

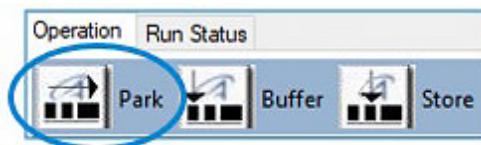
# Long Term Storage of Your Parallel CE System



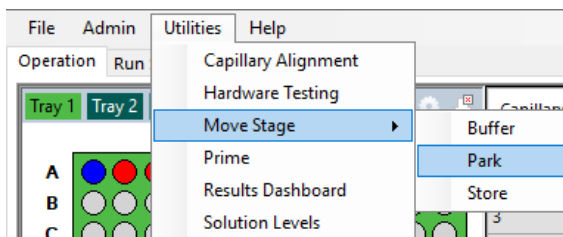
## Preparing the system for prolonged downtime or laboratory closure

For prolonged storage of the system while out of the laboratory the capillary array should be left installed in the system.

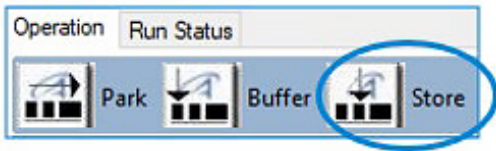
1. Ensure gel is in the instrument reservoir;
  - a. 5200/5300/5400 Fragment Analyzer, Femto Pulse, ZAG DNA Analyzer: Perform a Full Conditioning.
  - b. Oligo Pro II: Run a Step 3 for 15 minutes with gel only.
2. Select **Park** to place the storage plate in the drawer and move the stage to the bottom of the instrument;
  - a. 5200/5300/5400 Fragment Analyzer, Femto Pulse, ZAG DNA Analyzer



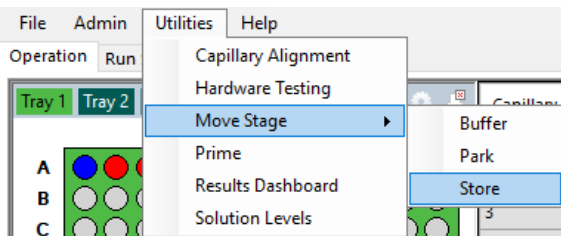
### b. Oligo Pro II



3. Replace storage solution plate with a new plate
  - a. 5200 Fragment Analyzer, Femto Pulse: Place 1.0mL/well of storage solution into Row H of buffer plate.
  - b. O5300/5400 Fragment Analyzer, ZAG DNA Analyzer, Oligo Pro II systems: Place 200  $\mu$ L/well of storage solution in fresh sample plate in drawer 3.
    - i) 48 capillary unit rows A-D only
    - ii) 96 capillary unit rows A-H
4. Select **Store** to move storage plate under the capillaries.
  - a. 5200/5300/5400 Fragment Analyzer, Femto Pulse, ZAG DNA Analyzer



b. Oligo Pro II



5. Close software, turn off PC and instrument.

## Powering on system after prolonged downtime or laboratory closure

After prolonged storage of the system it is recommended to perform a Method B – Hot Water Soak of the capillaries followed by a Method C – 0.5N NaOH flush

1. Power on the PC, instrument and software.
2. Replace the storage solution as described in steps 2-4 above.
3. Perform a Method B – Hot Water Soak as outlined in the system guide appendix.\*
4. Perform a Method C – 0.5N NaOH flush as outlined in the system guide appendix.\*
5. Prepare fresh conditioning solution and separation gel according to the Kit Guide or Quick Guide, update solution levels in the software and prime lines.

\*The Oligo Pro II system does not have appendices for Method B or Method C. Place a buffer tray filled with 1.0 mL/well of hot water (150 °F to 200 °F) in the buffer position and send to this location for 30 minutes.

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