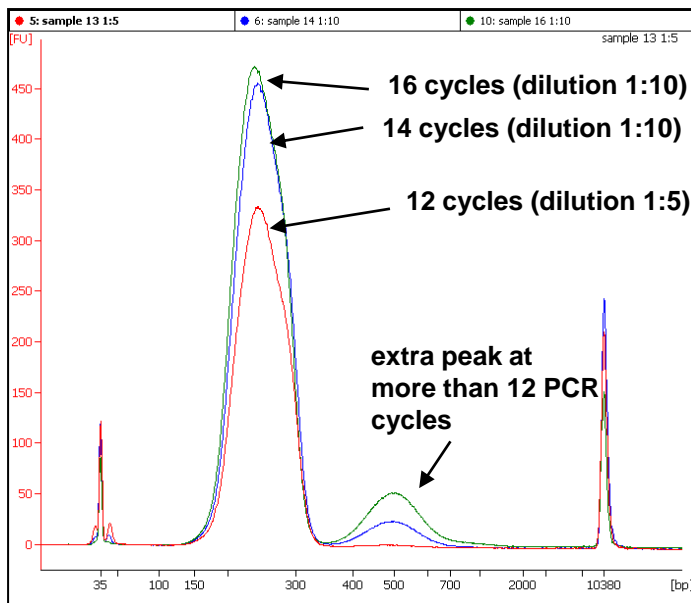
Sample ID	No. Cycles	Qubit BR ng/ul	Bioanalyzer ng/μl
4A	4	Too Low	0.090 0.015
4B	4	Too Low	0.083 0.007
6A	6	Too Low	0.304 0.017
6B	6	Too Low	0.301 0.010
8A	8	Too Low	1.13 0.07
8B	8	Too Low	1.34 0.07
10A	10	3.4	4.49 0.16
10B	10	3.4	4.54 0.17
A12	12	13.4	14.5 0.5
C12	12	12.5	13.9 0.6



# High Sensitivity DNA kit Applications – Reduction of Library Amplification Cycles

*Sizing and quantification of a Illumina GAll library. The library was enriched with Agilent's SureSelect Target Enrichment platform and amplified with varying PCR cycles.*

- Amplification cycles above a certain threshold can result in amplification artifacts.
- High Sensitivity DNA kit enables to reduce library amplification cycles



# Protein Kit Portfolio



Silver Stain Sensitivity



## P80

Range: 5 - 80 kDa  
Sensitivity: Coomassie  
Samples: 10

Samples  
-Antibodies (reduced)  
-Small Proteins



## P230

Range: 14 - 230 kDa  
Sensitivity: Coomassie  
Samples: 10

Samples  
-Antibodies (all types)  
-Standard Proteins

Coomassie Range (5 ng/μl BSA)



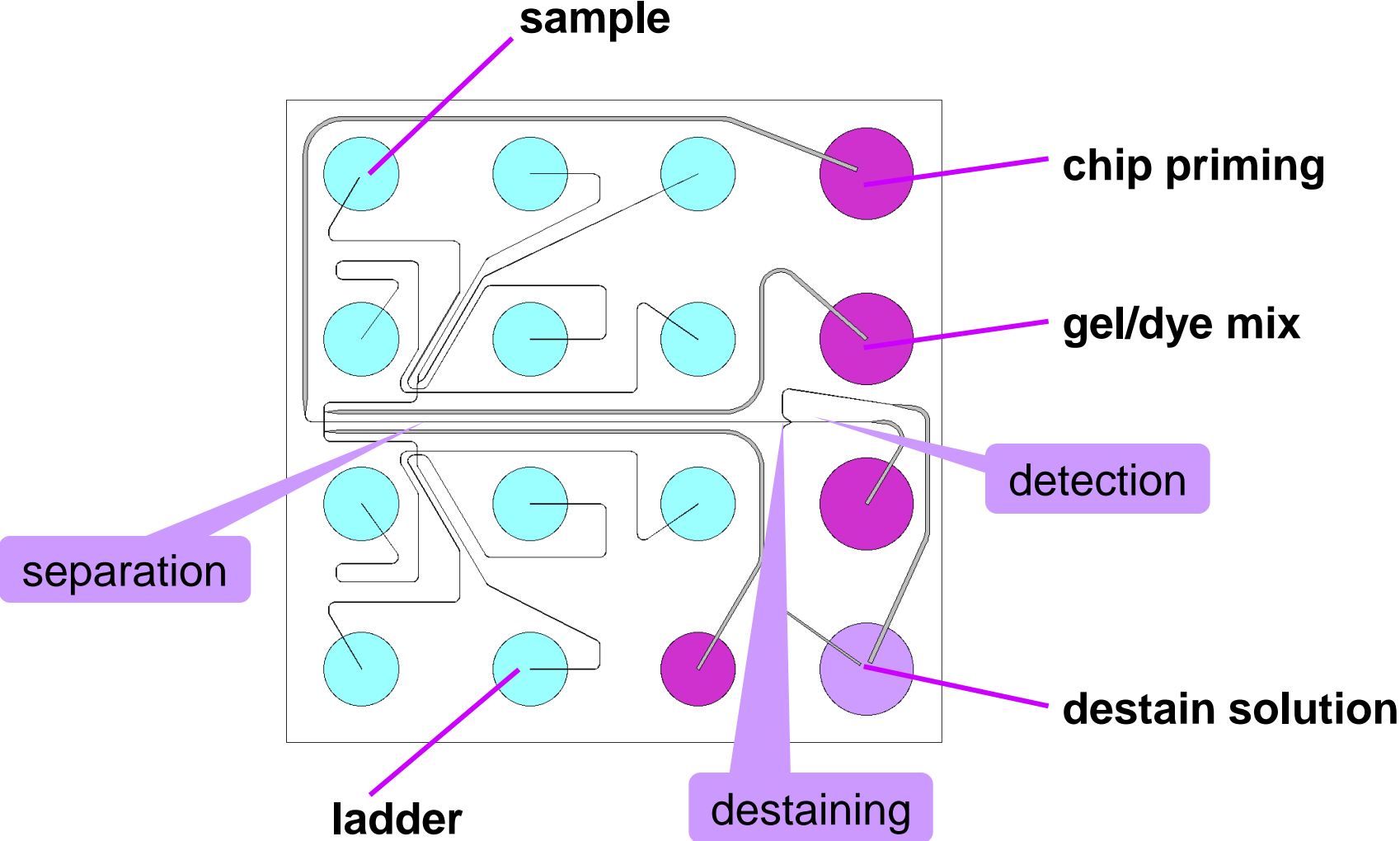
## HSP-250

Range: 10 - 250 kDa  
Sensitivity: 1 pg/μl BSA  
Samples #: 10 per Chip  
Chips #: 10 per Kit  
Labeling Conc: 1 ng – 1 μg /μl

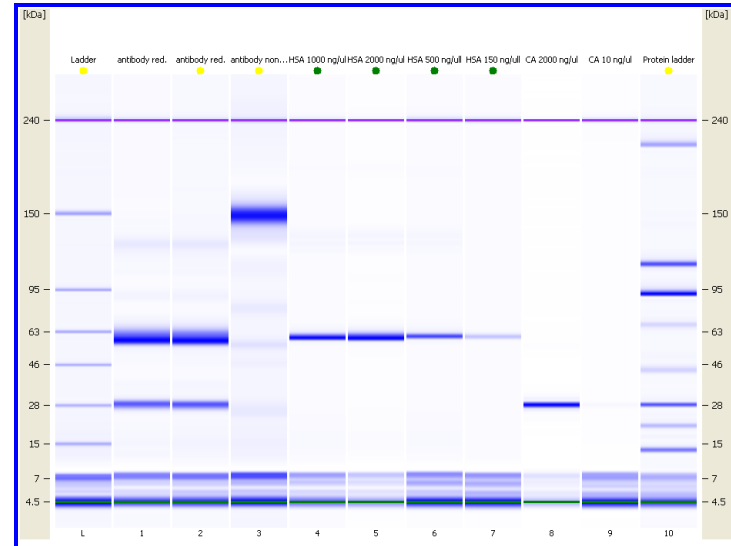
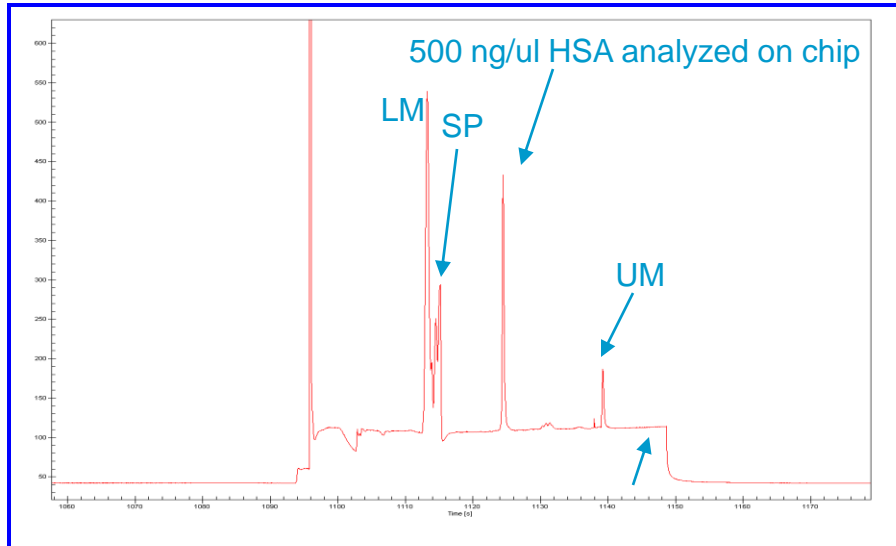
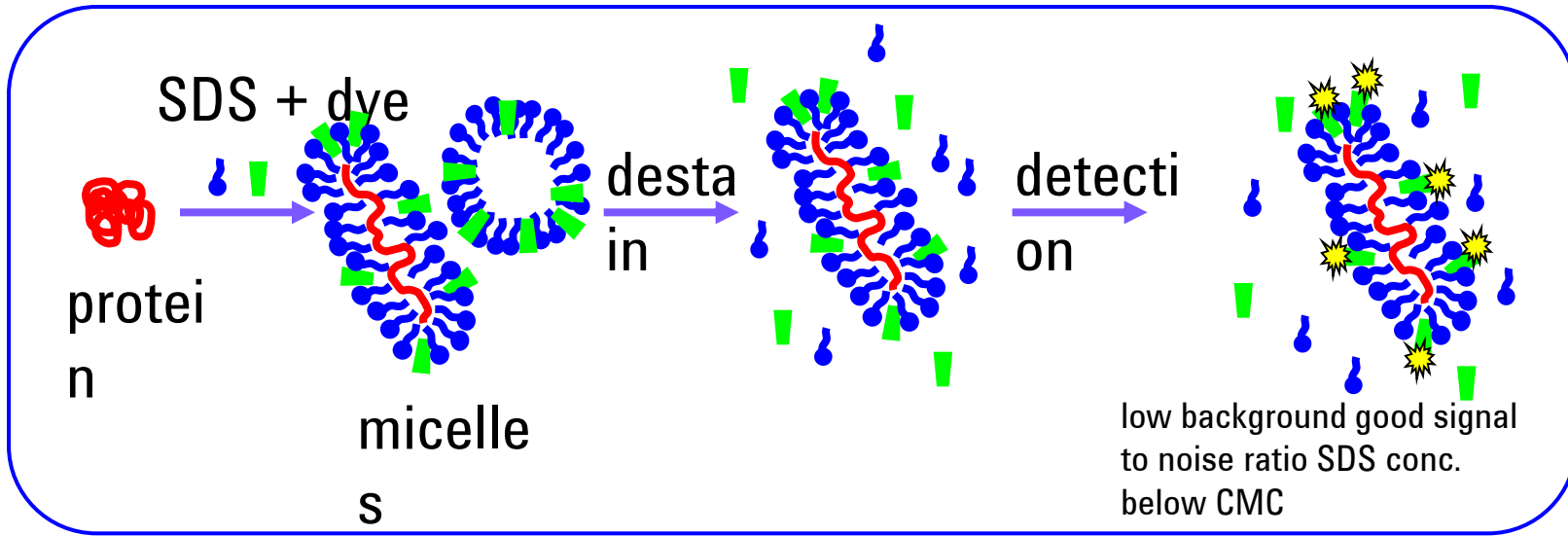
Requirements:  
Software: Expert B.02.06 (+)  
Instrument: all except G2938A

Silver stain Range (200 pg/μL BSA)

# Protein Chip Layout

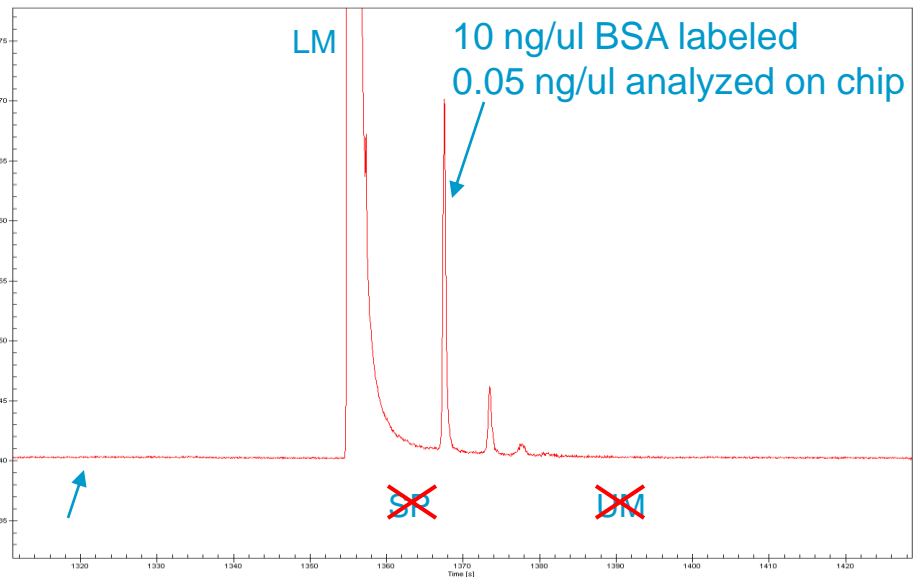
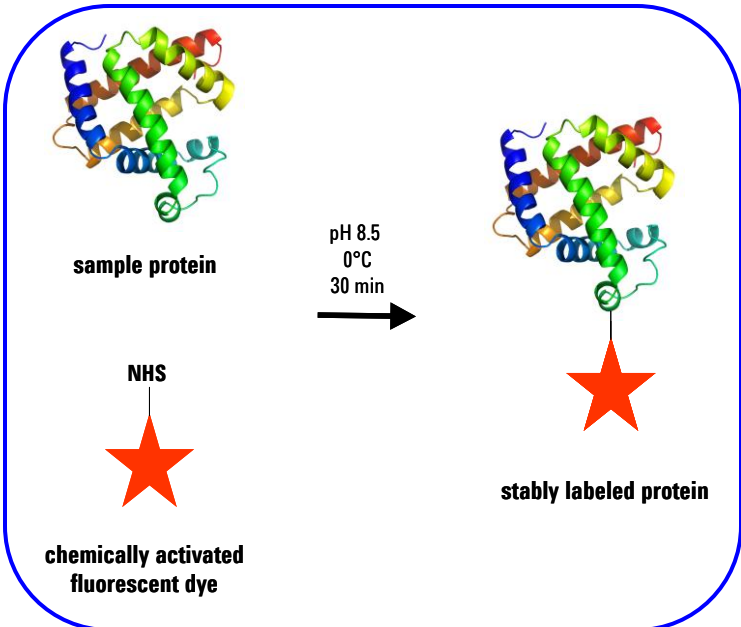


# Principles (I) -- P80/P230 Assay

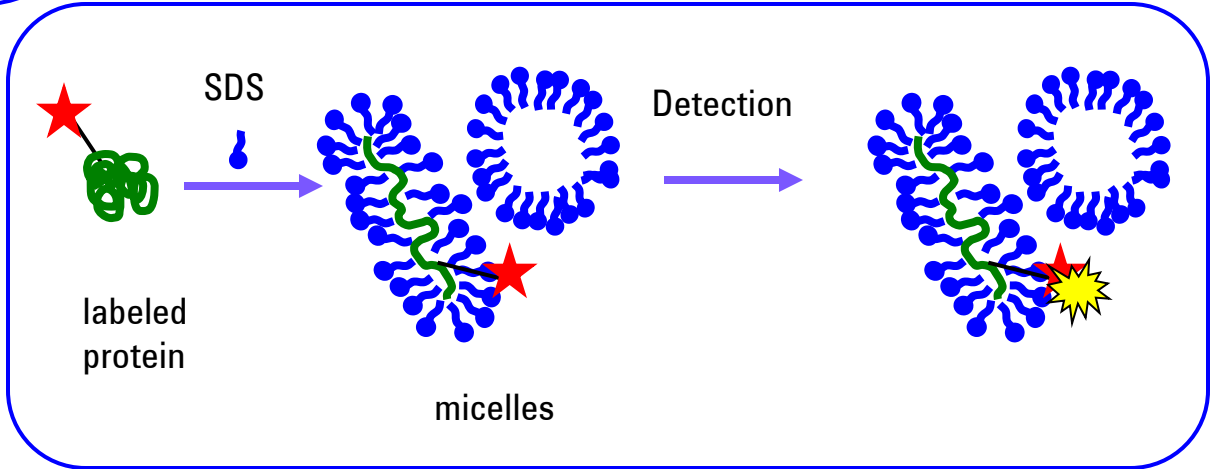




# Principles (II) -- High Sensitivity Protein Assay



Fractionation,  
Purification,  
Dilution, etc.



# Protein LabChip Applications



## Cell Lysates

- identification of over-expressed proteins
- comparison of different expression patterns

## Column Fractions

- monitoring of protein isolation and purification process
- check fractions for impurities

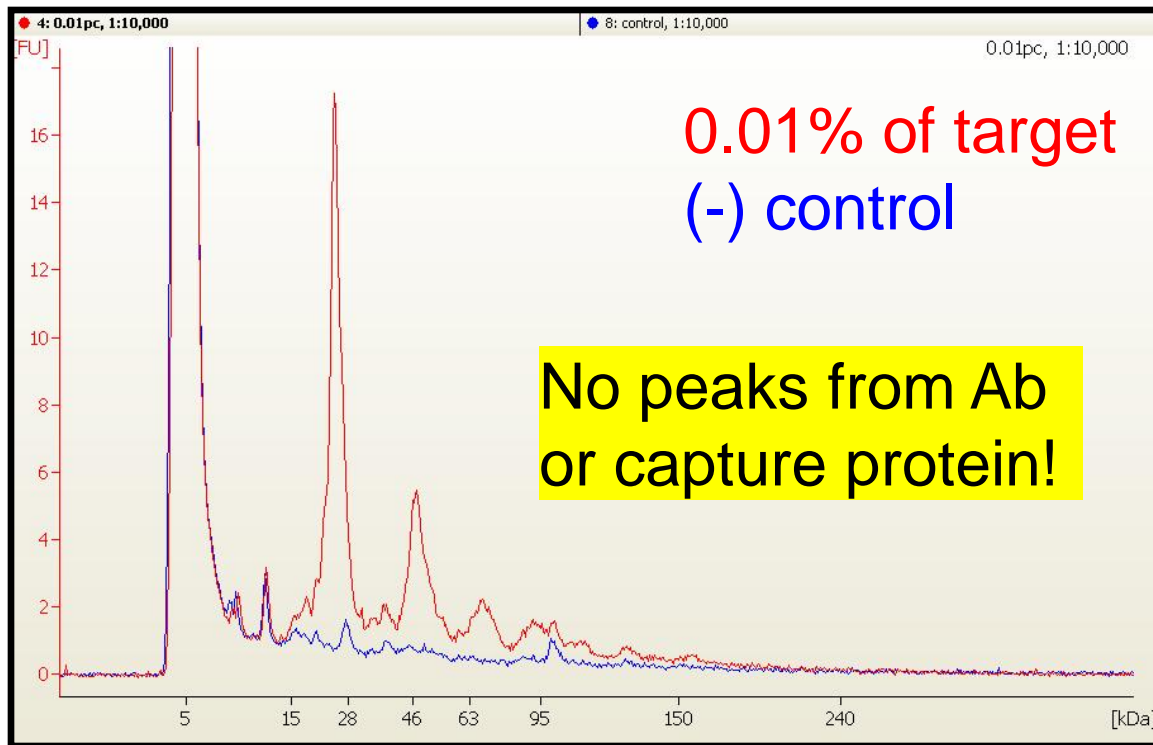
## Purified Proteins

- monitoring of impurities in protein preps
- integrity check for monoclonal or polyclonal antibodies (antibody QC)

# Immunoprecipitation: IP/HSP-250

His-tagged protein in *E. coli* background

Anti-His + Protein A magnetic beads



C. Wenz & A. Rüfer, *Electrophoresis* 2009, 30, 4264–4269

1 mg/ml *E. coli* Lysate  
+/- Target Protein(s)



HSP 250 Protein Labeling

Immunoprecipitation

Incubation with specific Antibody

Incubation with Protein A Beads

Wash with Buffer (3x)

Elution with 50% HSP-250 Sample Buffer

Direct On-Chip Analysis



2100 Bioanalyzer  
HSP-250 Assay



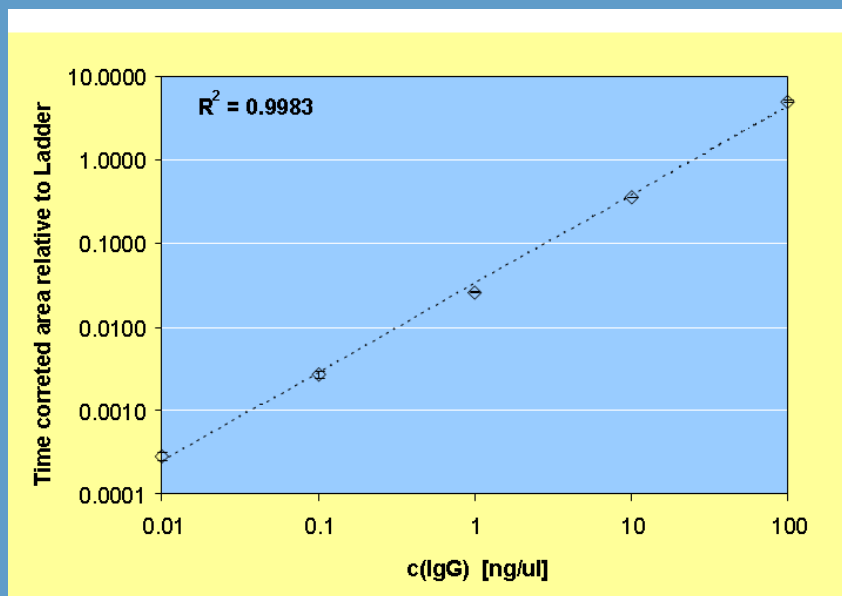
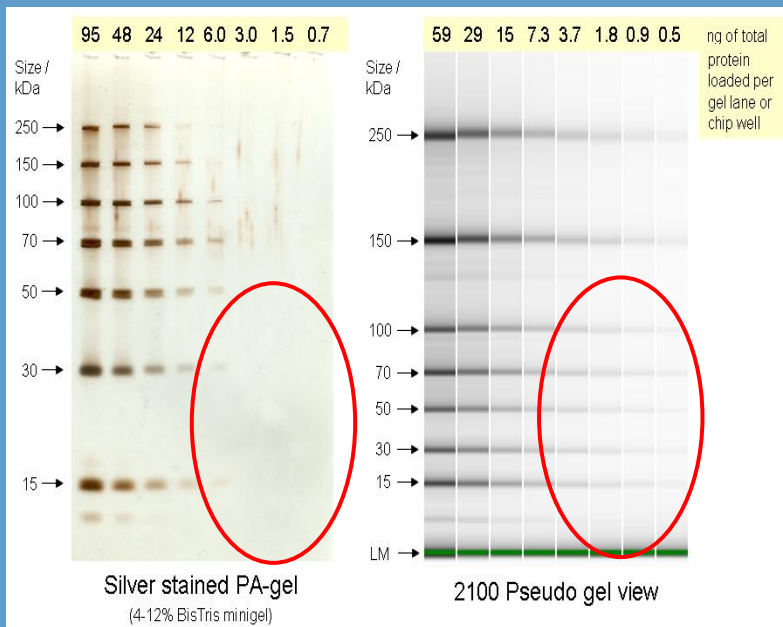
# High Sensitivity Protein 250 Kit (HSP-250)

## Highest sensitivity:

Labeled proteins can be measured down to  $\text{pg}/\mu\text{L}$  concentrations loaded on Chip

## Extra wide linear dynamic range:

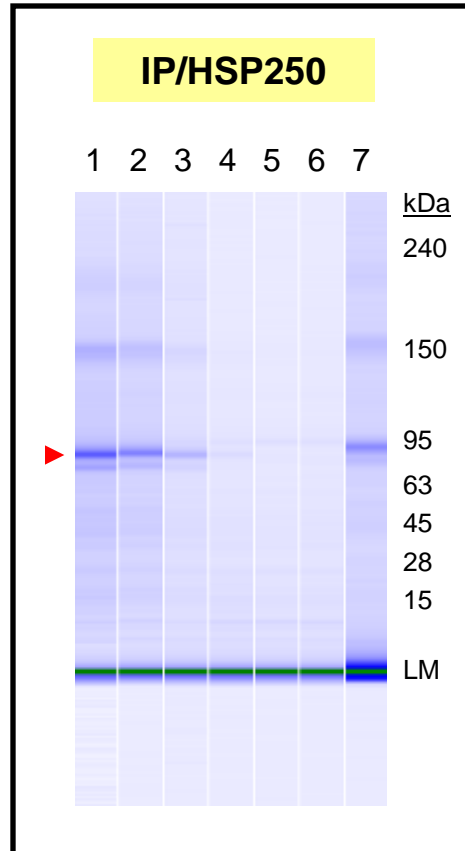
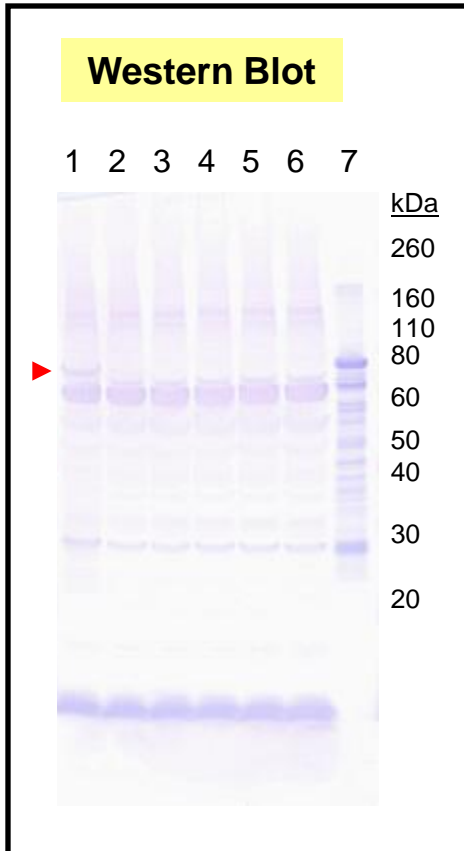
4 orders of magnitude linear dynamic range assuring excellent determination of impurities



Direct comparison of samples run on SDS-PAGE with Silver staining and on 2100 Bioanalyzer.



# Western Blot with PTEN-GST Fusion in *E. coli* Background



- 1: 1 % PTEN
- 2: 0.1 % PTEN
- 3: 0.01 % PTEN
- 4: 0.001 % PTEN
- 5: 0.0001 % PTEN
- 6: *E. coli* only
- 7: PTEN only

## Advantages of the IP/HSP-250 Method:

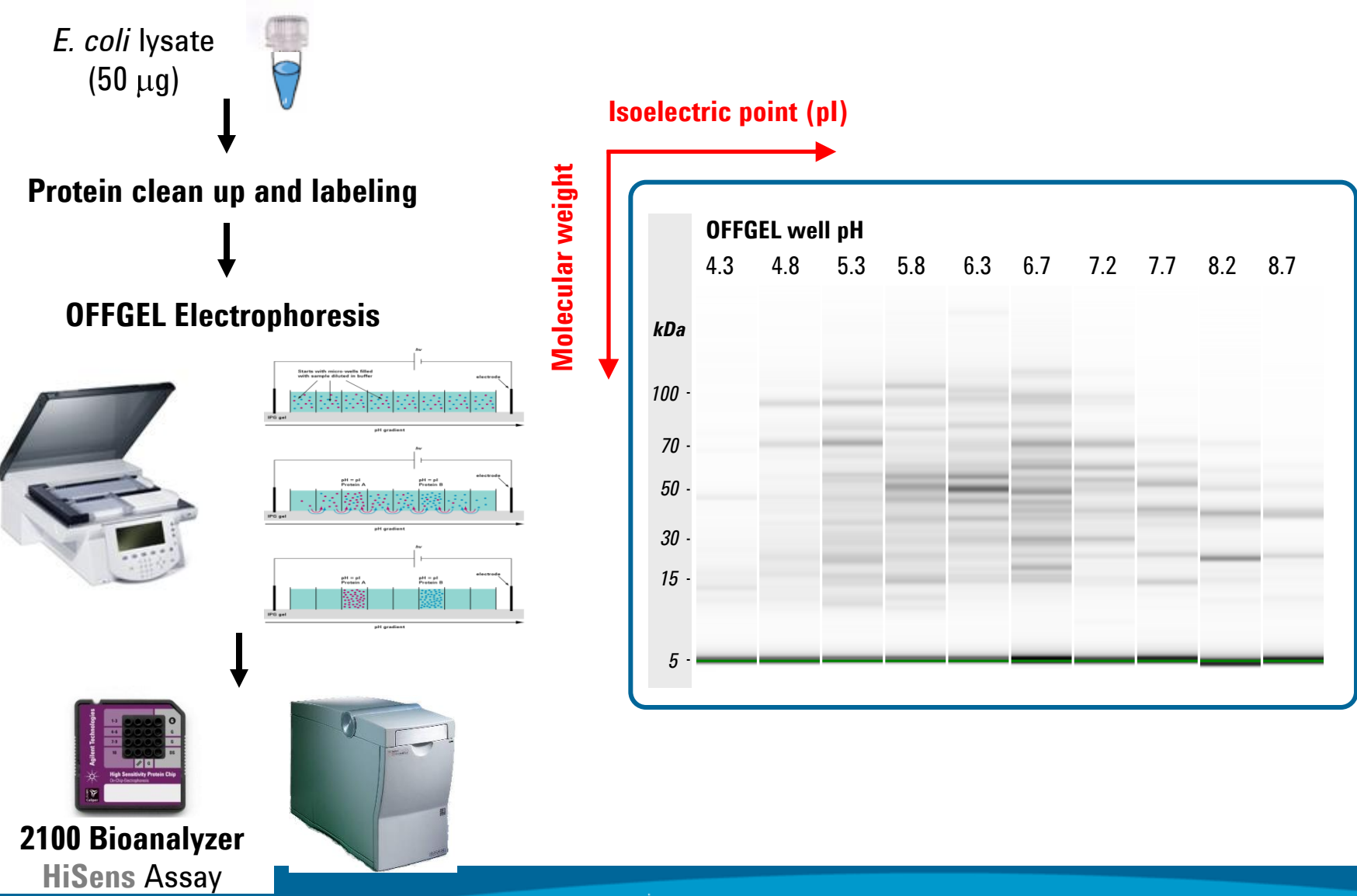
- Sensitivity
- Specificity
- Time-to-result: 3 hours
- Cost: less primary & no secondary antibodies



# High Sensitivity Protein 250 Applications Combination with IEF

5989-8489EN.pdf

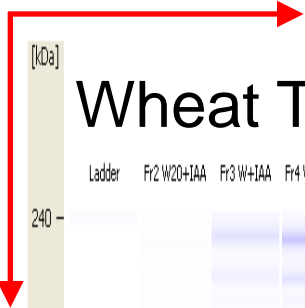
# Combination with OFFGEL Fractionation



# Wheat Analysis

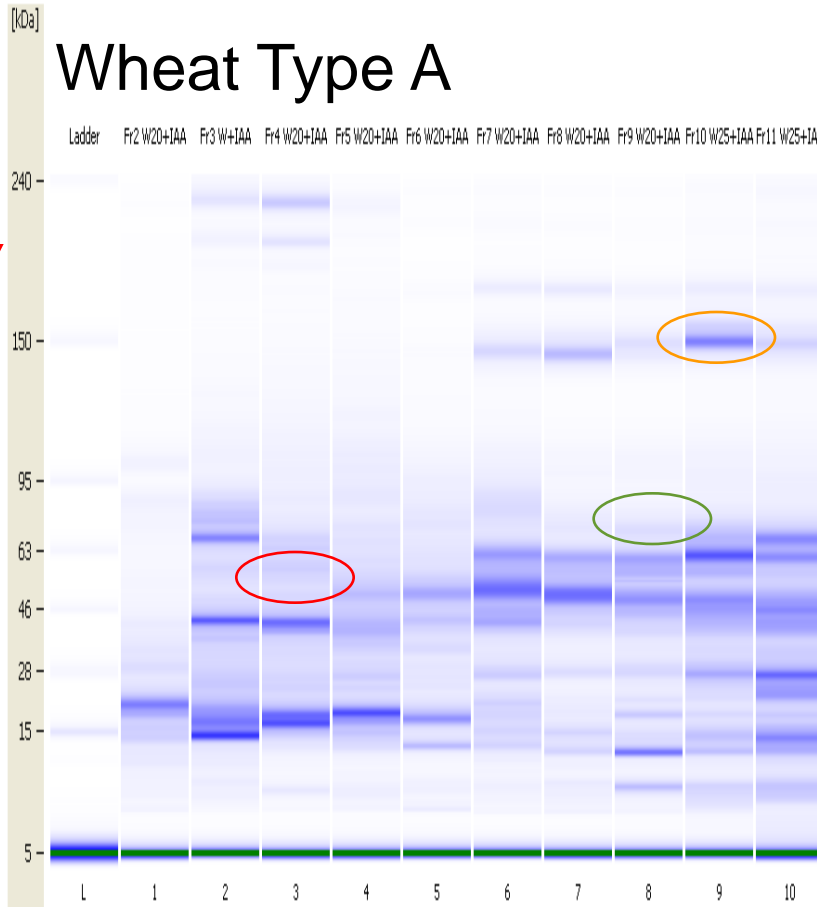
Protein extraction  
Alkylation with IAA  
Acetone precipitated  
Labeling at 10 ug/ul total protein (Bradford)  
100 ug labeled protein fractionated pH3-10  
Fractions undiluted analyzed with 250HSP-Assay

Isoelectric point (pI)



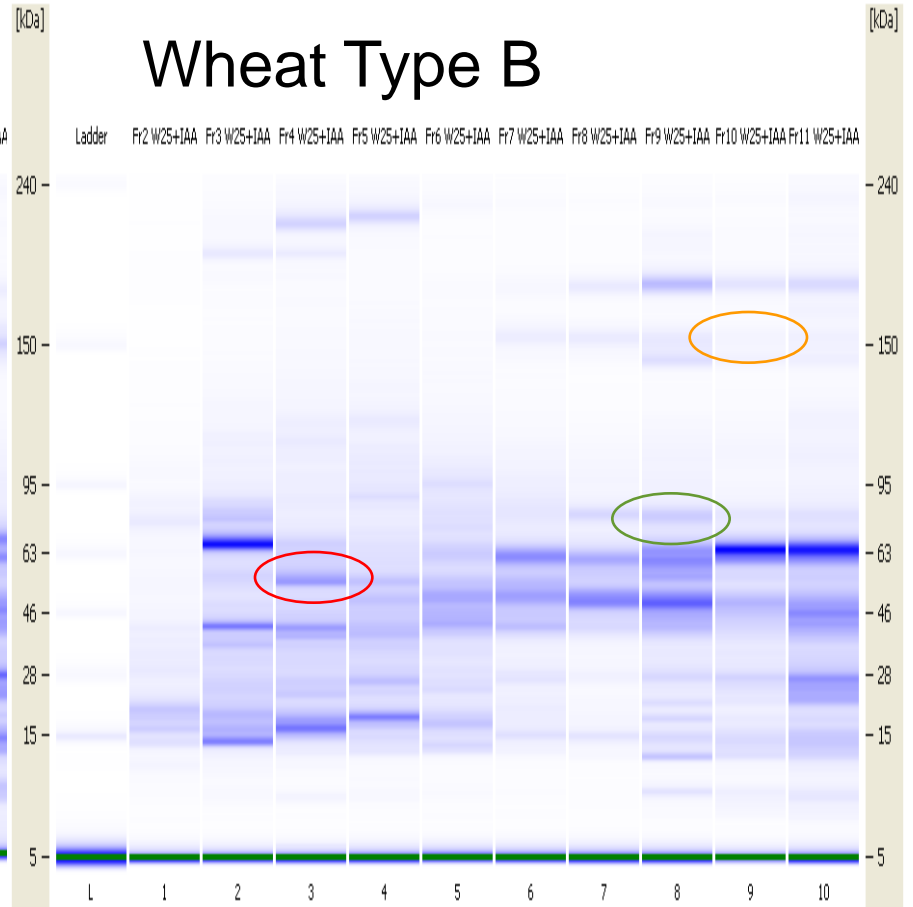
## Wheat Type A

Ladder Fr2 W20+IAA Fr3 W+IAA Fr4 W20+IAA Fr5 W20+IAA Fr6 W20+IAA Fr7 W20+IAA Fr8 W20+IAA Fr9 W20+IAA Fr10 W25+IAA Fr11 W25+IAA




## Wheat Type B

Ladder Fr2 W25+IAA Fr3 W25+IAA Fr4 W25+IAA Fr5 W25+IAA Fr6 W25+IAA Fr7 W25+IAA Fr8 W25+IAA Fr9 W25+IAA Fr10 W25+IAA Fr11 W25+IAA







BioAnalyzer Support:  
[Bioanalyzer@agilent.com](mailto:Bioanalyzer@agilent.com)  
Send .xad file not .pdf file

## Q & A