

Plant Metabolomics

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Node Leader, Metabolomics Australia
Past Immediate President, International Metabolomics Society



BIOPLATFORMS
AUSTRALIA

NCRIS
National Research
Infrastructure for Australia
An Australian Government Initiative



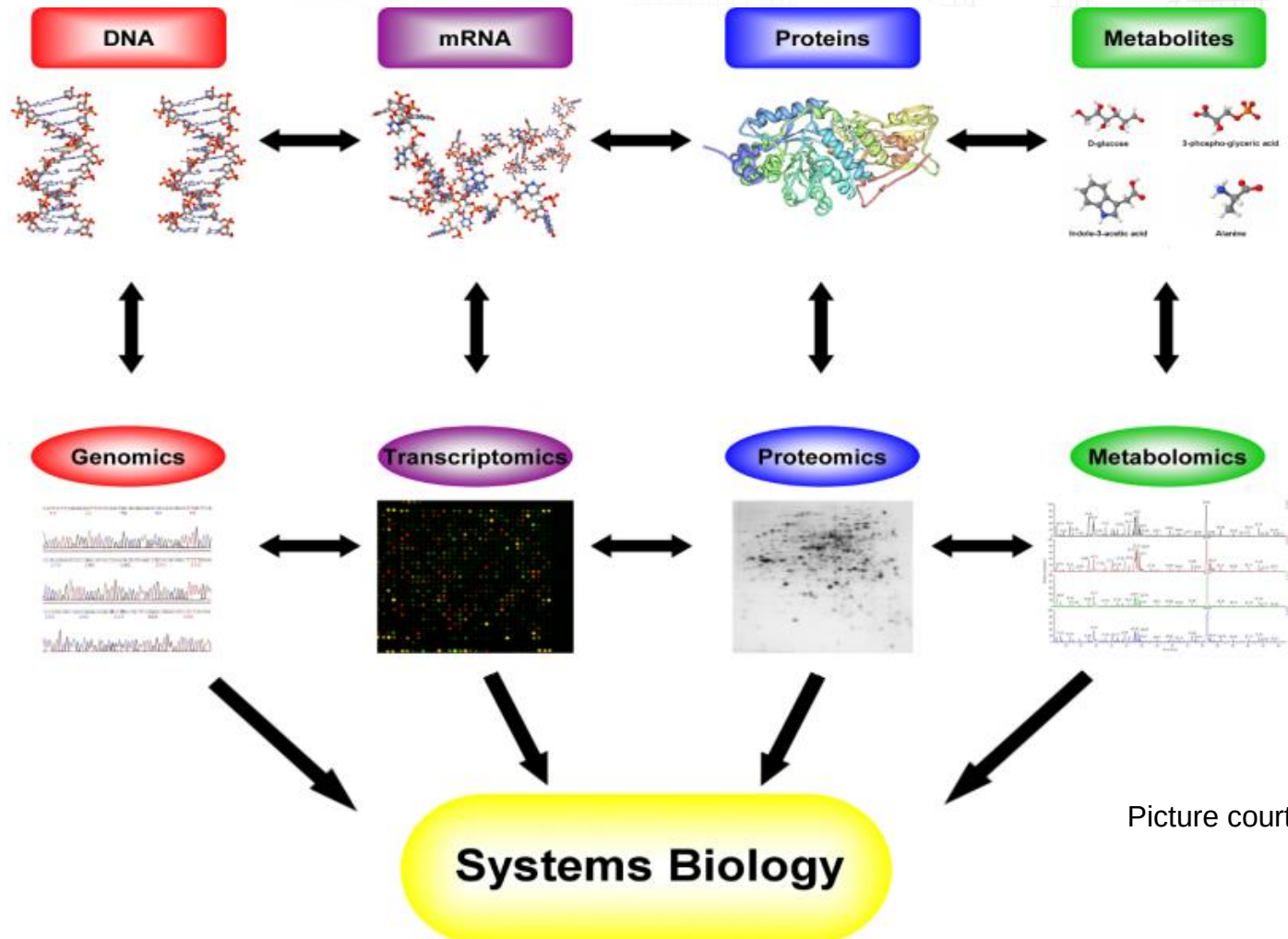


What can happen

What appears to be happening

What makes it happen

What is happening and has happened



Picture courtesy J. Bowne

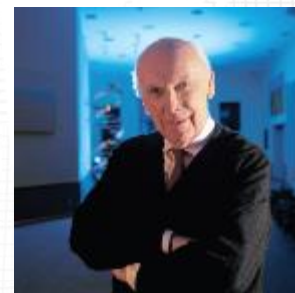
What is Metabolomics about?



This is the analytical approach of a large-scale, non-targeted and high-throughput determination of metabolites in a biological system

The real challenge of metabolomics is to comprehensively cover the analysis of many compounds having many different chemical structures

***“Think metabolism...
If I were doing a PhD [in
cancer research], I'd be
doing it in metabolomics”.***



Prof. James Watson, Nobel Laureate
Co-discoverer of the structure of DNA

The “Other” Metabolomes

(courtesy of Prof. David Wishart)



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3230 (T3DB)

Toxins/Env. Chemicals

1240 (DrugBank)

Drug metabolites

28500 (FooDB)

Food additives/Phytochemicals

1550 (DrugBank)

Drugs

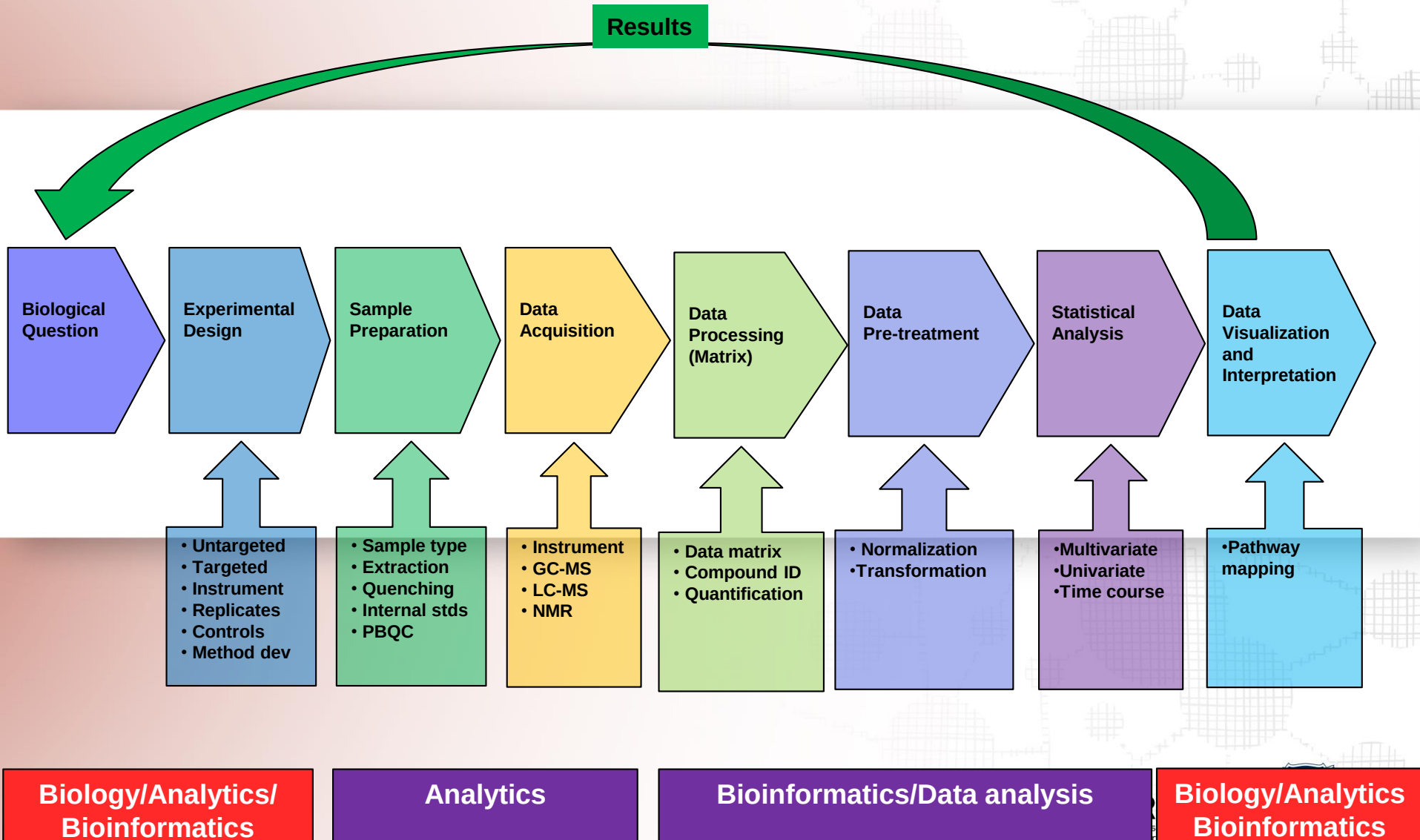
19700 (HMDB)

Endogenous metabolites



Courtesy Prof David Wishart

Metabolomics Workflow



What types of metabolites can be analysed?



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All low molecular weight metabolites
(<1500 Daltons)

Includes, but not limited to:

Sugars, sugar phosphates

Organic acids

Amino acids,

Fatty acids

Nucleosides

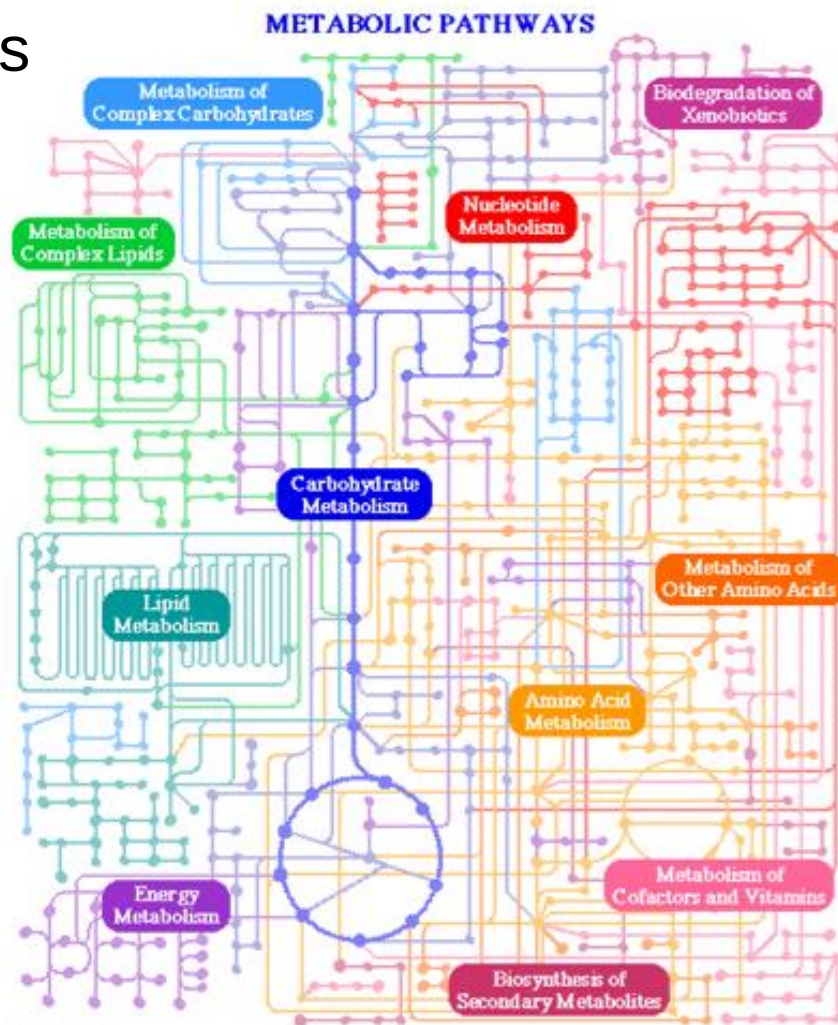
Lipids (i.e. sterols, phospholipids)

Small peptides

Other secondary metabolites

Drugs, insecticides, other xenobiotics

Total number of metabolites $> 500,000$

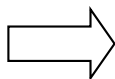


metabolites per cell $\sim 1,000$ to $6,000$

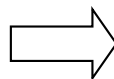
Comprehensive workflow



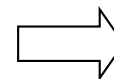
tissue harvest



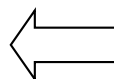
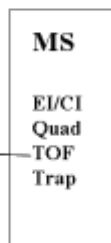
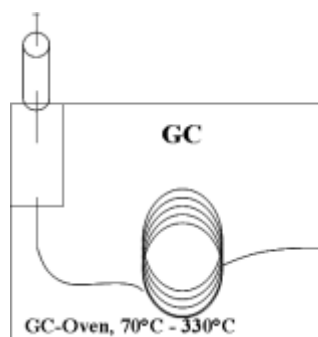
shock freeze



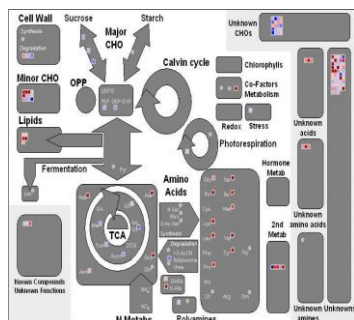
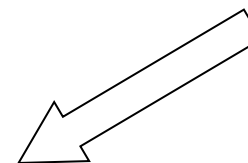
homogenization



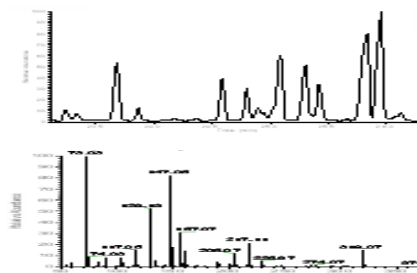
extraction procedure



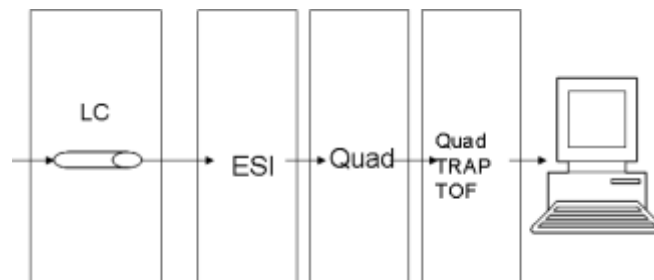
derivatisation



data interpretation

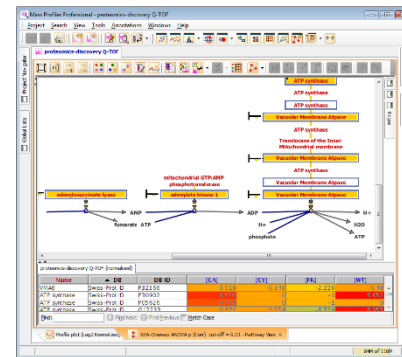
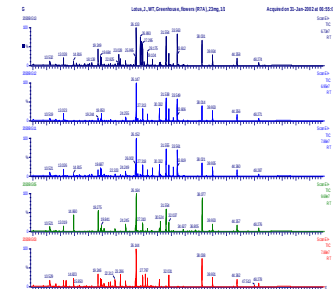


data analysis



extraction procedure

Metabolomics



Example: LC-MS approach



Sample



Polar extract (e.g. MeOH)

Lipid extract (e.g. chloroform)

Apolar
compounds

Polar
compounds

Lipid
profile

Lipid species
quantification

C18RP

+/- ESI QTOF

HILIC (ANP)

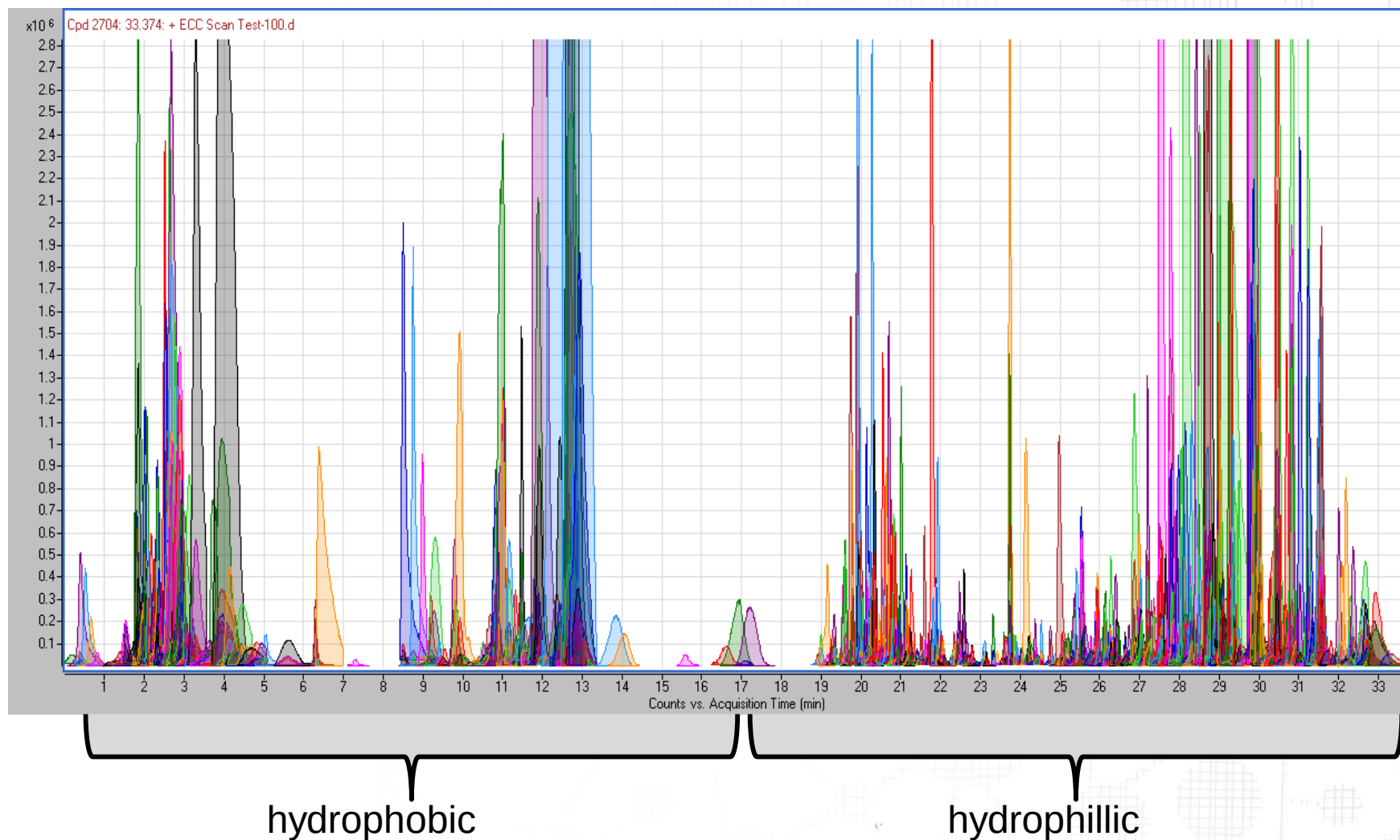
+/- ESI QTOF

LC-QTOF

MRM using QQQ

Tandem LC-QqTOF-MS

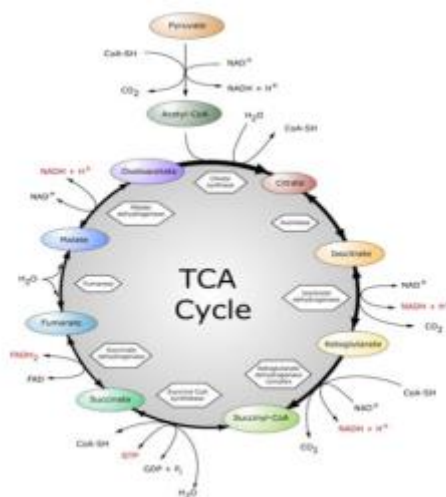
~ 2700 compounds



Targeted vs Untargeted

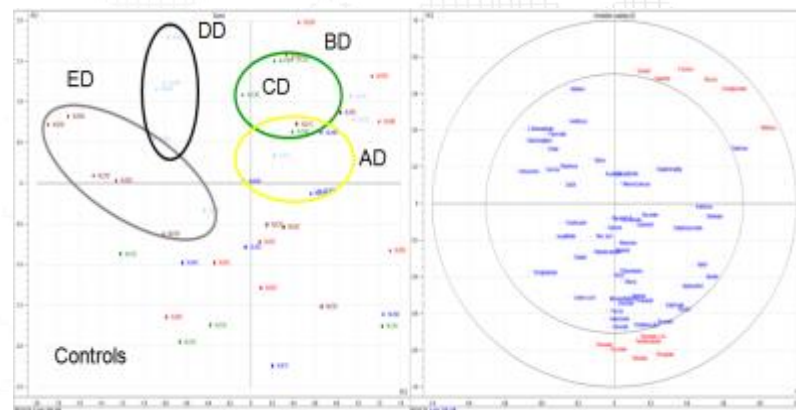
“detect the expected”

Absolute quantification
Pool sizes
You know what you measure
Not comprehensive
More accurately
Better sensitivity
Easier to interpret
Pathway mapping



“detect the unexpected”

Semi-quantitative
X-fold comparison
Biomarker discovery
More comprehensive
Loads of unknowns
Multivariate analyses

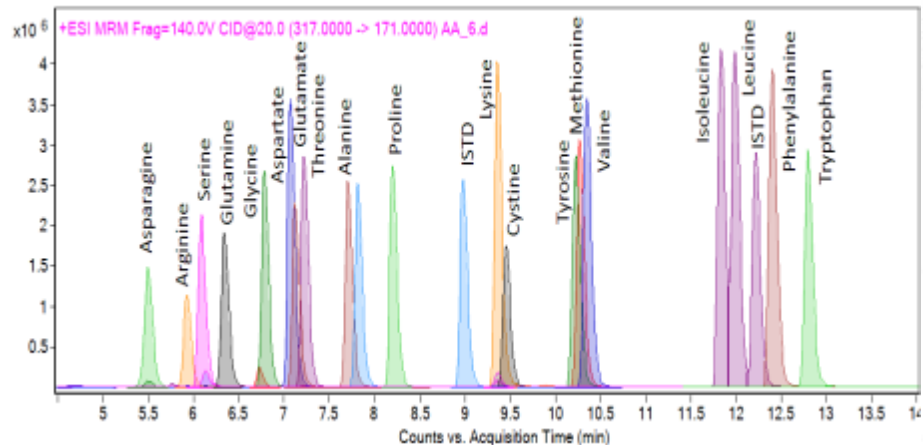


Quantitative metabolomics



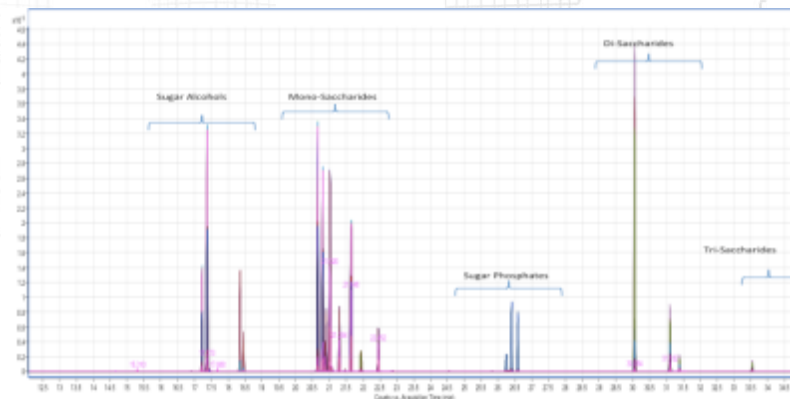
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Boughton et al. 2011

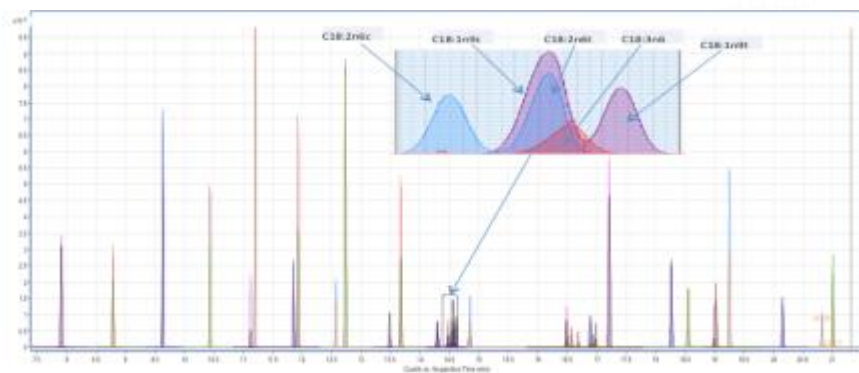


36 amino acids and 30 amines

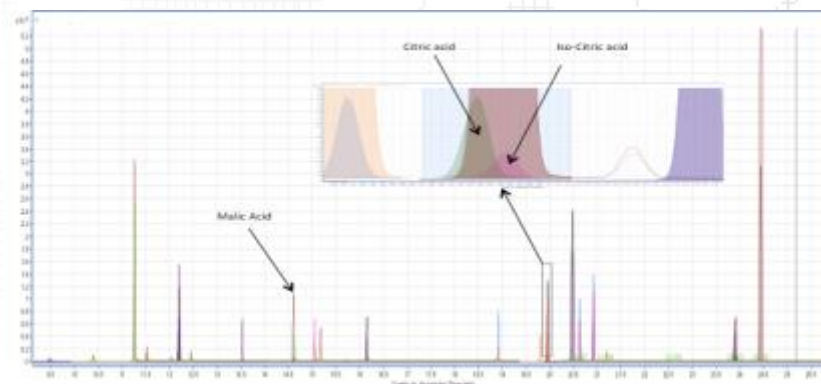
Dias et al. 2015



47 sugars and sugar alcohols



36 fatty acids



22 organic acids

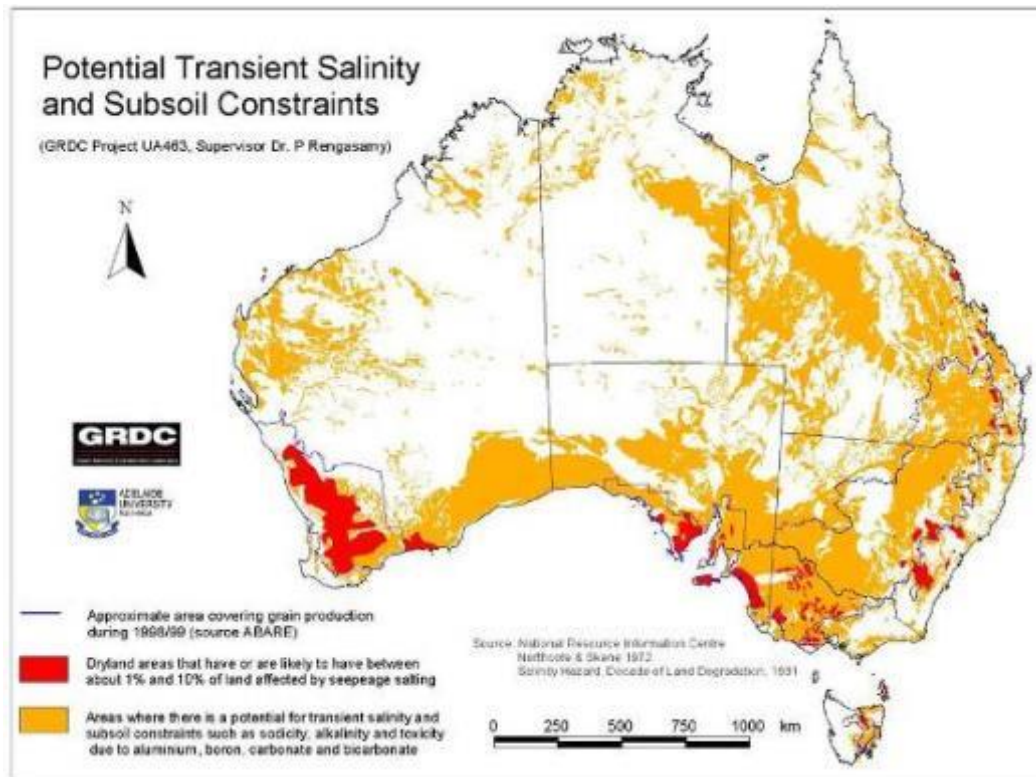
Identifying novel salinity tolerance mechanisms by spatial analysis of lipids in barley roots



Australian Government
Australian Research Council



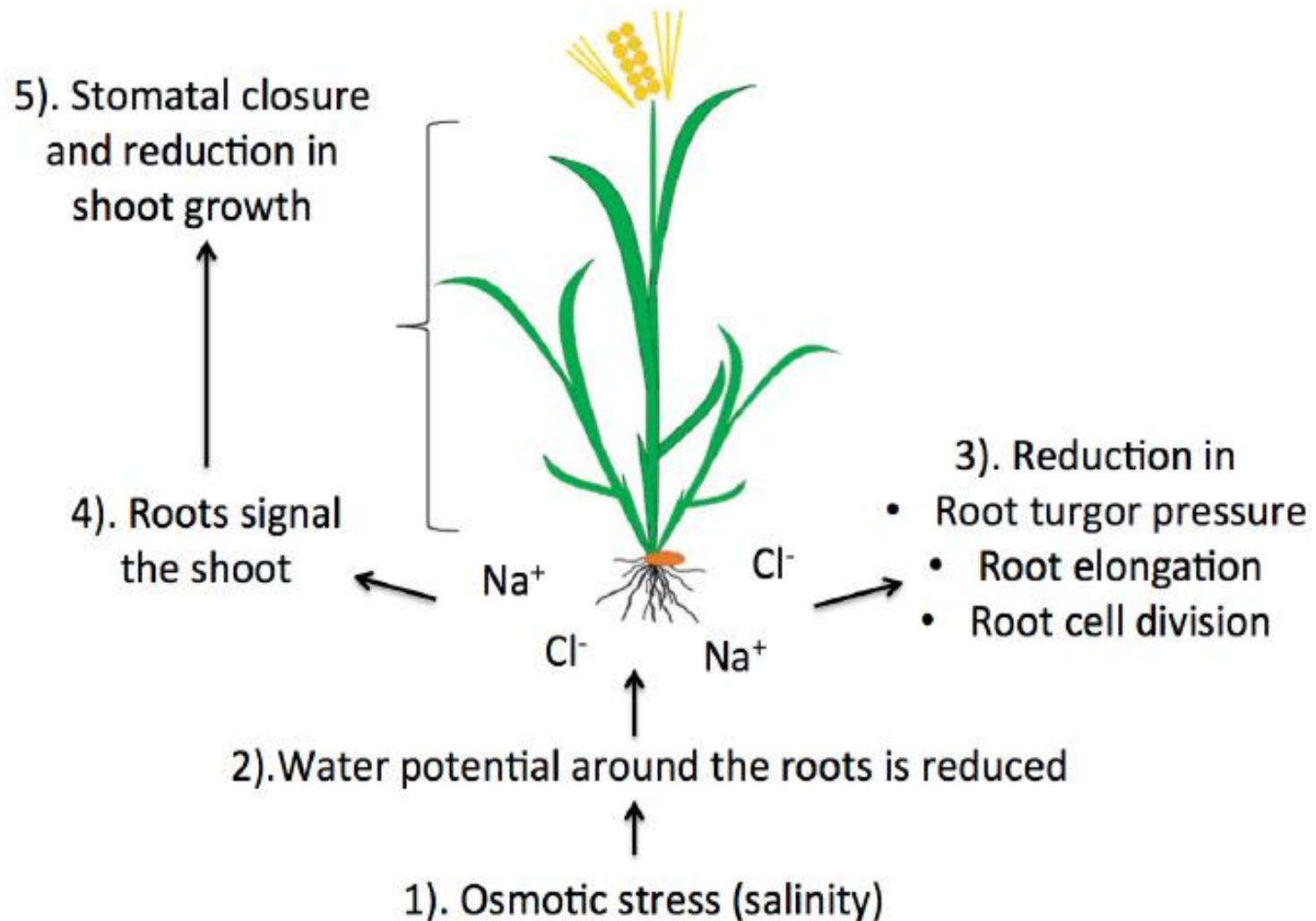
Abiotic stress affects agricultural productivity in Australia



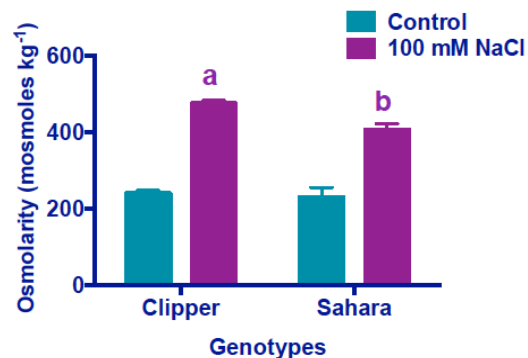
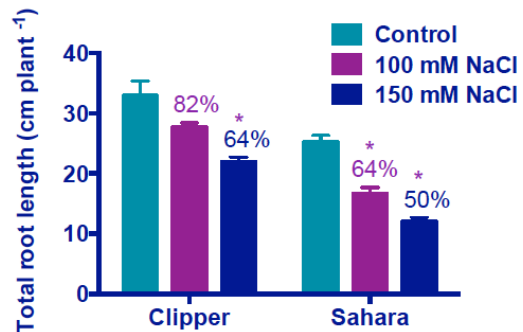
Australian wheat belt significantly affected by salinity
In danger: Australian wheat exports worth ~ \$4 billion p.a.

Rengasamy (2002) *Aust. J. Exp. Agric.* **42**: 351-361

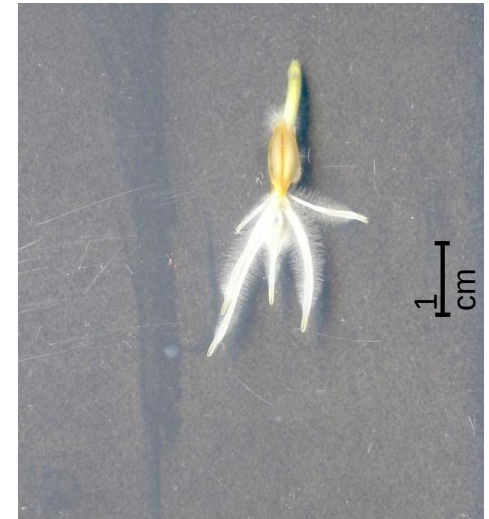
Effect of salt stress on plants



Effect of salt stress on roots



Control



Salt

CSIRO PUBLISHING

Functional Plant Biology

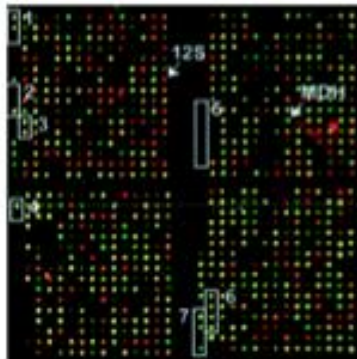
<http://dx.doi.org/10.1071/FP12290>

Genetic variation in the root growth response of barley genotypes to salinity stress

Megan C. Shelden^{A,E}, Ute Roessner^A, Robert E. Sharp^D, Mark Tester^B and Antony Bacic^C

Project AIM: Understand early tissue-specific salt responses

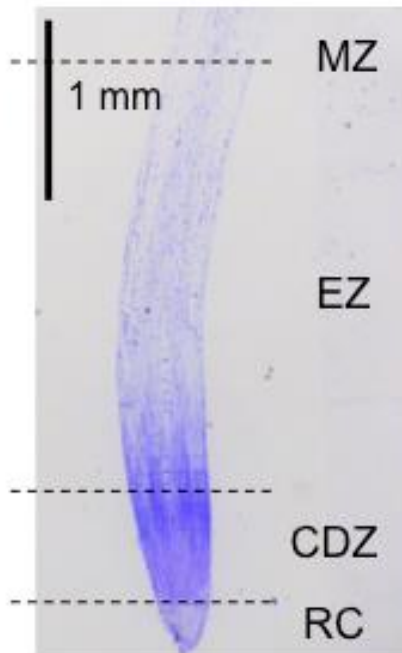
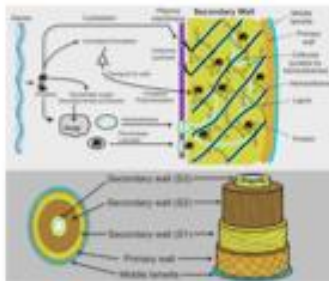
mRNA



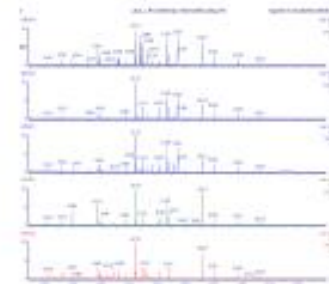
Lipids



Cell Wall



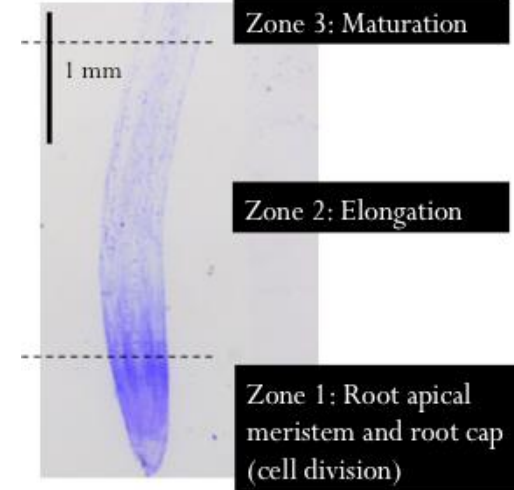
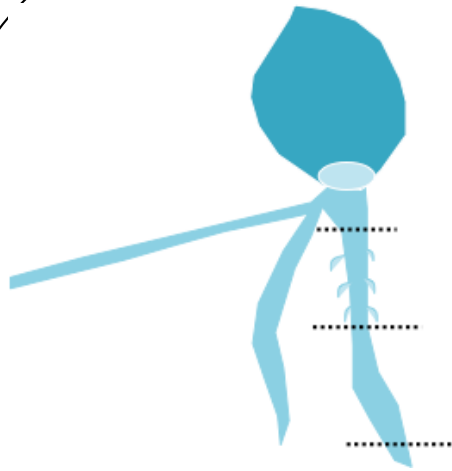
Metabolites



In collaboration with Plant Cell Wall Centre
T. Bacic, M. Doblin, A. Cassin

In collaboration with MA

Study Design



Barley Cultivars: Clipper and Sahara

Growth and Treatment on Agar Plates

Root Tissue Harvest
(3 Zones: Root Cap, Elongation and Maturation Zone)

1. Cell Wall
Polysaccharide Analysis

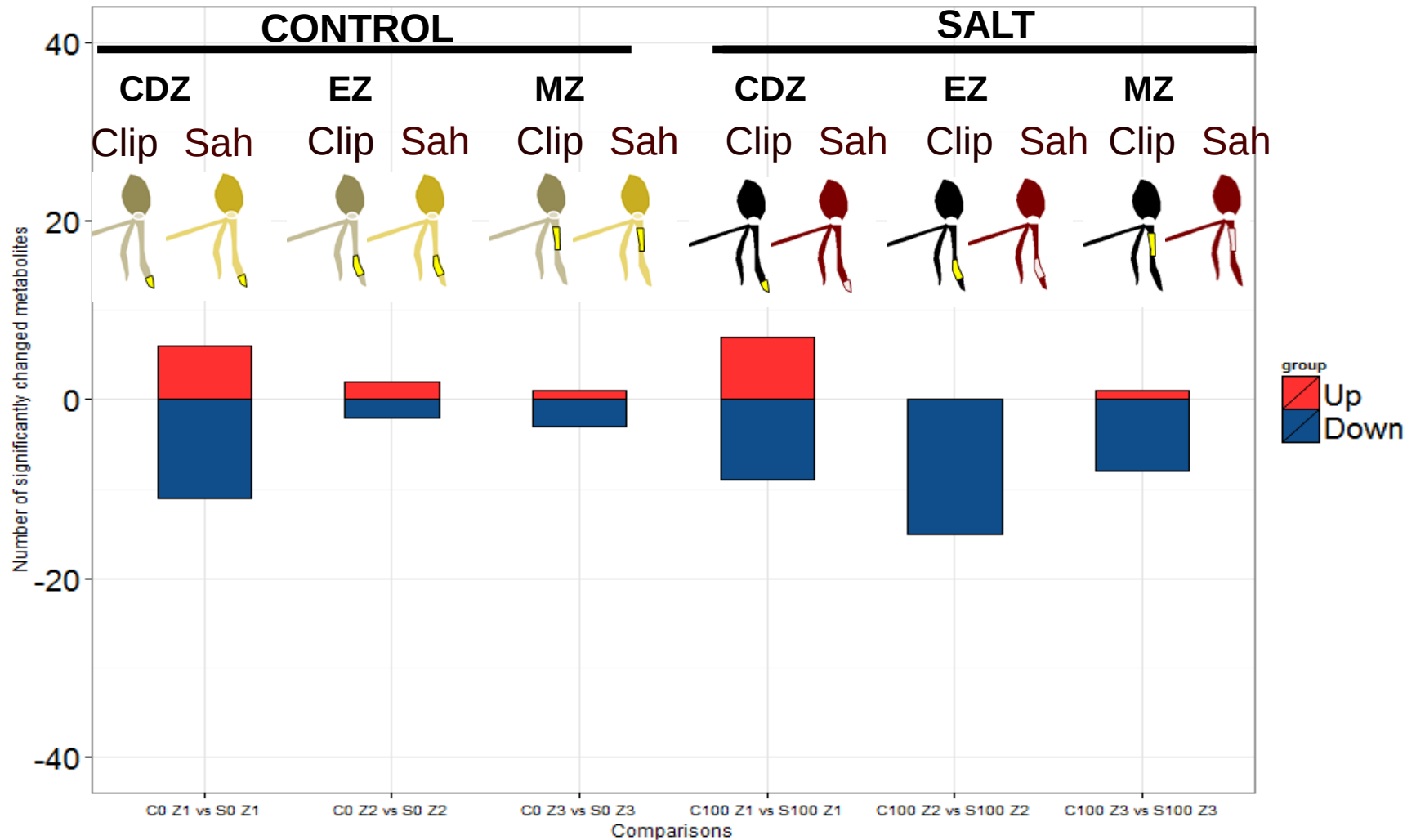
2. Targeted
Metabolite Profiling

3. RNA-Seq Analysis

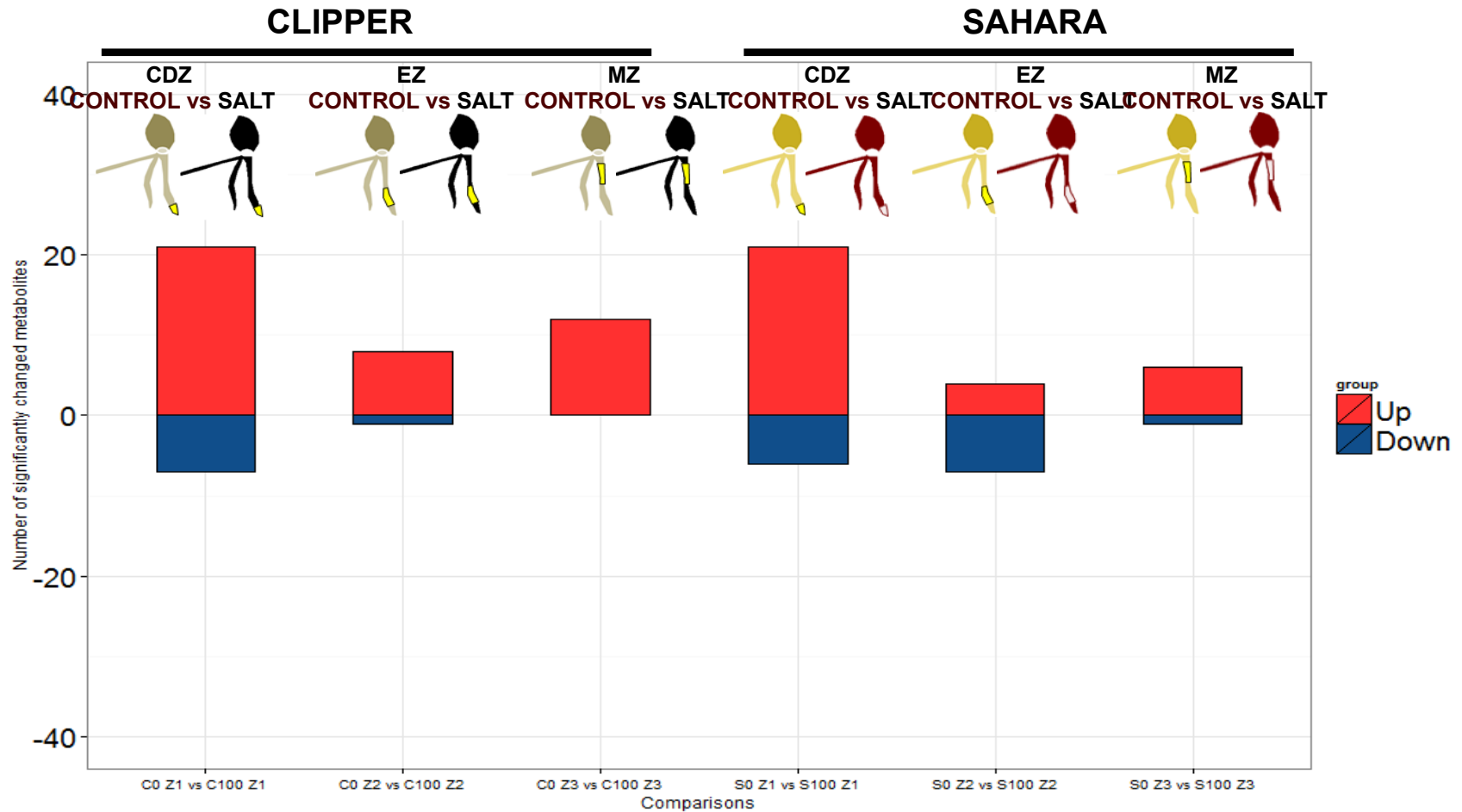
Data Analysis and Statistics

Data Integration and Biological Interpretation

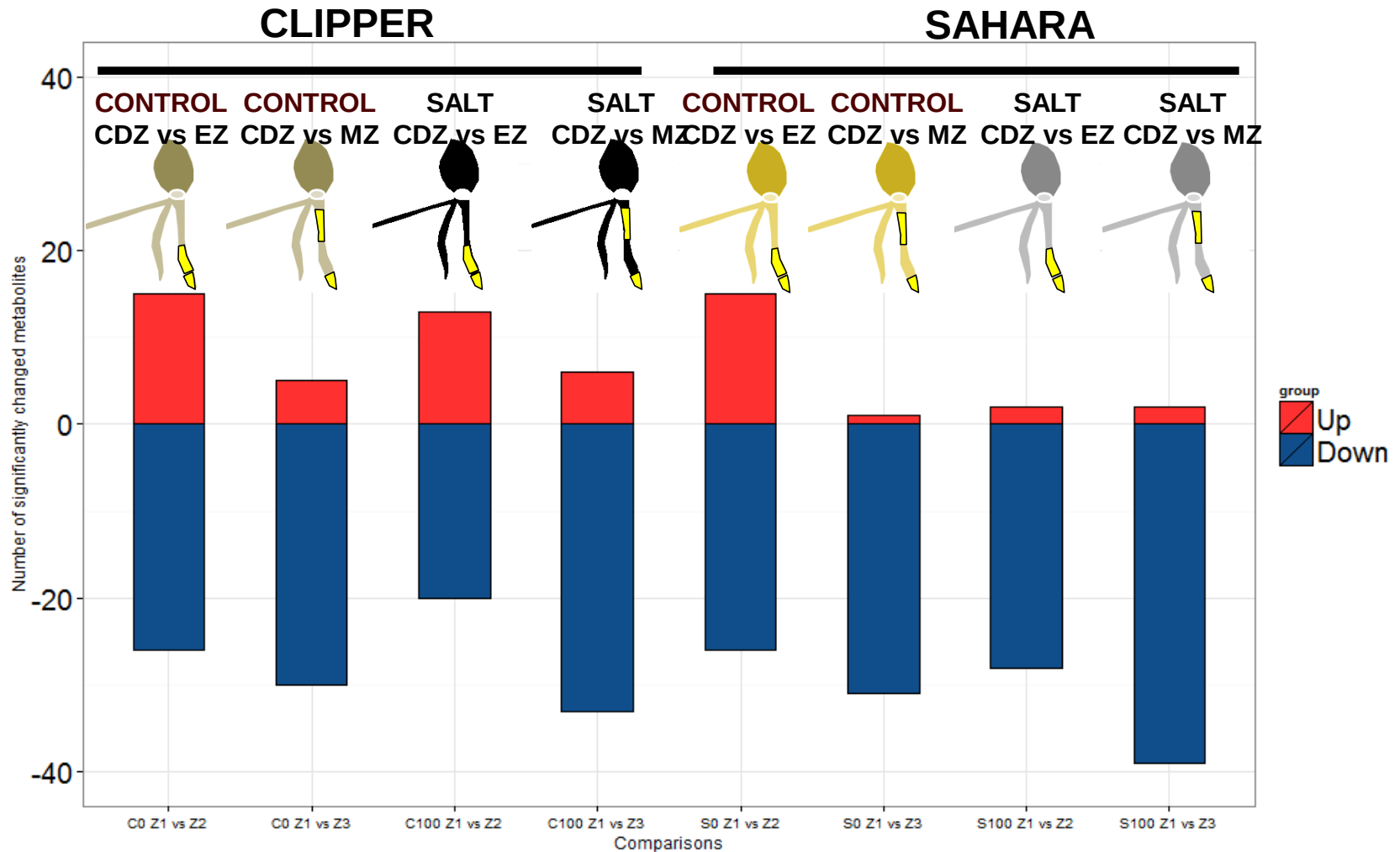
Significant differences (Sug, OA, AA) between cultivars



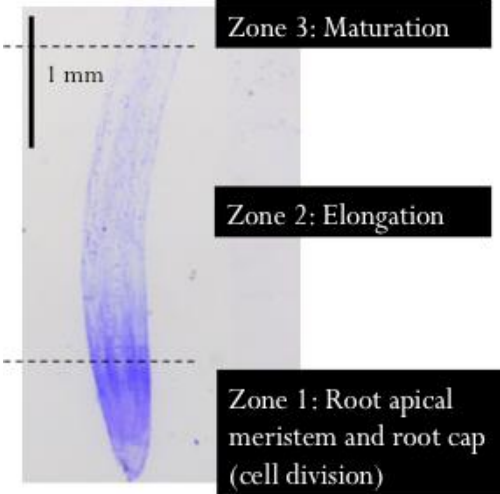
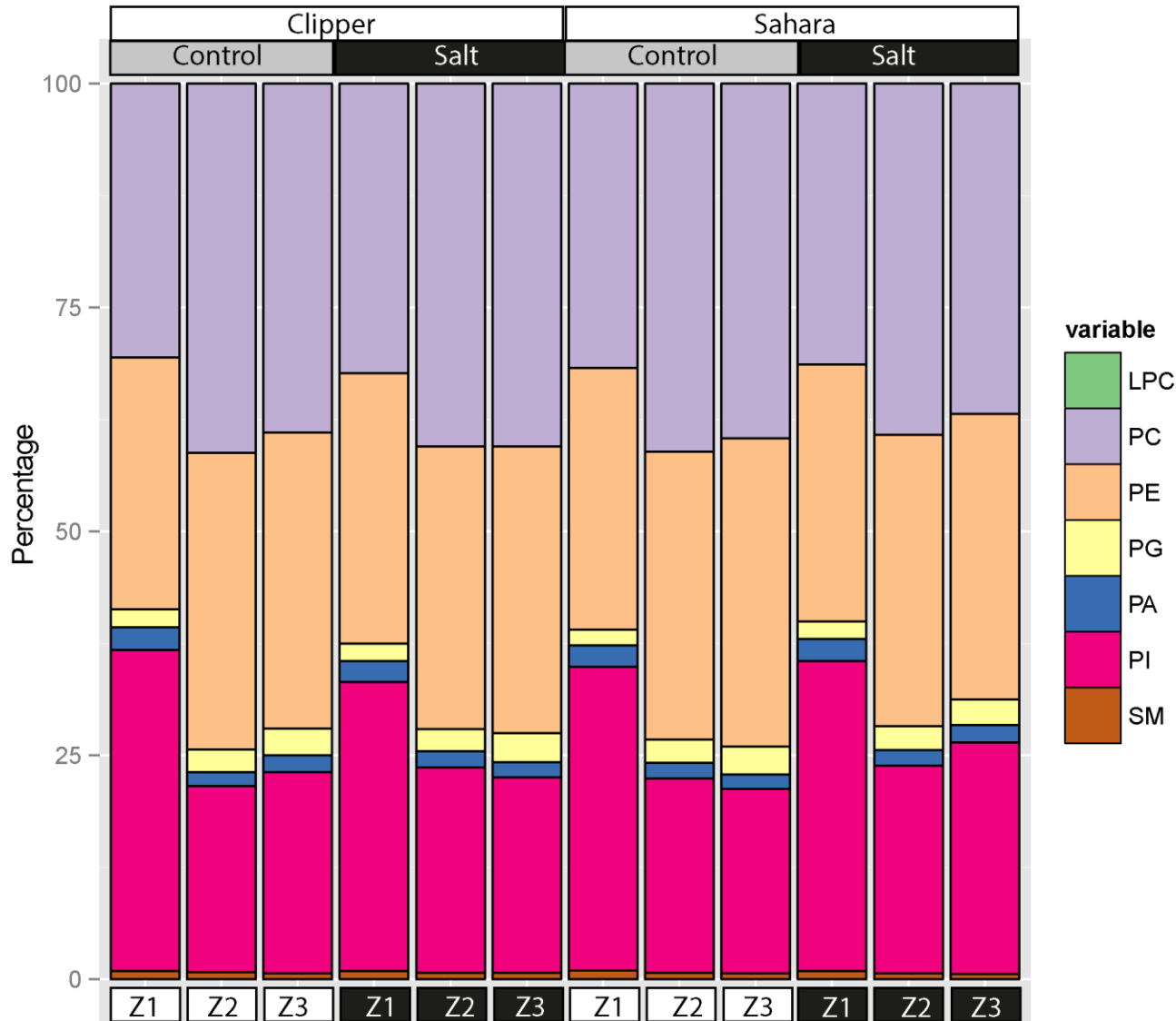
Significant changes (Sug, OA, AA) following treatment



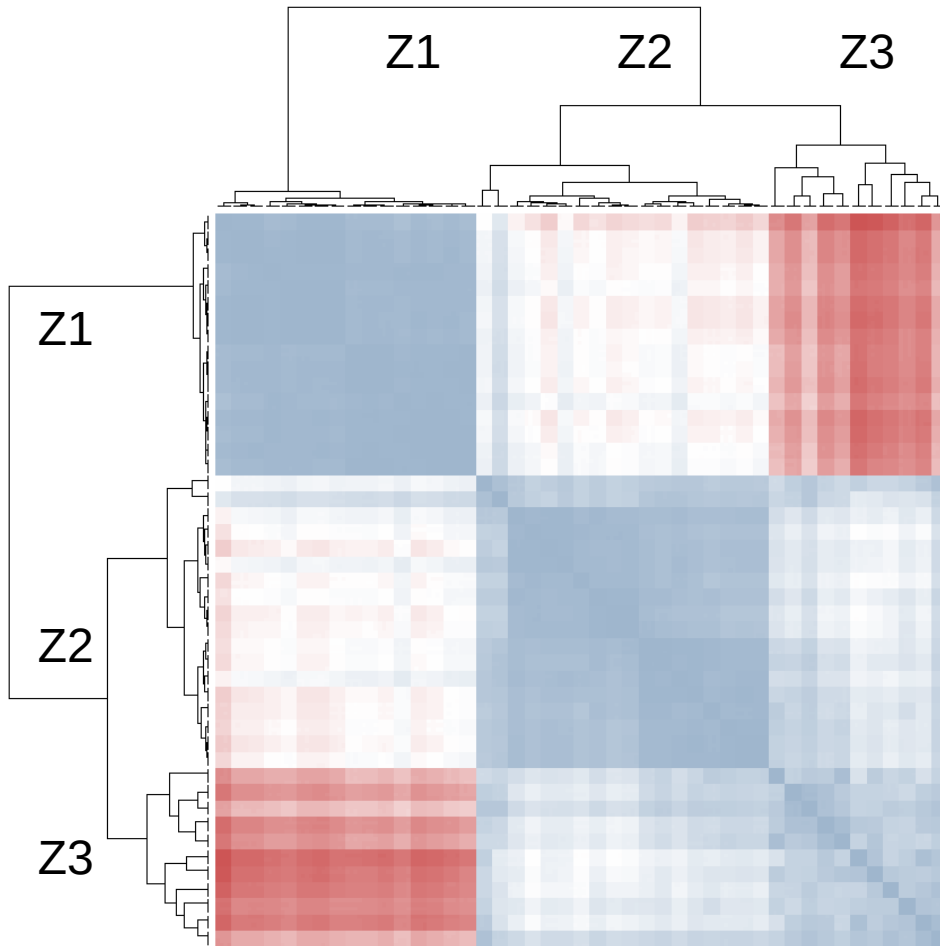
Significant differences (Sug, OA, AA) between zones



Tissue-specific lipid analysis



Next-Generation RNA Sequencing



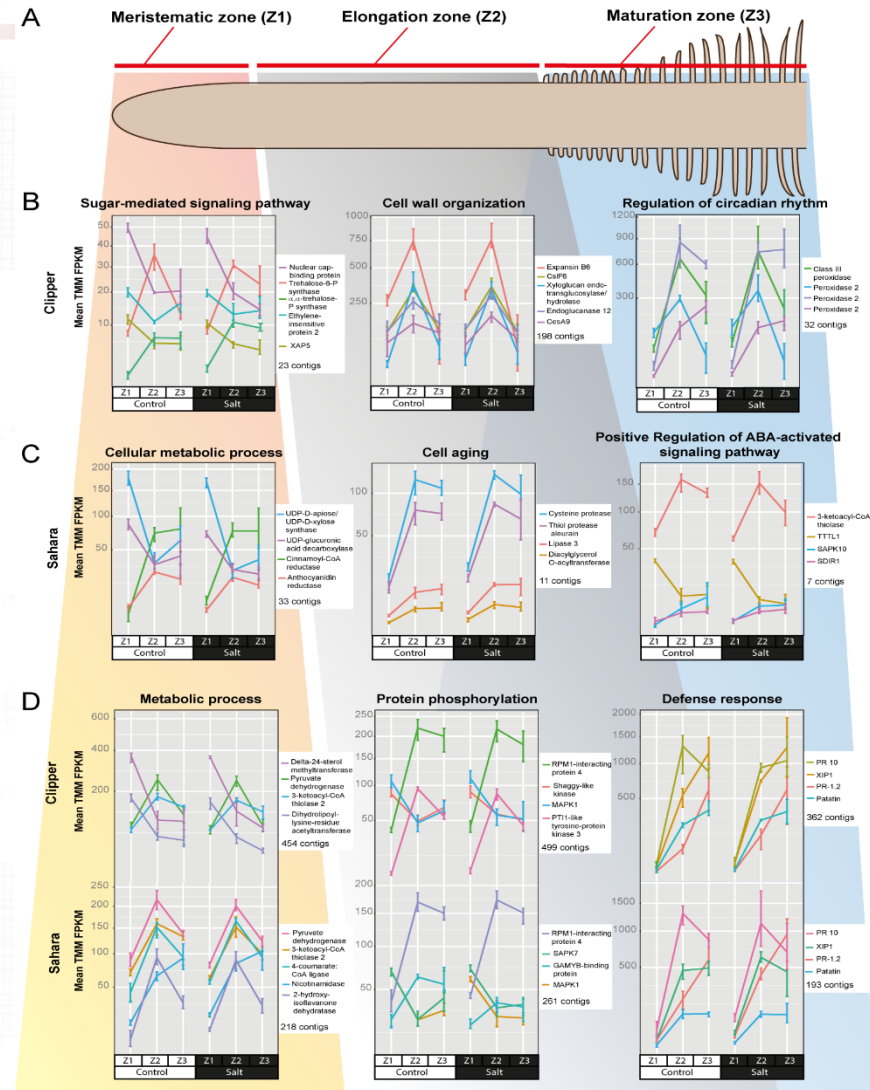
A clear per-zone (horizontal axis) response is observed in this Pearson's correlation plot illustrating cell differentiation along the root zone.

Transcriptomics analysis (short-term salt stress)



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- Root zones are highly specialized in their biological function
- Transcriptional differences are much larger between zones than between treatments.
- Biochemical processes are controlled in a root-zone specific manner



Hill, C. B., et al. (2016). Sci Rep **6**: 31558.

Significant metabolites - Clipper



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Z1 only

	DAM	FC		DAM	FC
Sugars (16)	Glucose 6-P	-3.57	Organic acids	Glucuronate	-1.85
	Maltose	+2.53		Nicotinic acid	-1.72
	Arabinose	-2.27		Quinate	-1.61
	Trehalose	-2.22		Caffeic acid	-1.59
	Raffinose	-1.89			
	Melibiose	-1.85	Amines (12)	Phenethylamine	-3.13
	Galactitol	-1.82		Serine	-2.86
	Xylitol	-1.79		Citrulline	-2.63
	Glucose	+1.78		Homoserine	-2.50
	Inositol	-1.75		Proline	+2.43
	Arabitol	-1.75		Glutamic acid	-2.22
	Fucose	-1.72		Glycine	-2.13
	Fructose 6-P	-1.67		Octopamine	-2.08
	Galactose	-1.64		Histidine	-2.04
	Turanose	-1.61		Aspartic acid	-2.00
	Xylose	-1.56		Ornithine	-1.89
				Agmatine	+1.65
Organic acids (14)	Fumarate	-4.00	Lipids (9)	PC (34:3)	+2.78
	Oxoglutarate	-3.23		LPC (18:3)	+2.34
	Succinate	-3.13		PG (32:0)	+2.48
	Malonate	-2.22		PC (34:1)	+2.18
	Salicylate	-2.08		PE (36:3)	+2.85
	2-keto gluconic acid	-2.00		PC (36:0)	+3.2
	Malate	-2.00		PC (38:5)	+2.18
	Aconitate	-1.96		PG (36:3)	-3.45
	Pipecolate	+1.92		LPC (18:1)	+1.87
	Citrate	-1.89			

Z1 or Z2 or Z3

	DAM	FC
N/A	N/A	N/A

Z1 or Z2

	DAM	(Z1) FC	(Z2) FC
Amines (2)	GABA	+2.24	+2.17
	Arginine	+1.71	+2.16
Lipids	PI (34:1)	-1.72	-2.44

Z2 only

	DAM	FC
Amines	Asparagine	+2.67
Fatty acids (3)	Elaidic acid (C18:1n9t)	+1.62
	Linoleic acid (C18:2n6c)	+1.60
	Palmitic acid (C16:0)	+1.45

Z2 or Z3

	DAM	(Z2) FC	(Z3) FC
Amines	Tyrosine	-2.22	-3.03
Fatty acids (3)	Nervonic acid (C24:1)	+1.29	-1.54
	Lauric acid (C12:0)	-1.25	-1.92
	Myristic acid (C14:0)	-1.19	-1.92

Z3 only

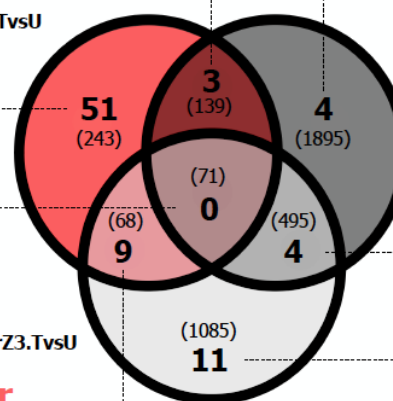
	DAM	FC
Sugars	Sucrose	+2.71
Amines (4)	Valine	-2.04
	Tryptophan	-1.92
	Isoleucine	-1.89
	beta-alanine	+1.84
Fatty acids (6)	Erucic acid (C22:1n9)	-2.22
	Heptadecanoic acid (C17:0)	-2.13
	cis-10-pentadecanoic acid (C15:1)	-1.79
	Stearic acid (C18:0)	-1.61
	Arachidic acid (C20:0)	-1.35
	Pentadecanoic acid (C15:0)	-1.32

DAM_ClipperZ1.TvsU

DAM_ClipperZ2.TvsU

DAM_ClipperZ3.TvsU

Clipper
(treatment-specific)



Z1 or Z3

	DAM	(Z1) FC	(Z3) FC		DAM	(Z1) FC	(Z3) FC
Amines (7)	Tyramine	-3.23	-2.50	Amines	Putrescine	-1.96	-4.55
	Glutamine	-2.94	-2.00		Leucine	-1.82	-3.12
	Threonine	-2.70	-2.56				
	Cysteine	-2.22	-1.82	Fatty acids (2)	Behenic acid (C22)	+2.65	+1.27
	Methionine	-2.00	-3.57		Lignoveric acid (C24:0)	+1.77	+1.43

Significant metabolites - Sahara



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Z1 only

	DAM	FC		DAM	FC
Sugars (2)	Arabinose	+3.63	Amines	Glutamine	-1.79
	Raffinose	+2.58		Homoserine	-1.75
Organic acids	Pipecolate	+4.89		Glycine	-1.75
				Phenylalanine	-1.59
				Threonine	-1.54
Amines (10)	Proline	+4.03	Lipids (3)	PA (39:0)	-1.41
	Ornithine	-2.70		PC (33:3)	-2.38
	Putrescine	-2.13		PC (36:6)	+2.09
	Citrulline	-2.08			
	4-hydroxyproline	+2.02			

Z1 or Z2

	DAM	(Z1) FC	(Z2) FC
Amines (8)	Tyramine	-2.44	+2.33
	Serine	-1.79	+5.07
	Glutaminic acid	-1.67	+4.11
	Arginine	+1.63	+1.93
	Taurine	-1.61	+4.64
	Octopamine	-1.59	+2.77
	Aspartic acid	-1.52	+2.34
	Cysteine	+1.44	-3.57
Lipids	PI (34:1)	-2.04	-3.57

Z2 only

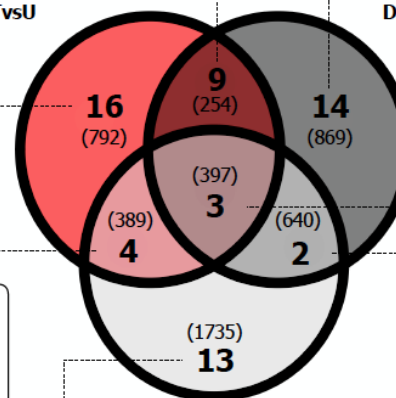
	DAM	FC		DAM	FC
Sugars	Sucrose	+4.76	Lipids (9)	PI (34:3)	+6.31
				PA (46:6)	+6.08
Amines (3)	Agmatine	+3.72		PC (38:5)	+4.88
	Phenethylamine	+3.62		PE (34:3)	+4.33
	Lysine	+1.92		SM (41:1)	+3.94
Fatty acids				PE (32:2)	+3.73
	Stearic acid (C18:0)	-4.00		PC (34:1)	+3.63
				PG (32:0)	+3.08
				LPC (18:3)	+3.07

Z1 or Z3

	DAM	(Z1) FC	(Z3) FC
Amines (4)	Alanine	+1.69	+2.49
	Valine	+1.55	-2.63
	Tryptophan	-1.56	-2.94
	Histidine	-1.52	-2.78

DAM_SaharaZ1.TvsU

DAM_SaharaZ2.TvsU



DAM_SaharaZ3.TvsU

Z1 or Z2 or Z3

	DAM	(Z1) FC	(Z2) FC	(Z3) FC
AmineAA (3)	Tyrosine	-2.33	-3.57	-2.08
	Methionine	-1.45	-4.76	-4.76
	beta-alanine	+1.40	+6.56	+2.48

Z2 or Z3

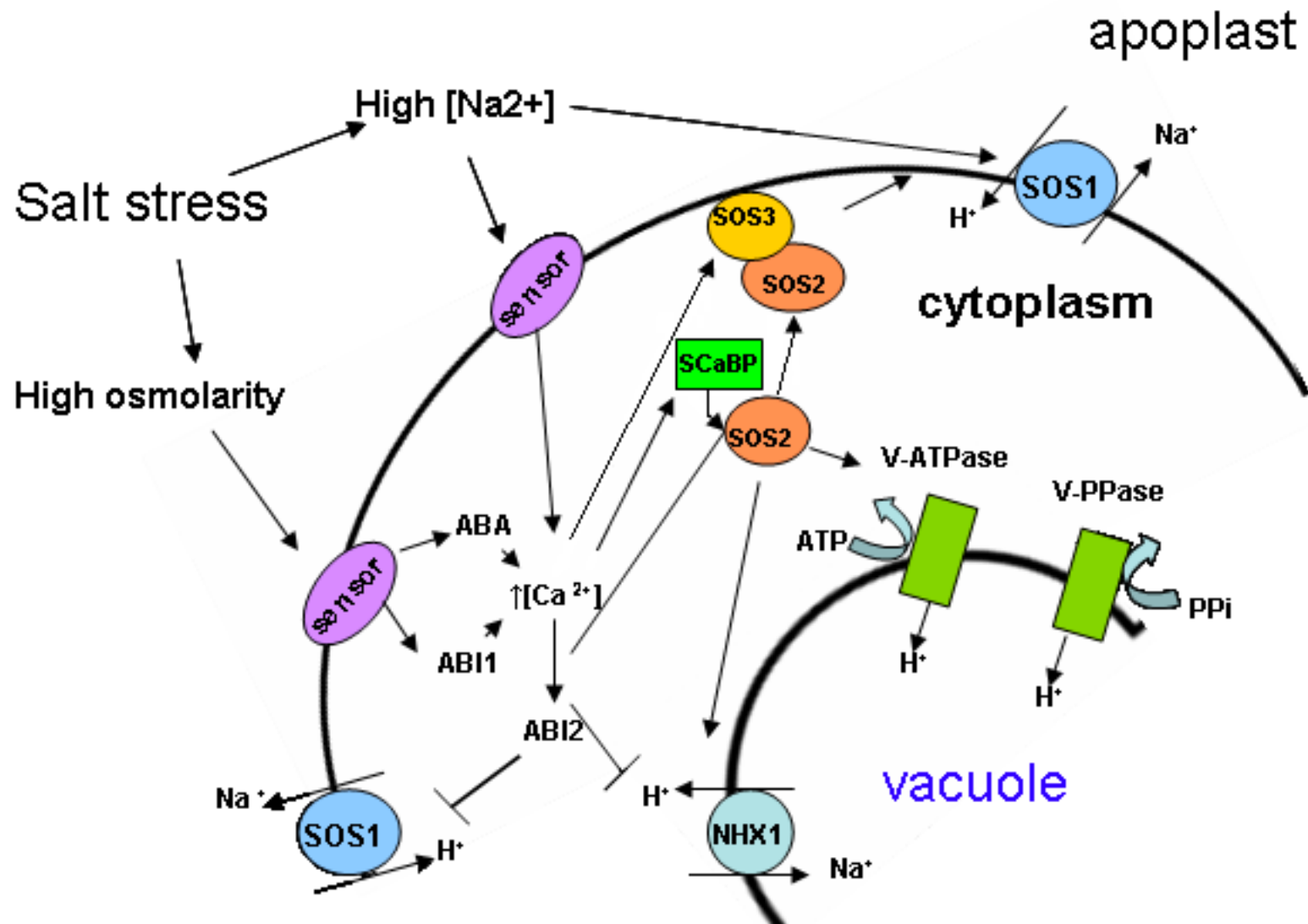
	DAM	(Z2) FC	(Z3) FC
FattyAcids	Nervonic acid (C24:1)	-2.50	-1.69
Lipids	PI (36:4)	-3.45	-2.38

Z3 only

	DAM	FC		DAM	FC
Organic acids	Malate	-3.57	Fatty acids	Lignoveric acid (C24:0)	+1.39
				Elaidic acid (C18:1n9t)	+1.40
Amines	Leucine	-2.13		Behenic acid (C22)	+1.59
				Palmitic acid (C16:0)	+1.26
				Pentadecanoic acid (C15:0)	-1.25
Fatty acids (10)	Heptadecanoic acid (C17:0)	-1.75	Lipids	PI (36:5)	-3.03
	Erucic acid (C22:1n9)	-1.64			
	Myristic acid (C14:0)	-1.49			
	cis-10-pentadecanoic acid (C15:1)	-1.47			
	Lauric acid (C12:0)	-1.49			

Sahara
(treatment-specific)

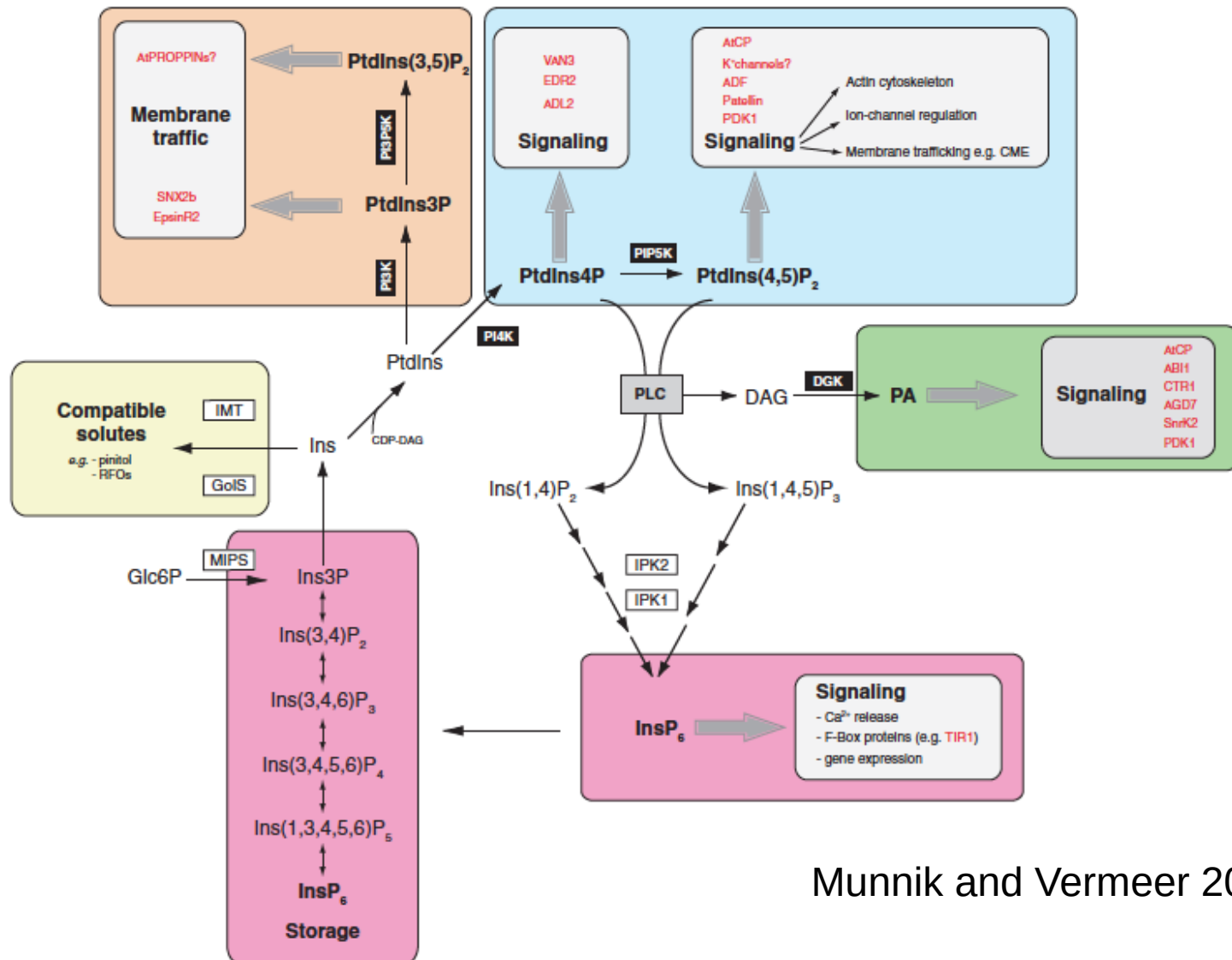
Salt signalling



Proposed lipid based signalling

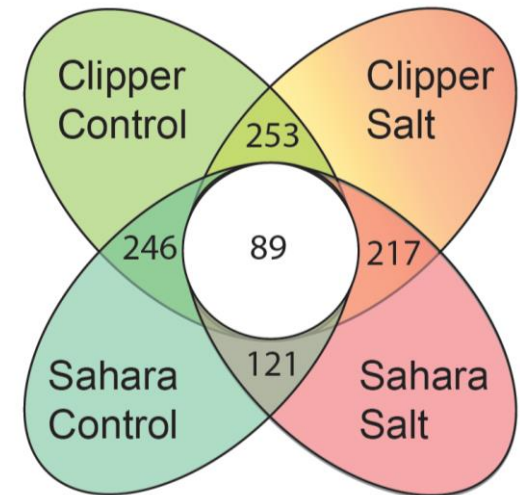
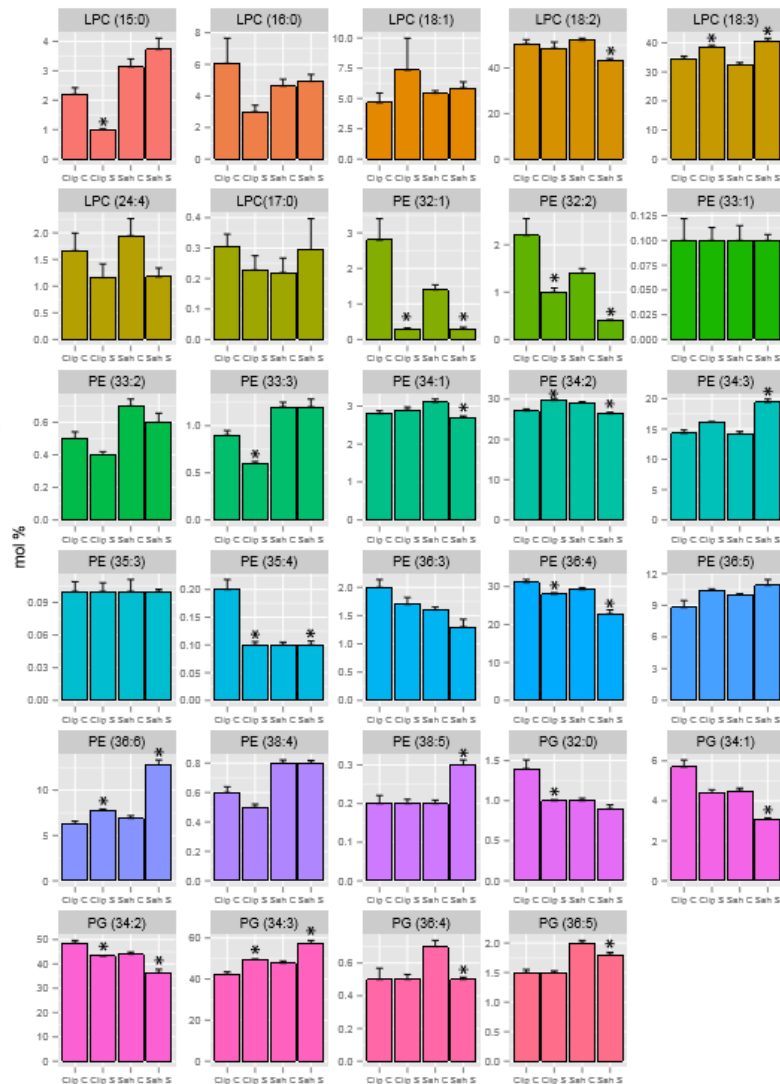
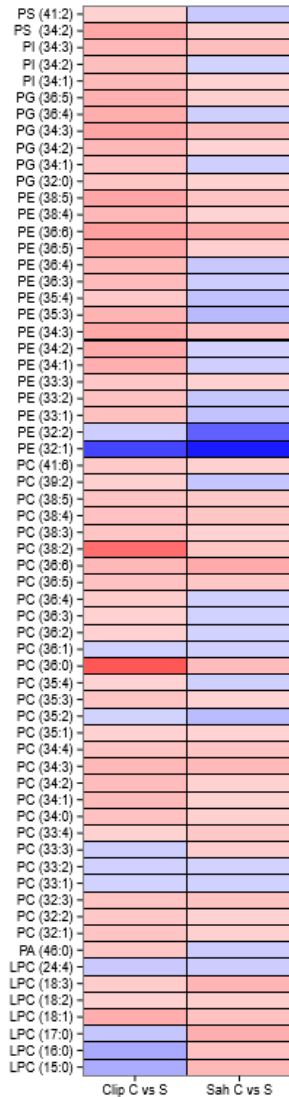


METABOLOMICS
AUSTRALIA



Munnik and Vermeer 2010

Total lipid analysis in salt treated roots



The barley root lipidome

Gradient Category and Source Polarity

Gradient 1/Neg

PC

PE

PG

PS

PI

SQDG

DGDG

MGDG

Cardiolipin

LysoGlycerolipid

PA

Gradient 2/Pos

Ceramide

HexCer

DAG

ASG

SG

Gradient 3/Pos

GIPC

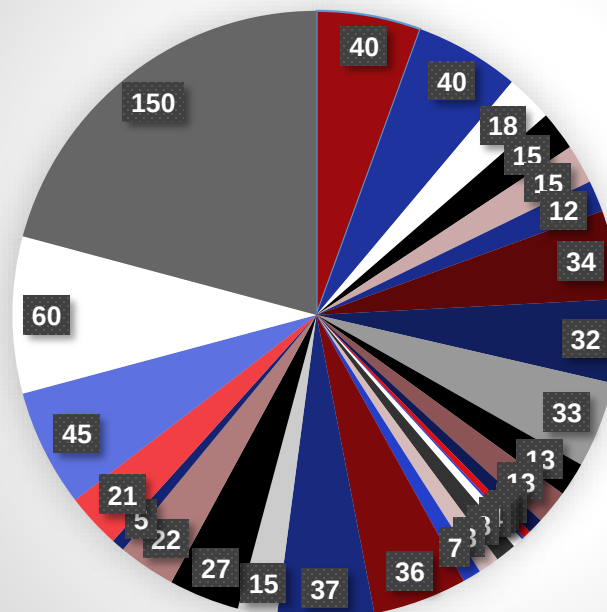
OHC

Gradient 4/Pos

TAG

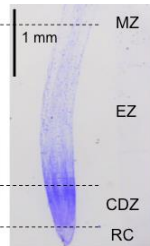
SE

Number of lipid species identified

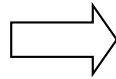


Yu et al 2018

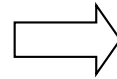
Where to next?



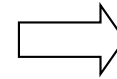
tissue harvest



shock freeze

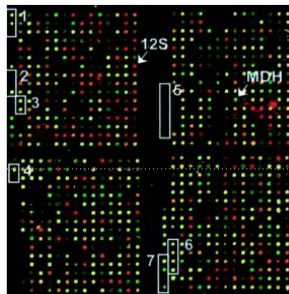


homogenization

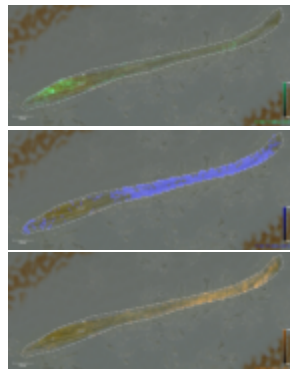
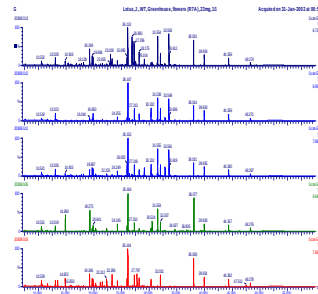


extraction

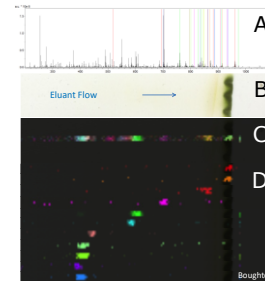
Transcriptomics



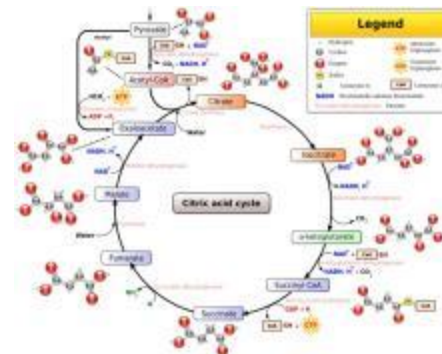
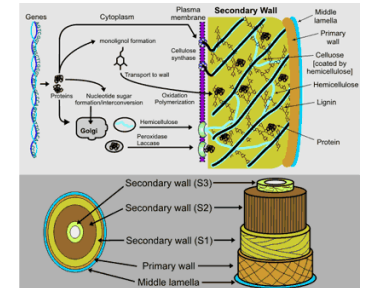
Metabolomics



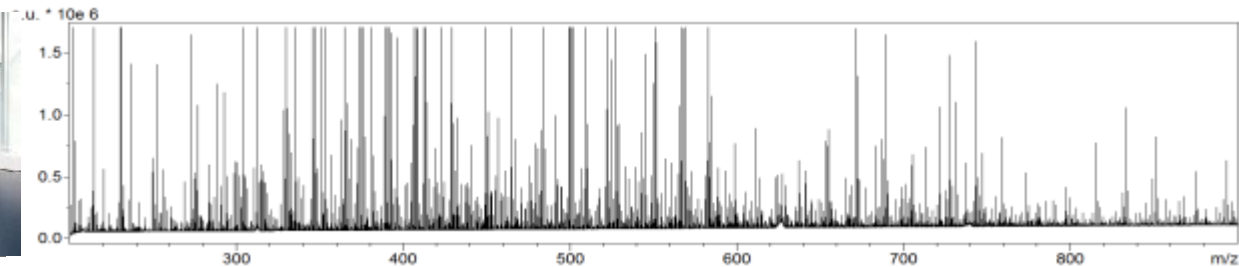
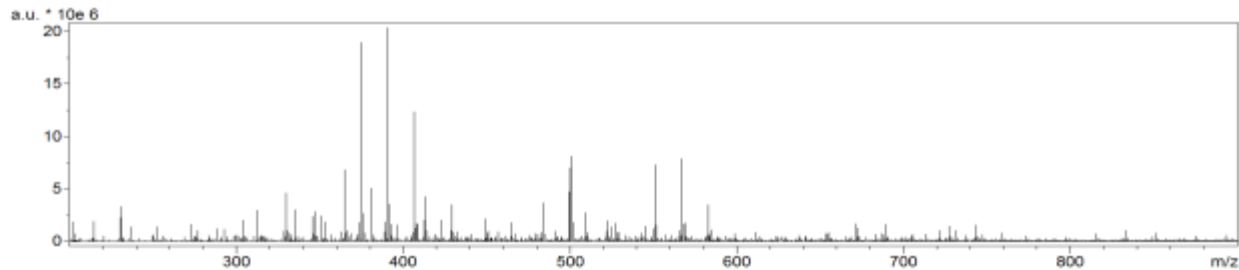
Lipidomics



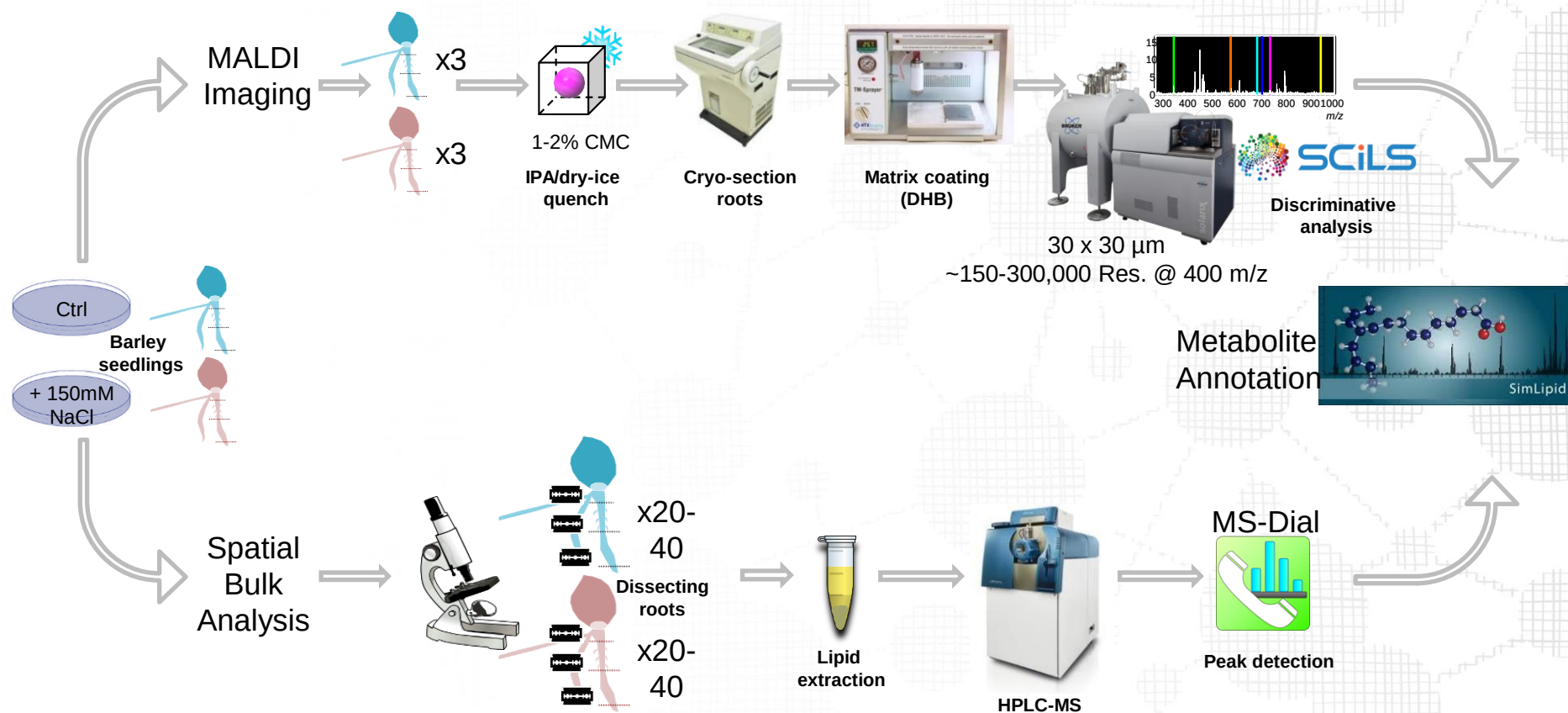
Cell wall



MALDI-IMS of barley roots



Workflow

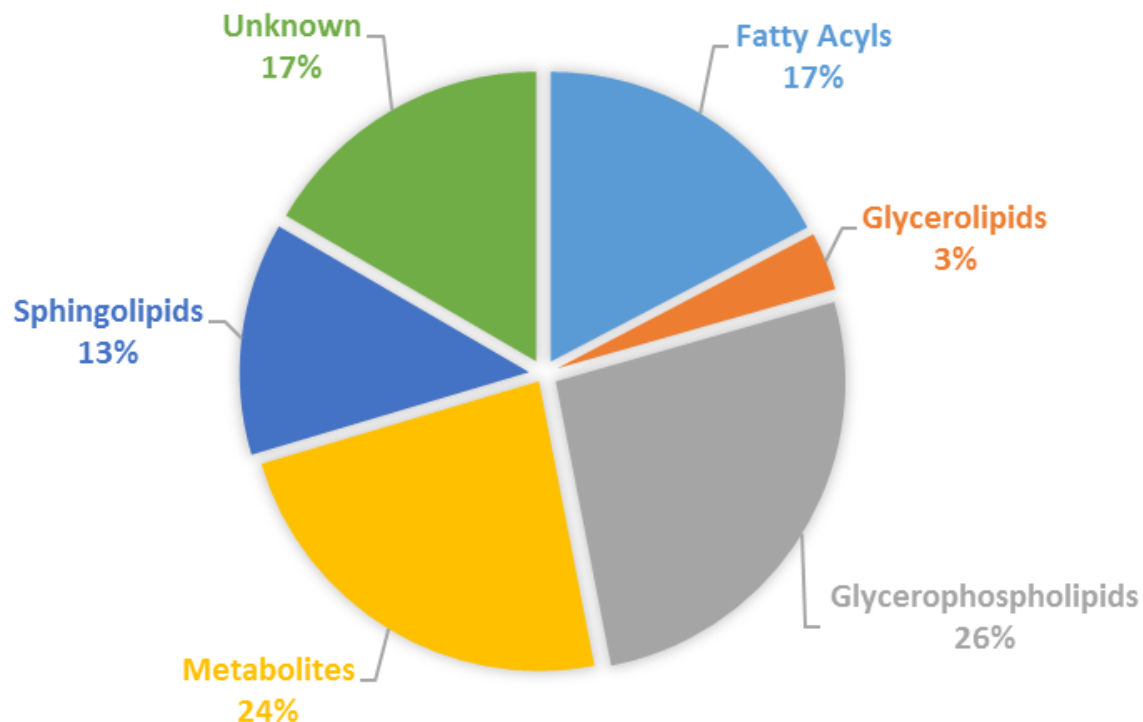


Tentative Annotations via MSI

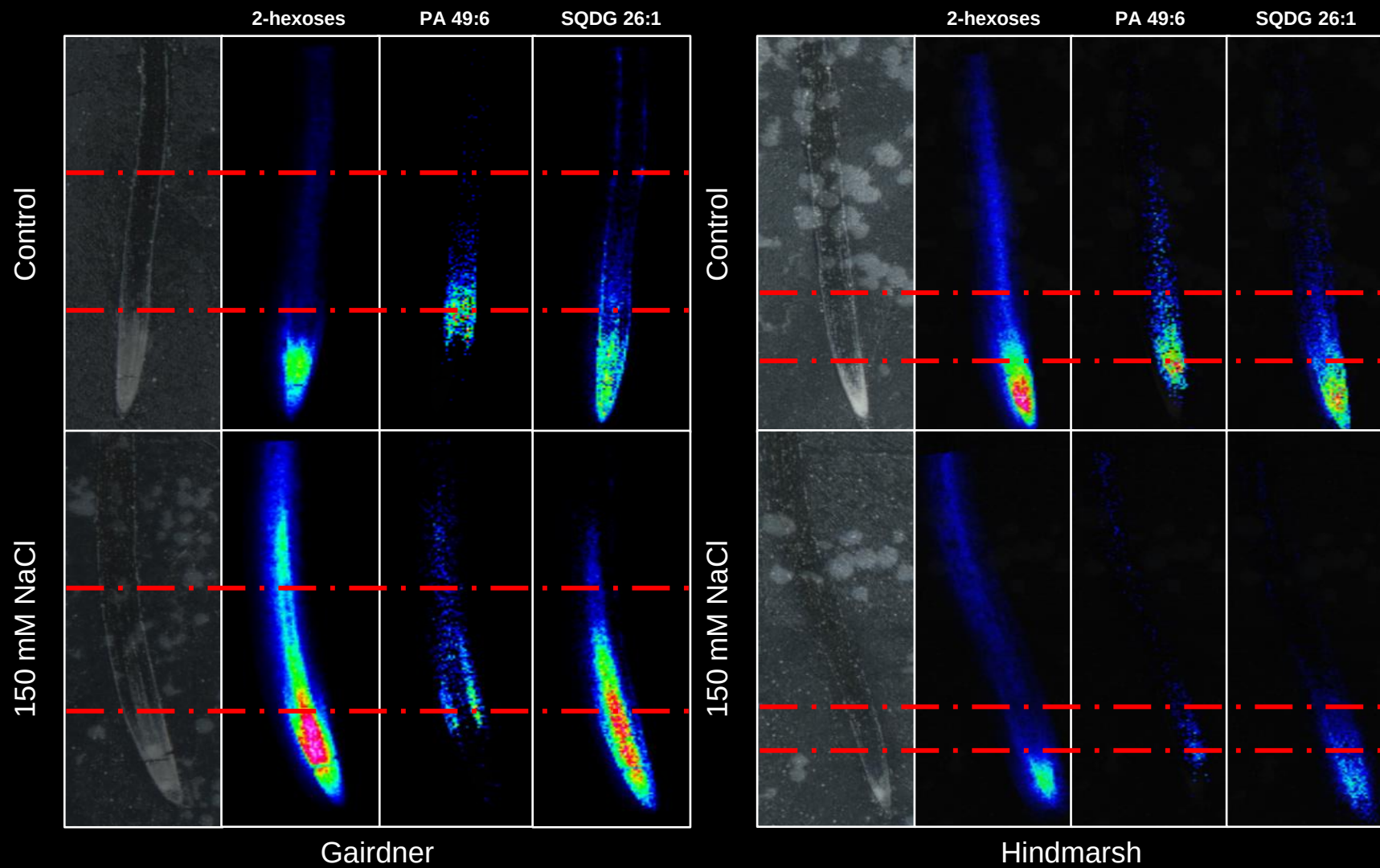


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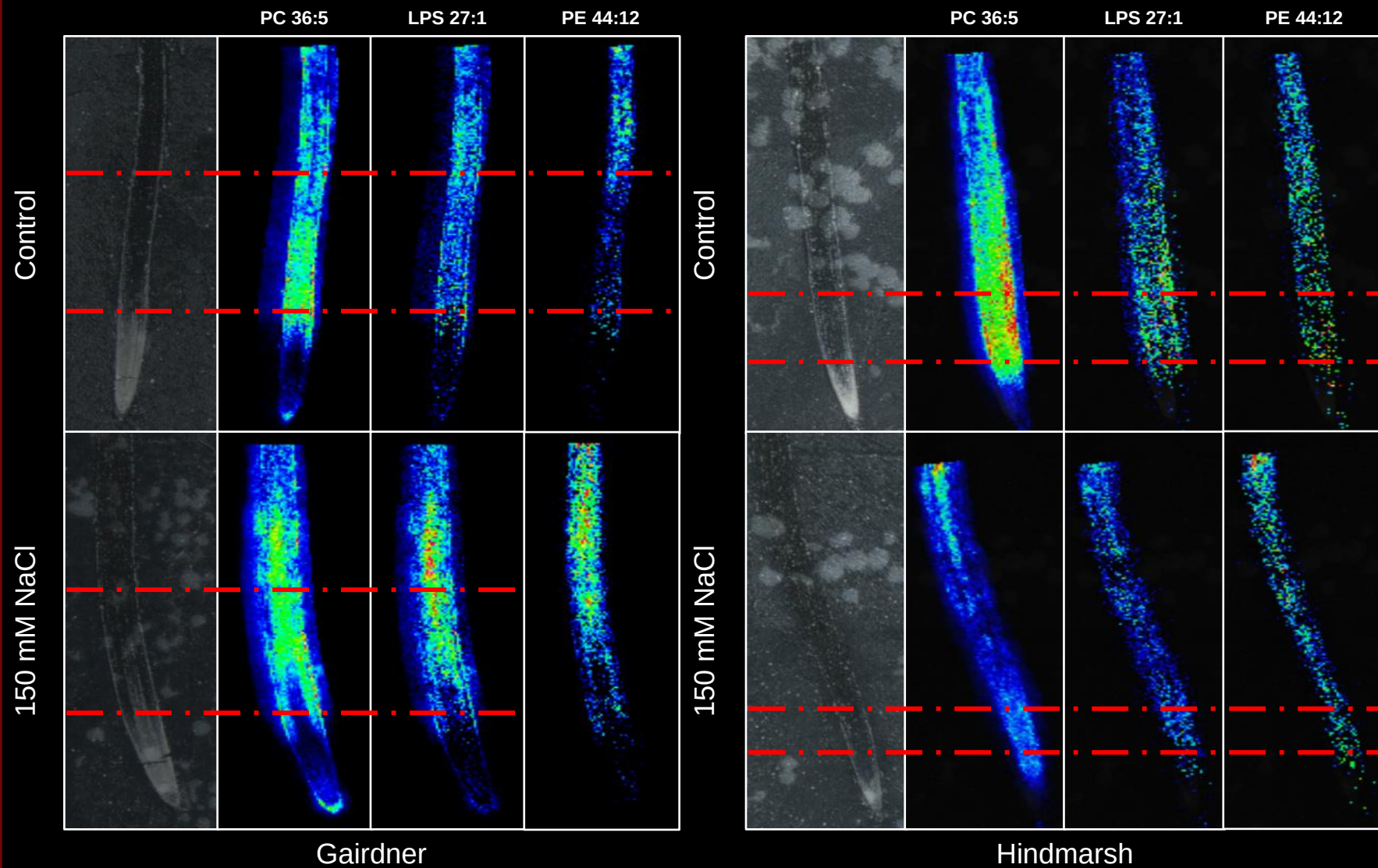
- **230** spatially distributed ions across different barley cultivars (Gairdner and Hindmarsh)
- Annotated using SimLipid and Metlin (5 ppm), and curated for multiple adducts & isobars



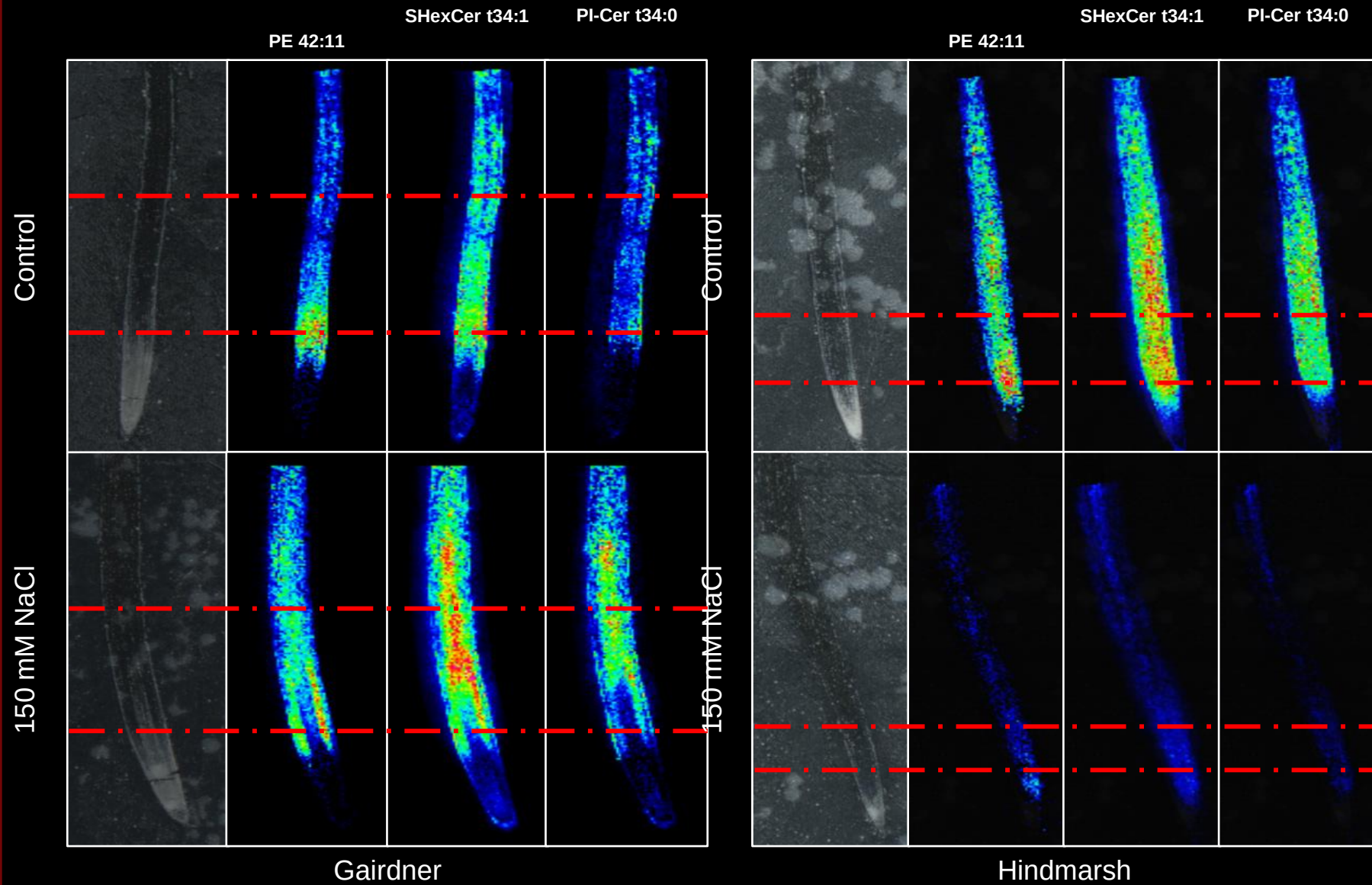
Results



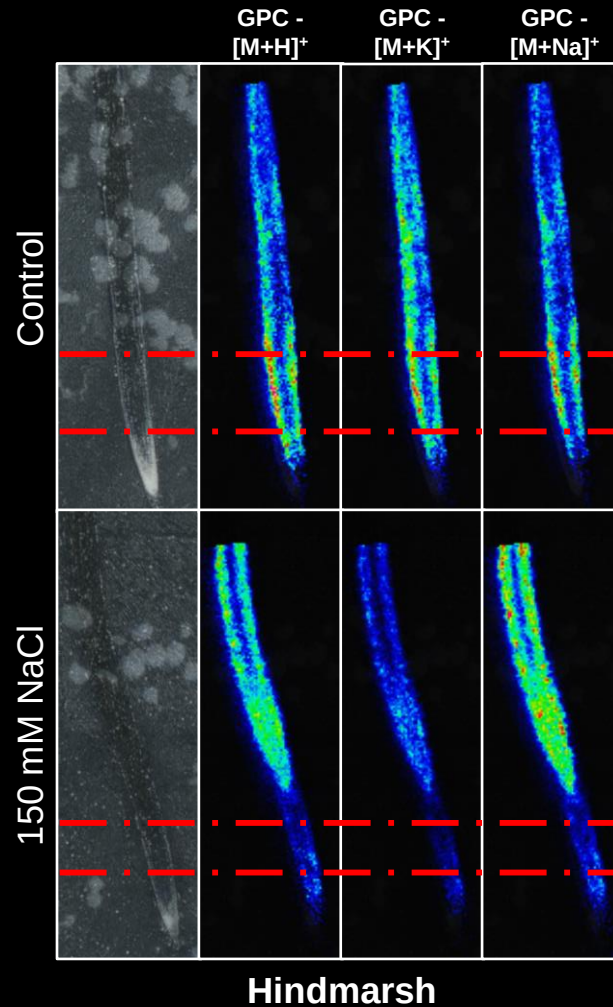
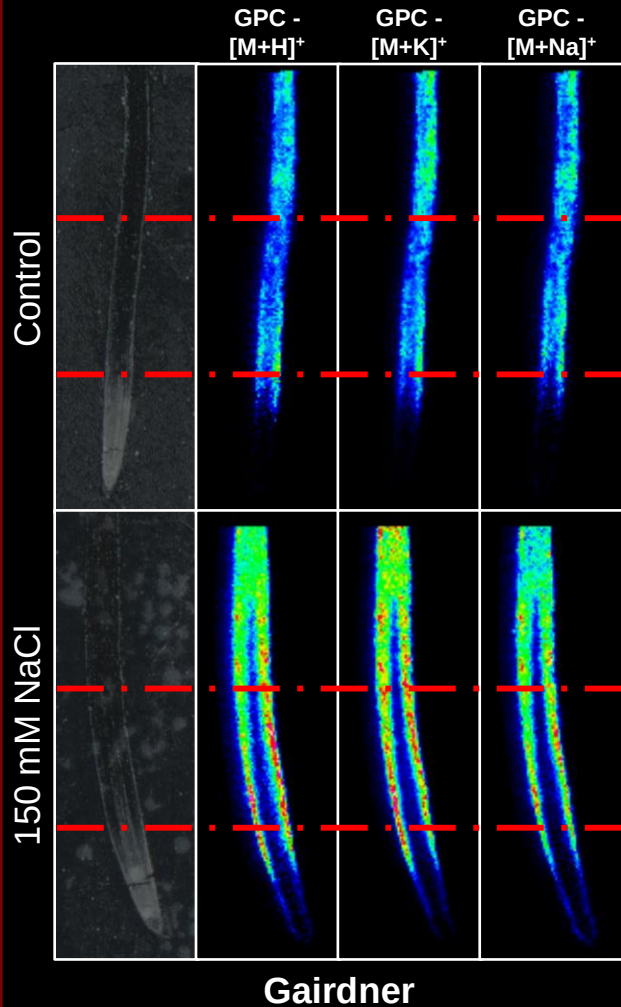
Results



Results



Results

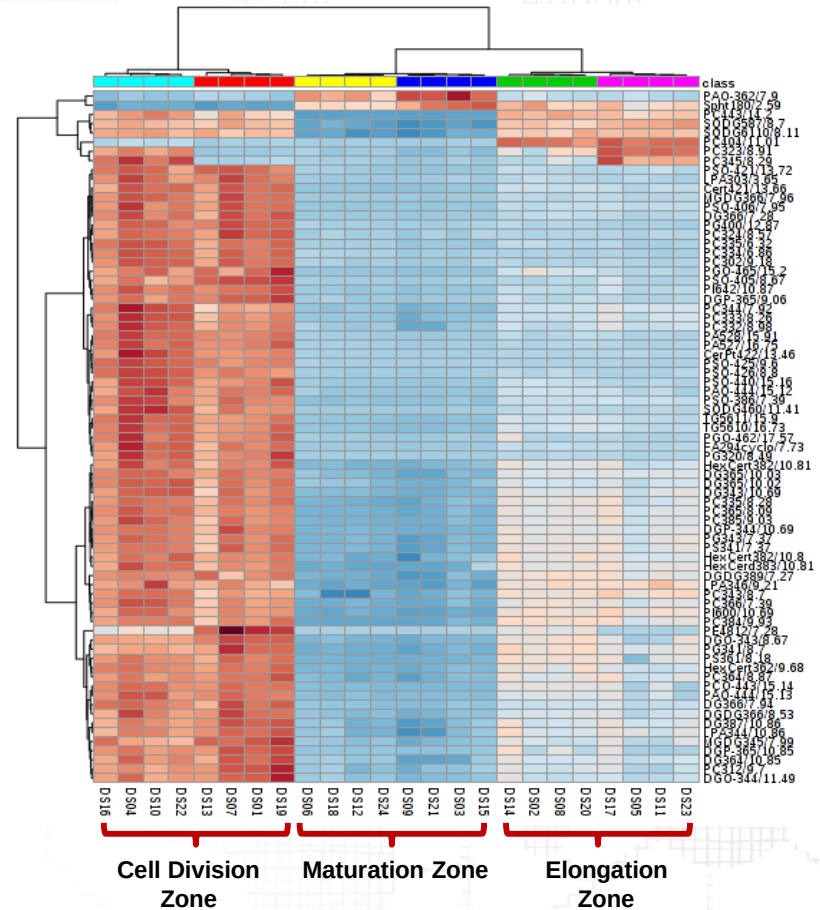
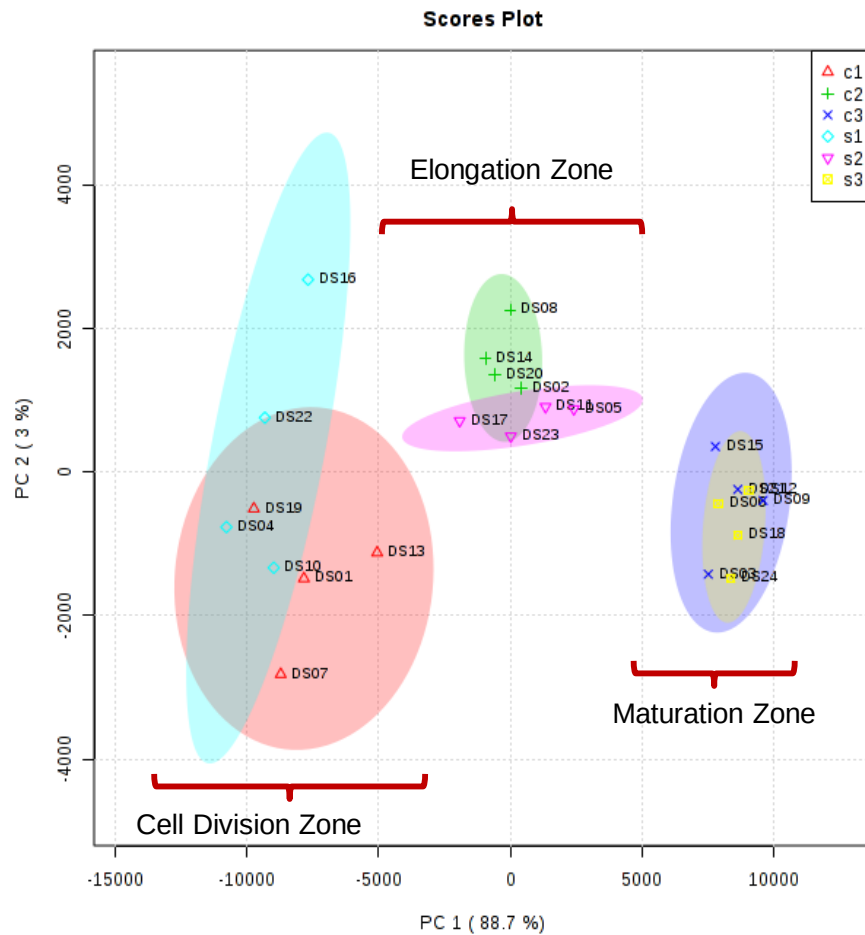


- GPC is a degradation product of phosphatidylcholine, resulting from the removal of fatty acids by the activity of phospholipase A1, phospholipase A2 and lysophospholipase (Sahsah et al., 1998).
- The presence of GPC is generally interpreted as a stress-induced membrane turnover or degradation (Aubert et al., 1996).

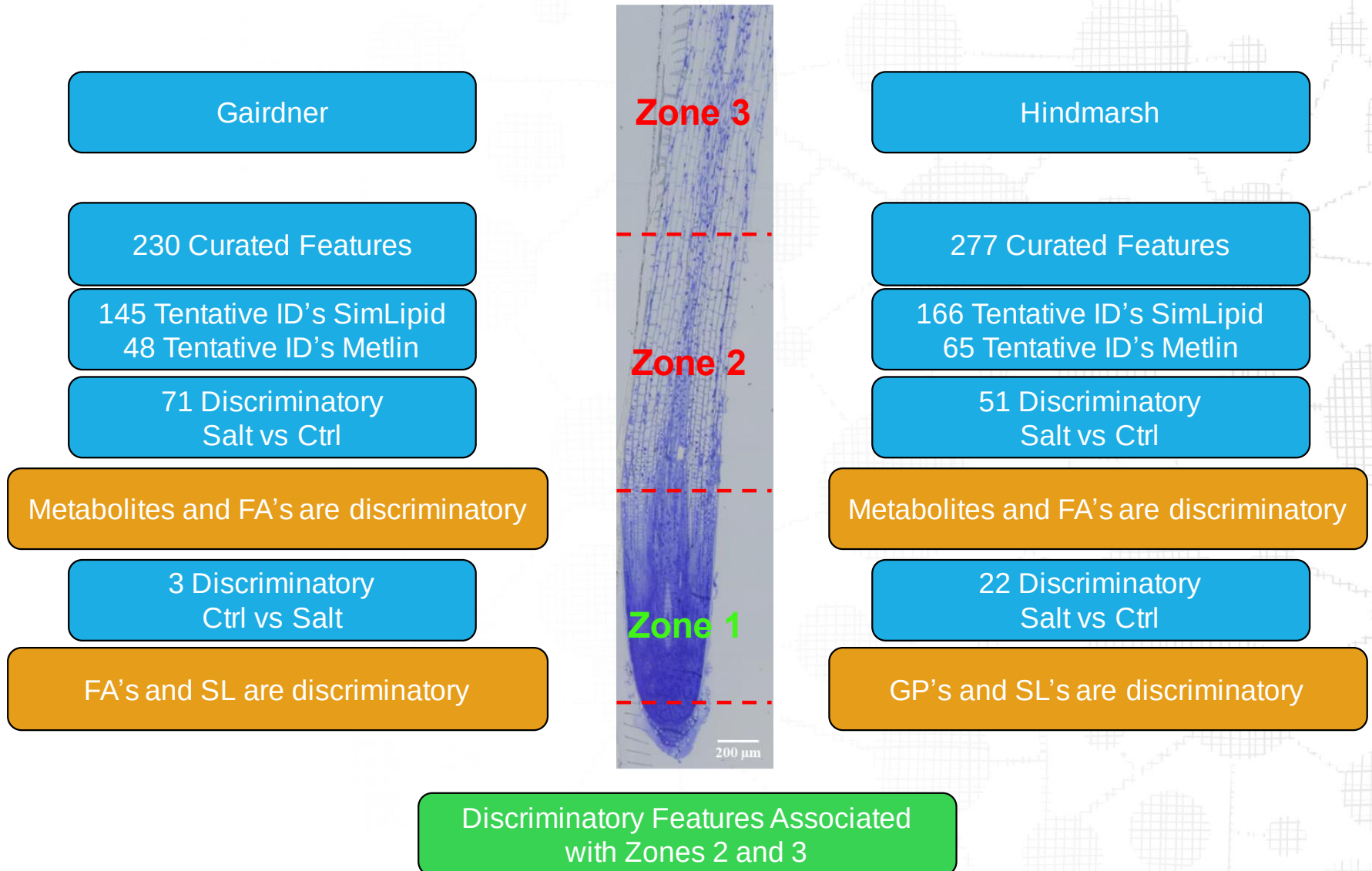
Spatial Bulk Analysis – LC-MS



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Summary

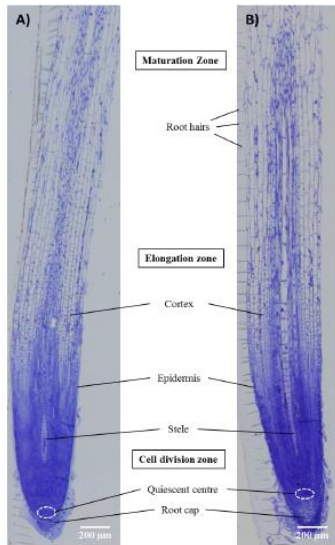


Summary

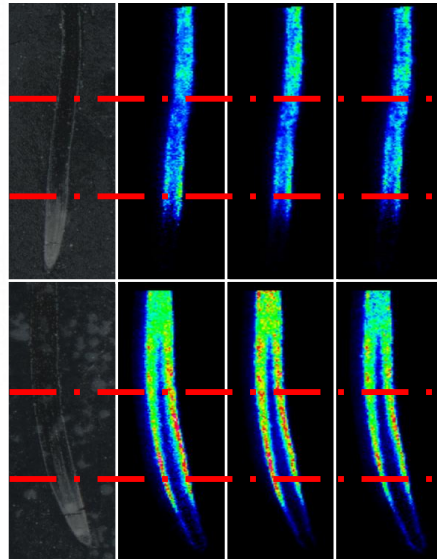
- Tissue type-resolved analyses demonstrate tissue-specificity of metabolism
- Large differences between genotypes of the same species
- Lipids are important players in salinity response
- Imaging MS provides new means of tissue-specific metabolite analysis

Future for spatial analysis

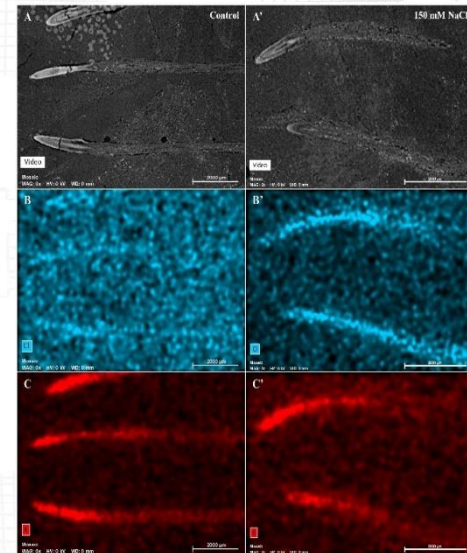
Microscopy



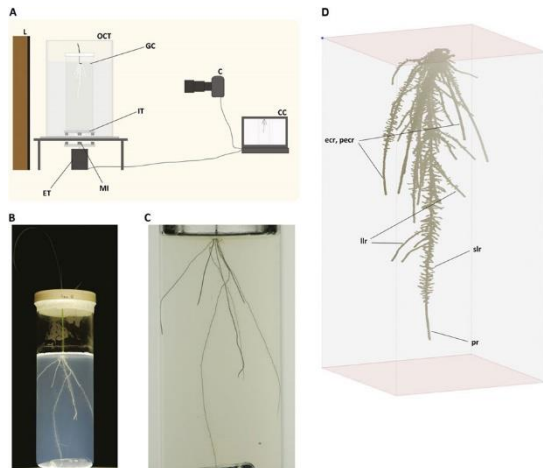
Imaging MS



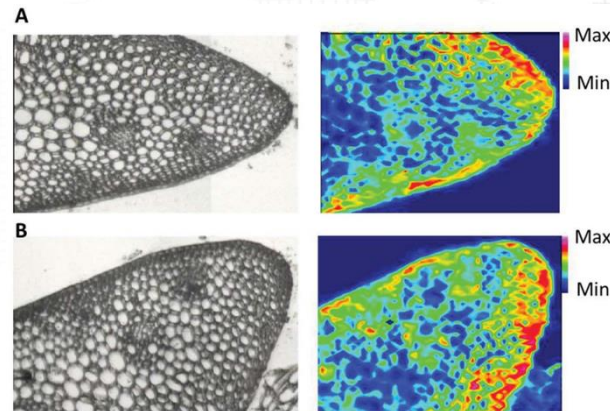
Micro-XRF



Root phenotyping



FT-IR



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