

Automated preparation of samples for fatty acid analysis using the Agilent workbench

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*Workbench?:

What it does and it's function

*Fat extraction methods we employ:

Various methods in fat determination and samples that are analysed

*How the workbench is incorporated into these methods:

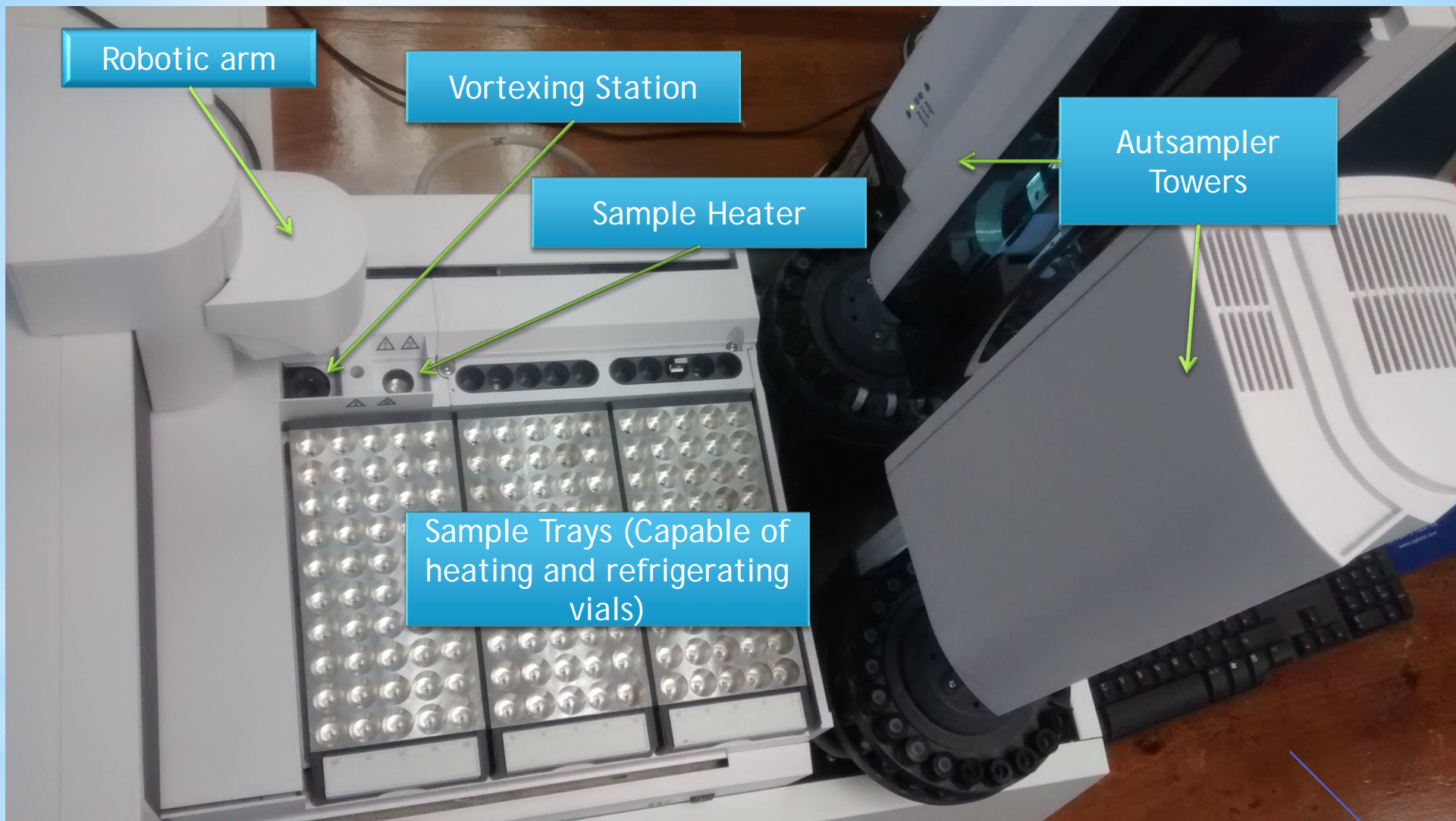
Its use in methods and its advantages

*Overview



- * Agilent 7696A Sample prep Workbench
- * Standalone automated sample preparation platform
- * Capable of volumetric transfer of liquids between 2ml vials
- * Performs accurate addition of reagents, dilutions, vortexing, heating of samples etc.
- * Compliment to a GC/HPLC instrument

* **Workbench**



Robotic arm

Vortexing Station

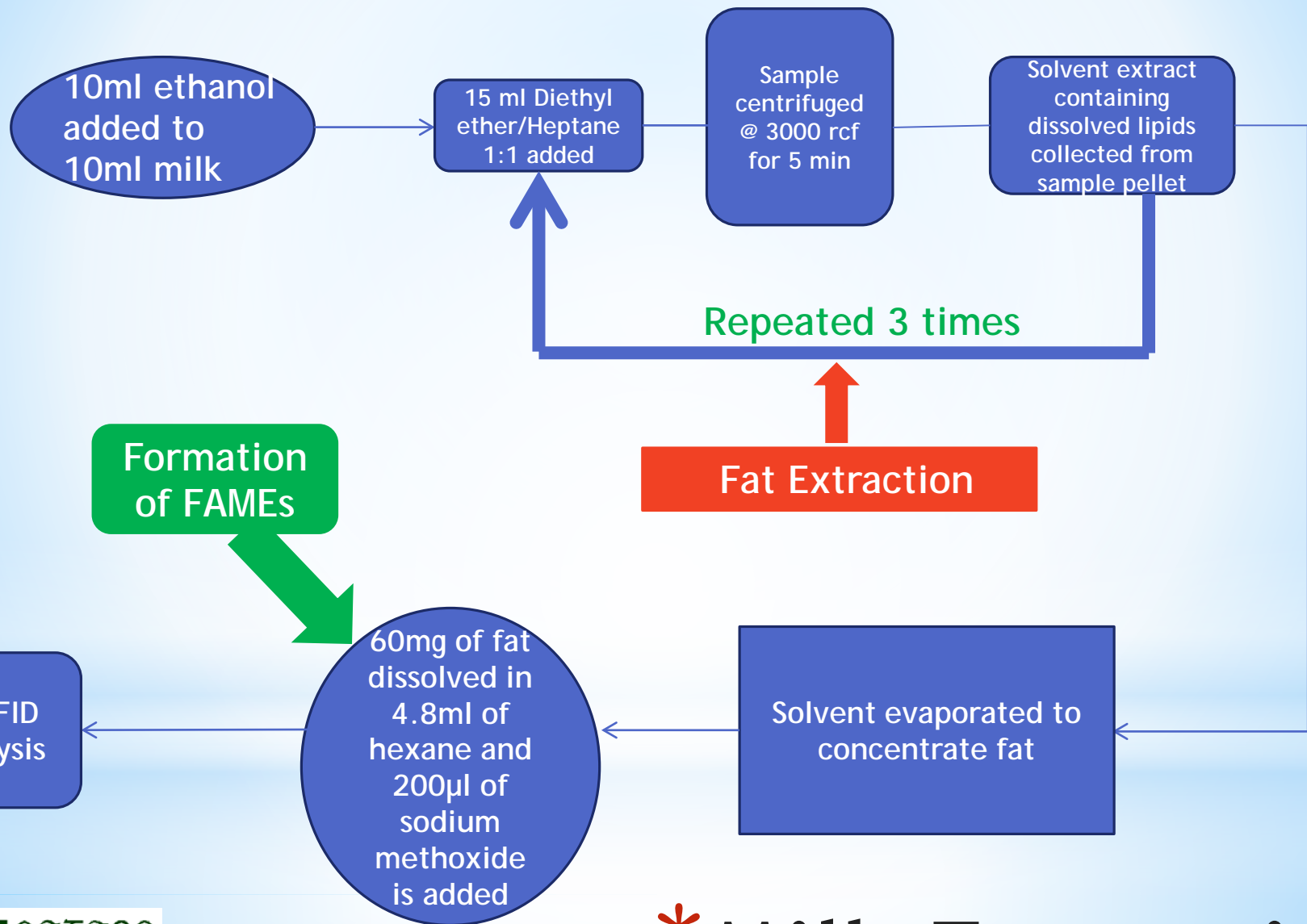
Sample Heater

Autosampler
Towers

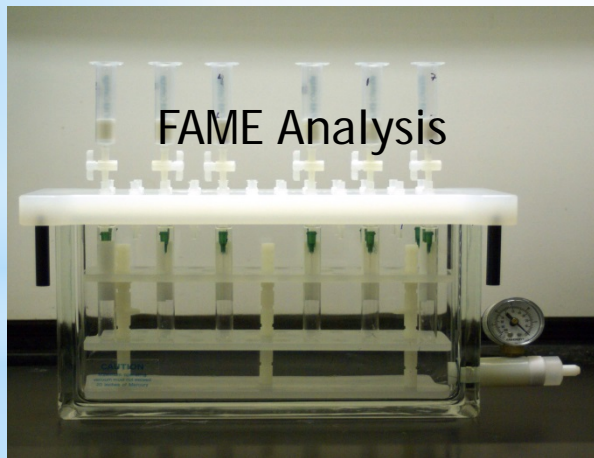
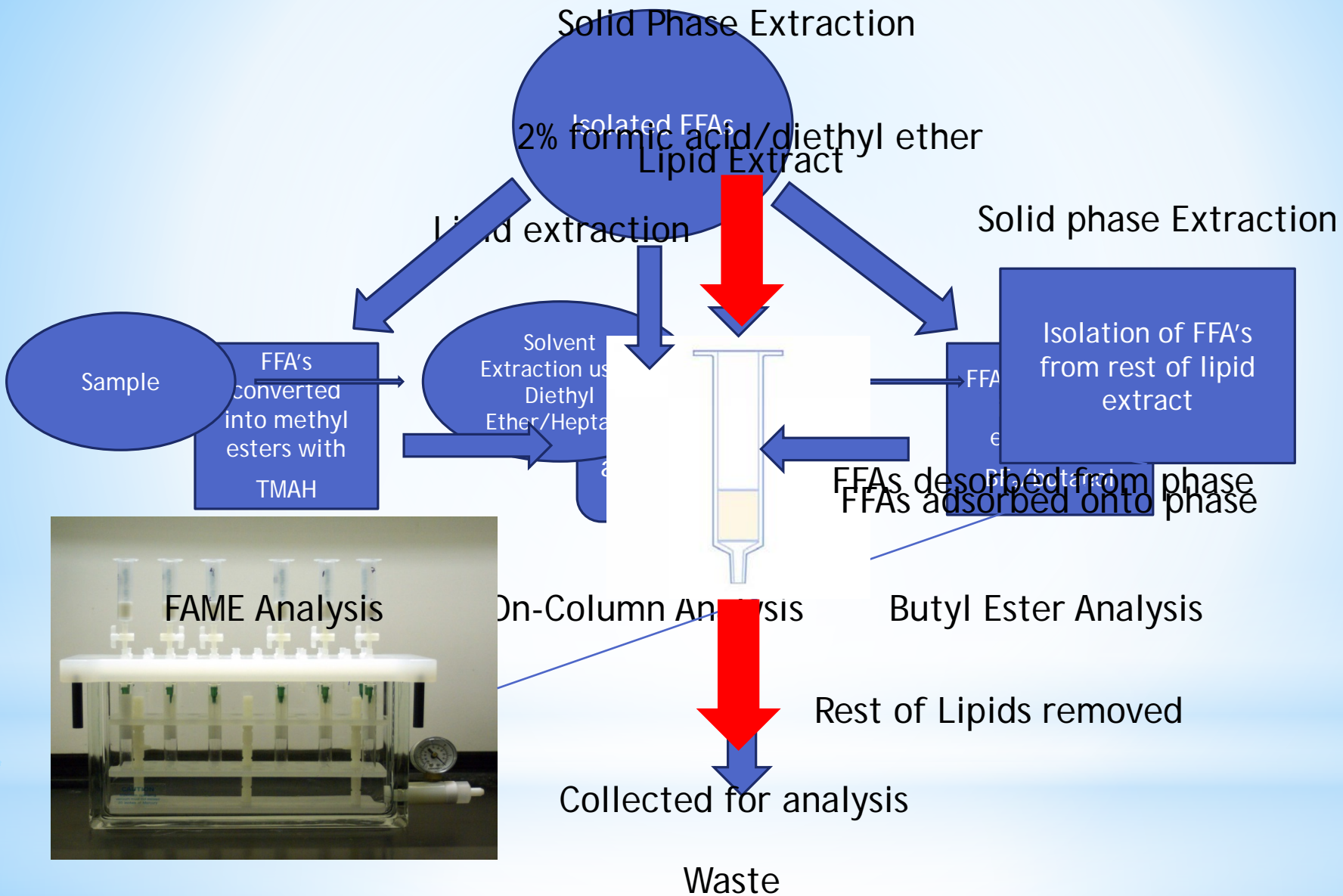
Sample Trays (Capable of
heating and refrigerating
vials)

- * Triglyceride Determination → Analysed as fatty acid methyl esters (FAMES) using sodium methoxide in methanol (NaMeO)
- * Free Fatty Acids → Analysed as Free fatty acids, FAMES using tetramethylammonium hydroxide (TMAH) or butyl esters using boron trifluoride (BF₃) in butanol
- * Analysis of milk, cheese, milk powders, infant formula etc..

* Fat Extraction



* Milk Extraction



* FFA extraction

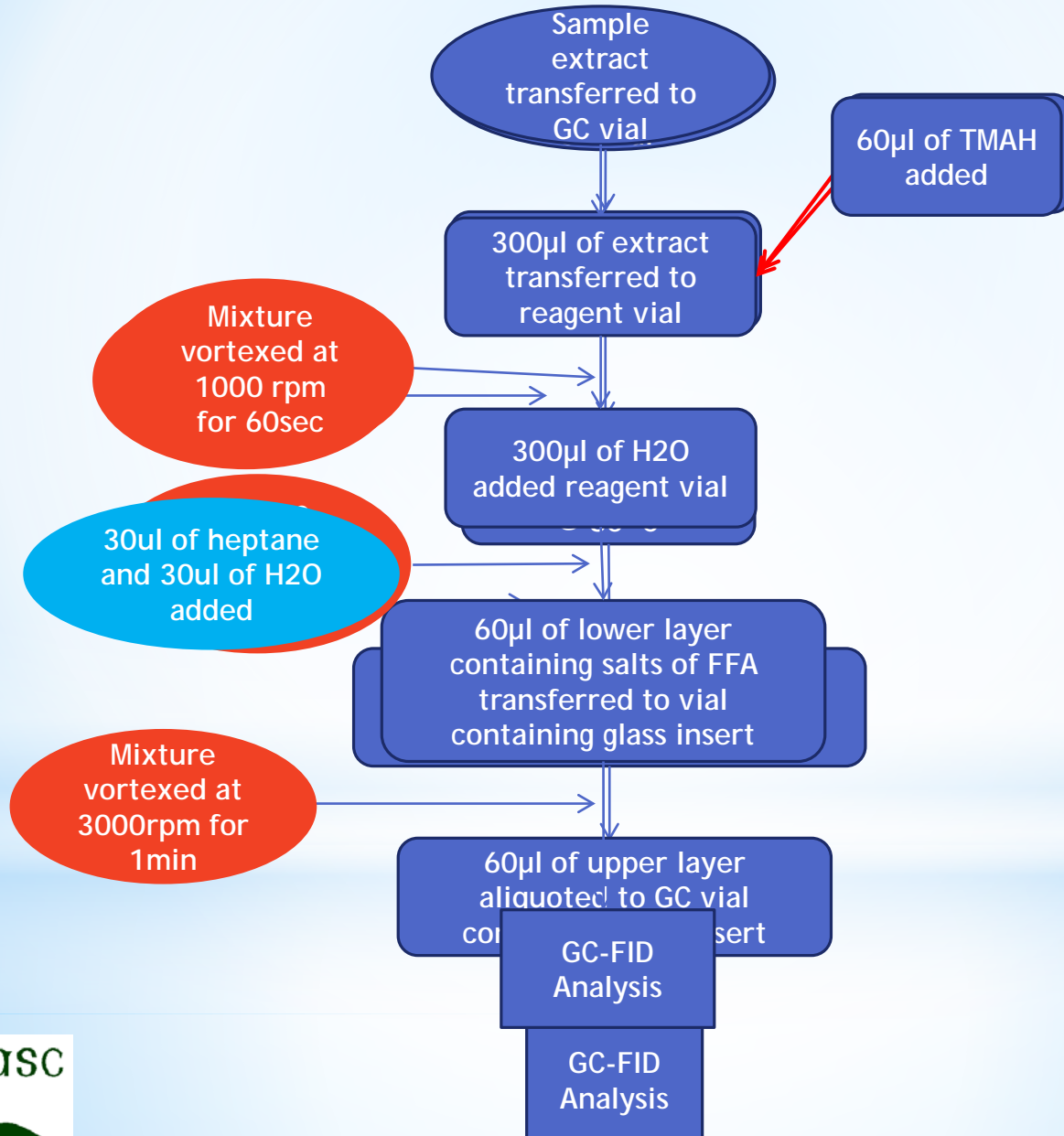
Methods:

- * Derivatisation of triglycerides to FAMES using NaMeO
- * Derivatisation of FFAs to FAMES using TMAH
- * Derivatisation of FFAs to butyl esters using BF₃/Butanol

Incorporating the Workbench:

- * Used to automate the derivatisation steps
- * Used in sample and standard preparation eg. Dilution of sample extract, preparation of calibration levels etc.

Butyramide



Robotic arm

Butyl Ester Derivatisation

Autsampler Towers

Sample mixture vortexed

Reagents added to samples

Sample and reagent vials

Butyl esters transferred to GC vial for analysis

Advantages:

* Less labour
* Reduced solvents and reagents (60 µl of BF₃ per sample)
* Improved accuracy and precision (no human error)
* Sample and reagents are always contained during preparation

Sample trays (capable of heating and refrigerating vials)

Internal Standard Analysis

Curve Type: Linear

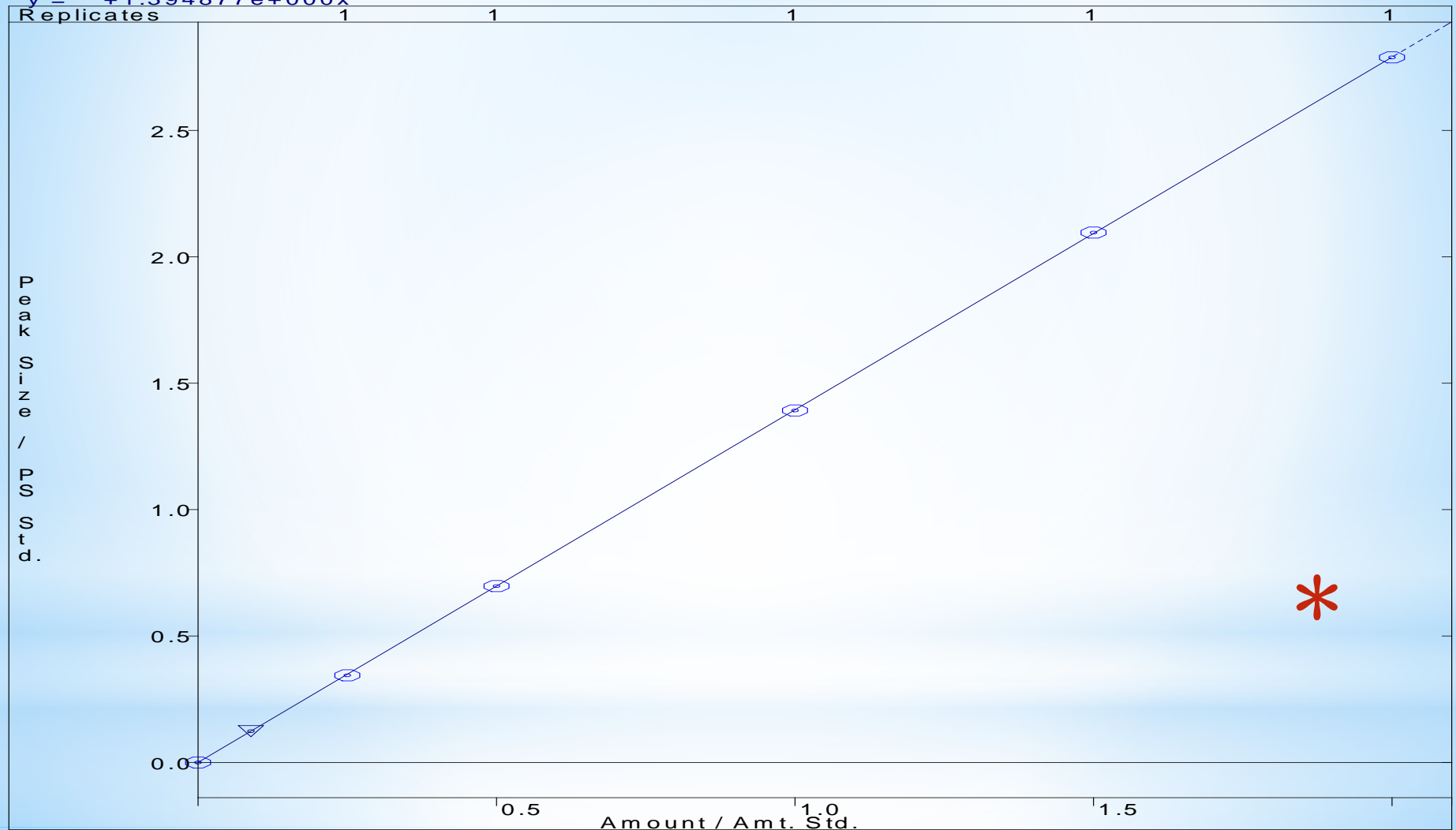
Origin: Force

$y = +1.394877e+000x$

C10

Resp. Fact. RSD: 0.5560%

Coeff. Det.(r²): 0.999995



	On-Column	FAME	Butyl Ester	RSD (%)
Brie	1135	1067	1183	5.2
Processed Cheese	1104	952	1246	13.4
Light Cheddar	969	827	814	9.8
Mild Cheddar	1525	1627	1436	6.2
Special Reserve	2289	2316	1936	9.7
Blue Stilton	3109	2856	3355	8.0
Milk Powder	337	333	296	7.1
Infant Formula	173	227	217	13.8
Butter	1492	1295	1960	21.6
Yoghurt	182	246	223	14.9
Ice Cream	323	176	661	64.3
Milk	290	286	90	51.4
Nat. Butter Powder	73652	90949	67206	15.9
Nat. Cream Powder	48680	49274	44712	5.2
Nat. Cream paste	126615	114969	131622	6.9
Nat. Butter paste	106079	98129	106748	4.6
Nat. Blue Cheese paste	116085	104376	113377	5.5

	On-Column (R ²)	FAME (R ²)	Butyl Ester (R ²)
C4:0	0.9999	0.9978	0.9996
C6:0	0.9998	0.9995	0.9995
C8:0	0.9997	0.9997	0.9964
C10:0	0.9999	0.9999	0.9999
C12:0	0.9999	0.9998	0.9998
C14:0	0.9999	0.9998	0.9978
C16:0	0.9999	0.9999	0.9996
C18:0	0.9998	0.9999	0.9999
C18:1	0.9998	0.9998	0.9999
C18:2	0.9998	N/A	0.9999
C18:3	0.9994	N/A	0.9999
LOD (ppm)	0.7	5	8
LOQ (ppm)	3	20	20

Analysis of 17 dairy samples. Displayed are the average ppm (mg/kg) values obtained from six replicates (n=6)

Validation data obtained, included is correlation coefficient (R²), limit of detection (LOD) and limit of quantification (LOQ)

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Comparison and validation of 2 analytical methods for the determination of free fatty acids in dairy products by gas chromatography with flame ionization detection

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*** Any Questions?**