# **GC Method Development**



### What to Consider

The Sample

Method of injection

Inlet

**Detector** 

**Carrier Gas** 

Column

## **COMPOUND REQUIREMENTS FOR GC**

Only 10-20% of all compounds are suitable for GC analysis

### The compounds must have:

- ✓ Sufficient volatility
- ✓ Thermal stability

NO Inorganic Acids and Bases

Be mindful of salts!

# **Sample Considerations**

Sample matrixresidues?dirty samples?

### 2. Analyte Composition

- 1. Isomers?
- 2. Polar vs. non-Polar?
- 3. Organinc Acids?
- 4. Light Gases?
- 5. Nobel Gases?
- 6. Halogens?

# **Sample Residues**

Semi-volatile residues

Bake out

Back flush

Non-volatile residues

Guard column

Bake out

Back flush

**Dirty Samples** 

Sample clean up?

Back flush

# **Use What You Know About the Analytes**

Complex Mixture?

Few analytes?

Homologous Series?

Mixture of polar and non-polar?

Labile analytes?

Volatility?

Gas or Liquid Sample?

# We have a thought about the sample ...What's next?

# Let's Get the Sample Onto the Column...

Manual Injection

Liquid Injection

Headspace

Purge & Trap

Gas Sampling Valve

SPME

**Thermal Desorption** 

Custom

### The Inlet

Volatiles Interface

Cool-On-Column

**Purged Packed** 

PTV

Split / Splitless

Multi-Mode

### **Volatiles Interface**

Used for 'volatile' samples

Sample is already a vapor

Headspace

Purge & Trap

## **Volatiles Interface**

Mode	Sample Concentration	Sample to Column	Comments
Split	High	Very little, most is vented	
Splitless	Low	All	Can switch to split mode electronically
Direct	Low	All	Must physically disconnect split vent, plug the interface, and reconfigure the GC. Maximizes sample recovery and eliminates possibility of contamination to pneumatic system.

### Cool-On-Column

Good for Labile Samples

Sample is deposited "ON" the column

Temperature of inlet follows Oven Temperature

Good for 'Active' analytes

Minimizes inlet discrimination No inlet Liner\*

- Good for Trace Analysis
- High Boiling Point Compounds

No Inlet Discrimination

Guard Column Highly Recommended

# **Purged Packed**

Good for HIGH flow applications

Used with Packed columns

Can be used with 0.53 mm and 0.32 mm ID columns

\*\*Has a minimal capacity for sample expansion

\*\*Back Flash\*\*

# PTV (Programmable Temperature Vaporization)

Mode	Sample Concentration	Sample to Column	Comments
Split	High	Very Little	
Pulsed Split	High	Very Little	
Splitless	Low	All	
Pulsed Splitless	Low	All	
Solvent Vent	Low	All	Multiple injections concentrate analytes and vent solvent.

# **Split / Splitless**

Mode	Sample Concentration	Sample to Column	Comments
Split	High	Very Little	
Pulsed Split	High	Very Little	Useful with large injections
Splitless	Low	All	
Pulsed Splitless	Low	All	Useful with large injections. *better transfer of sample to column*

### **SPLIT INJECTOR**

### **Split Ratio**

- Too low: Poor peak shape
   Column overload
- Too high: Poor sensitivity
   -Wastes carrier gas (gas saver)
- Usually non-linear
   <u>Do not</u> use ratio as a dilution factor

### MINIMUM RECOMMENDED SPLIT RATIO

	mm I.D.	Lowest ratio
Hig	0.10	1:50 - 1:75
her f	0.18 - 0.25	1:10 - 1:20
low r	0.10 0.18 - 0.25 0.32 0.53	1:8 - 1:15
ates	0.53	1:2 - 1:5

\*Want to have 20 mL/min flow through the inlet\*

# **Multimode**

Mode	Sample Concentration	Sample to Column	Discussion
Split	High	Low	
Pulsed Split	High	Low	
Splitless	Low	All	
Pulsed Splitlss	Low	All	
Solvent Vent	Low	All	Multiple Injections concentrate sample and vent solvent
Direct	Low	All	



# **Sample Expansion...Liners?**

Split / Splitless Inlet

Multimode Inlet

Packed inlet

PTV

# **Inlet Liners - Purpose**

Glass Inlet Liners provide an "inert" space for liquid samples to be uniformly vaporized to a gas and moved to the column.

Liquid-gas phase change involves a significant change in volume.

Gaseous sample volume depends on

- the solvent type
- column head pressure
- temperature of inlet

These aspects should be optimized for your sample volume and application.

Solvent	Volume
(1µL, ambient)	(µL at 250°C and 20psig)
n-Hexane	140
Acetone	245
Acetonitrile	350
Methanol	450
Water	1010

See "A Practical Guide to the Care, Maintenance, and Troubleshooting of Capillary GC Systems", Third Revised Edition, by Dean Rood, Wiley-VCH, New York, 2001.

# **Liners - 3 Key Aspects Govern Applications**

**Liner Volume** 

**Liner Treatments or Deactivation** 

Special Characteristics (glass wool, cup, taper, etc.)

When choosing a liner for your application, consider all three aspects to give you the best chromatography.

You must also determine what type of inlet is in your GC

Then consider the application itself, and the types of liners and injection techniques used for it:

- Split
- Splitless

# **Liner Volume**

Choose a liner with enough volume to accommodate the vaporized sample.

Important, especially for polar solvents with large vapor volumes.

If vapor volume of sample exceeds liner volume, samples may back up (backflash) into carrier gas supply lines, causing ghost peaks and reproducibility problems in chromatography.

# Liner Volume (contd.)

Agilent liners are primarily 2mm or 4mm in inner diameter (without tapers and additional features) and 78mm long.

Thus, 2mm liners hold approx. 0.245 mL or 245 μL of vapor
 4mm liners hold approx. 0.972 mL or 972 μL of vapor

Recommended injection volumes are 1-2 $\mu$ L or less for organic solvents, 0.5 $\mu$ L for water.



### **Liner Volume**

How Do we Calculate the Vapor Volume?

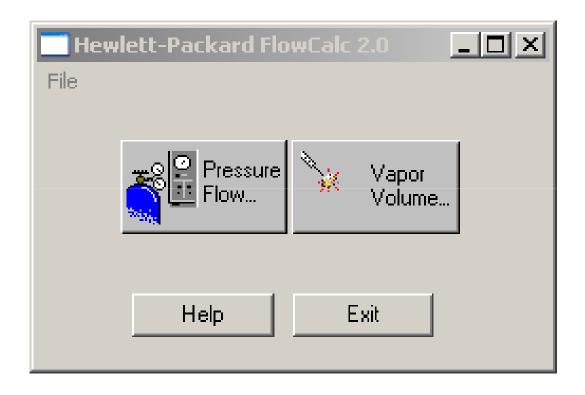
Pressure / Flow Calculator

Free download from our Website

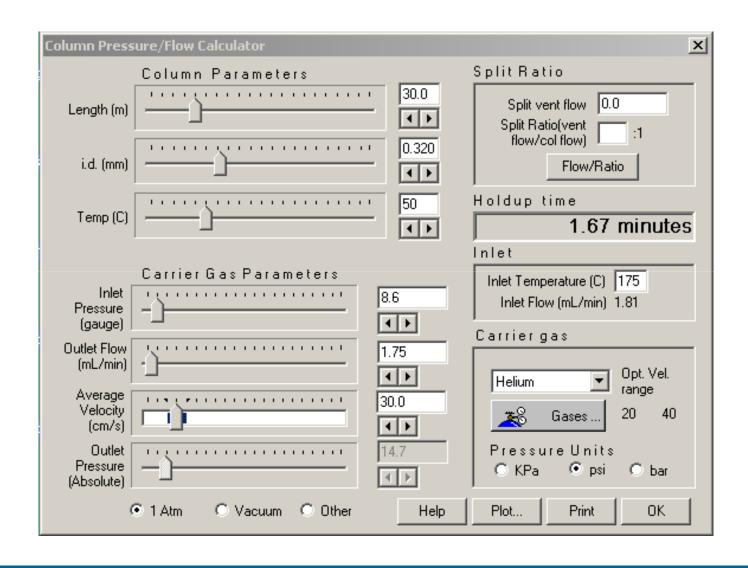
www.chem.agilent.com

<u>http://www.chem.agilent.com/en-</u>
<u>US/Support/Downloads/Utilities/Pages/GcPressureFlow.aspx</u>

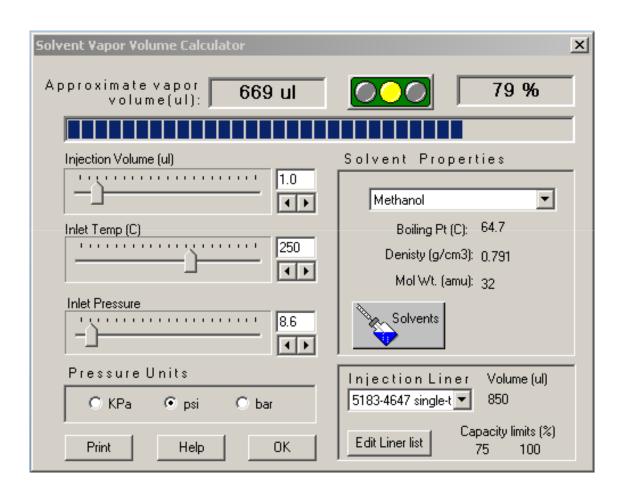
### **Pressure / Flow Calculator**



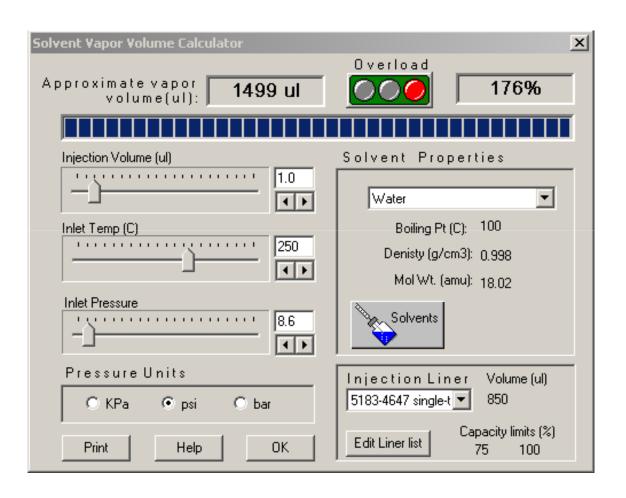
## Determine what the inlet pressure will be:



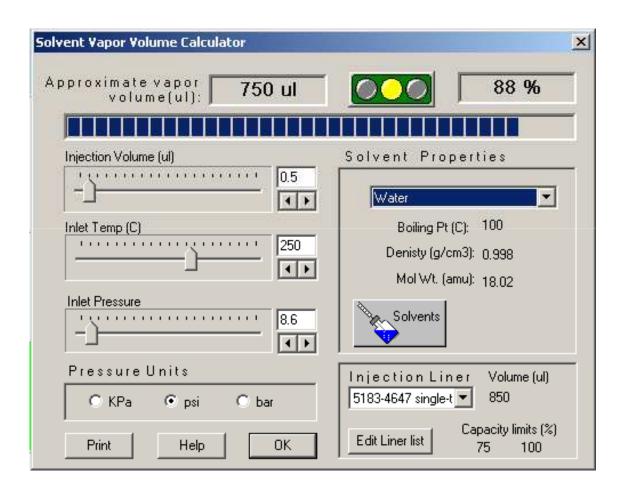
## **Test Inlet Conditions For Solvent Expansion**



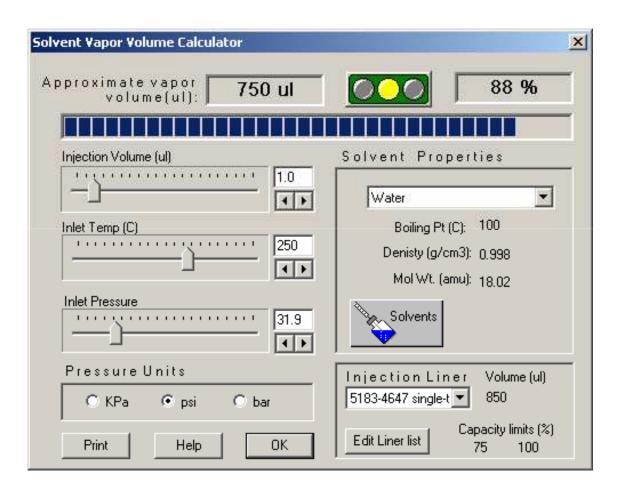
### **Water as Solvent**



# Water as Solvent Cut Injection Volume in Half



# Water as Solvent Pulsed Injection



## **Liner Treatments or Deactivation**

Minimizes possibility of active sample components from adsorbing on active sites on the liner or glass wool surface.

Unwanted sample adsorption leads to tailing peaks and loss of response for polar compounds.

Although not necessary for all applications, deactivated liners provide added insurance against possible sample adsorption.

Deactivation of borosilicate glass liners is often done with a silylating reagent like Dimethyldichlorosilane (DMDCS)



### **Special Characteristics**

Electronic Pressure Control with direct connect liners.

Some liners have special features that are necessary for different injection techniques. For example outlet inlet Taper (gooseneck), minimizes sample contact with gold seal. Dual taper, also minimizes sample contact with inlet weldment and reduces potential for backflash. Glass wool and shelf to hold it in place, prevents non-volatiles from reaching column and removes residual sample from needle. Glass wool should be deactivated. Jennings cup, normally used for efficient sample mixing in split inlets, reduces sample discrimination and prevents non-volatiles from reaching the column. Not for very dirty samples. Press fit (direct) connection end to hold capillary column firmly (virtually all sample goes onto the column). Side hole needed for

### **Special Characteristics (contd.)**

### Other special characteristics include:

- Baffles
- Spiral paths
- Glass or ceramic frits or beads
- Laminar cups (elongated version of Jennings cups)
- Column packings with stationary phases

### All designed to provide:

- a turbulent sample flow path for sample mixing
- protrusions, barriers, or adsorbents to collect high molecular weight sample components or particles
- surfaces for efficient vaporization of sample components.

# **Split Injection Liners**

Liner	Part	Comments
_	No.	
	19251- 60540	Simplest split liner, glass wool, no-deactivation, large volume, 990µL volume. Use for general purpose applications for compounds with low glass adsorption activity. Also used for Splitless mode.
Glass nub	5183-4647	Glass wool (held near needle entrance to remove residual sample on needle), deactivated, 870µL volume. Glass nub ensures that gap remains below liner for split injection. Efficient, for most applications, including active compounds. Fail-safe insertion into injection port. Needle length is important.
	18740- 80190	Liner with Jennings cup, no glass wool, 800µL volume. For manual injection only. Use for general purpose applications, high and low MW compounds. Reduces inlet discrimination.
	18740- 60840	Liner with Jennings cup, glass wool, and column packing, 800µL volume. For manual injection only. For dirty samples, traps non-volatiles and particulates well. For high and low MW compounds. Not recommended for use with EPC.

### <u></u>

# **Splitless Injection Liners**

Liner	Part No.	Comments
E	5181-3316	Single taper, deactivated, 900µL volume. Taper isolates sample from metal seal, reducing breakdown of compounds that are active with metals. For trace samples, general application.
	5062-3587	Single taper, deactivated, with glass wool, 900µL volume. Glass wool aides volatilization and protects column. For trace (dirty) samples.
8	5181-3315	Double taper, deactivated, 800µL volume. Taper on inlet reduces chance for backflash into carrier gas lines. High efficiency liner for trace, active samples.
Side hole	G1544-80730 G1544-80700	Direct connect liners, single and dual taper, deactivated. Capillary column press fits into liner end, eliminating sample exposure to inlet. Ultimate protection for trace, active samples. Side hole permits use with EPC.

# GLASS WOOL Liner Packing Recommendations

Amount, size and placement must be consistent for consistent results

Can be broken upon installation into the liner, exposing active sites

Liner deactivation with glass wool plug in place is ideal

# GLASS WOOL Placement in Liner

### Near top of liner:

- Wipes syringe needle of sample
- Can improve injector precision
- Helps to prevent backflash

### Near bottom of liner:

- Helps in volatilization of high MW components
- Increases mixing

Both positions help retain <u>some</u> non-volatile residues from reaching the column

## **Carrier Gas Considerations**

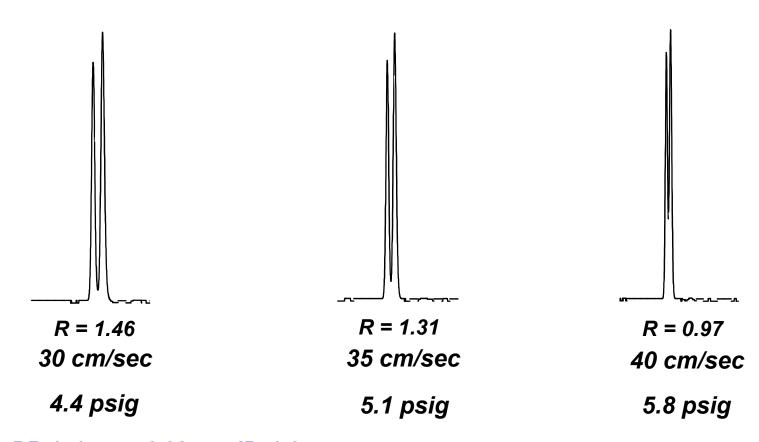
Carries the solutes down the column

 Selection and velocity influences efficiency and retention time

### **RESOLUTION VS. LINEAR VELOCITY**

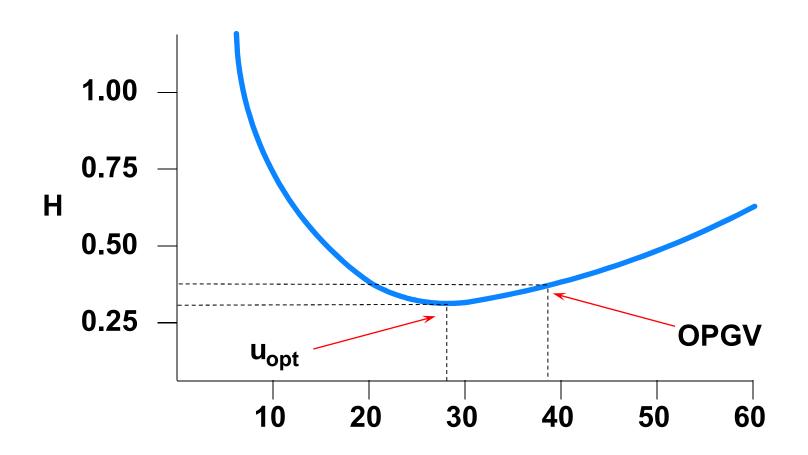
### Helium

**Resolution of 1.5 = baseline resolution** 



DB-1, 15 m x 0.32 mm ID, 0.25 um 60°C isothermal 1,3- and 1,4-Dichlorobenzene

## **VAN DEEMTER CURVE**



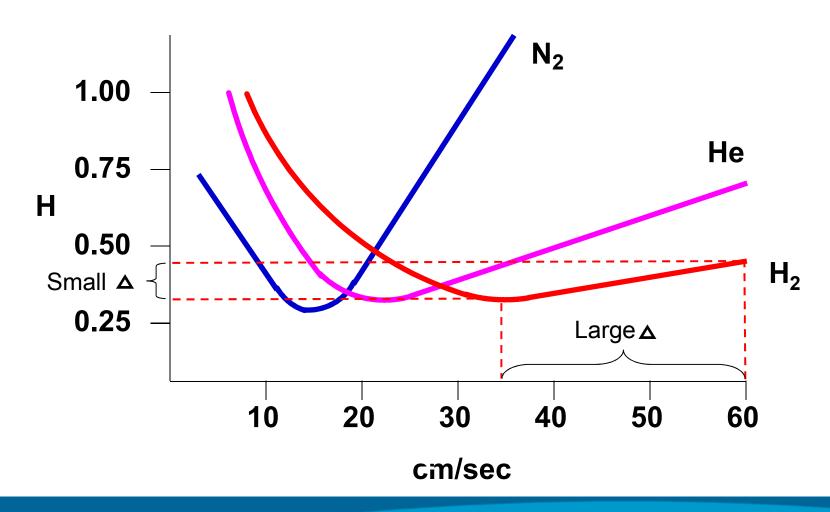
## $\overline{\mathbf{u}}_{\text{opt}}$ and OPGV

**U**opt: Maximum efficiency

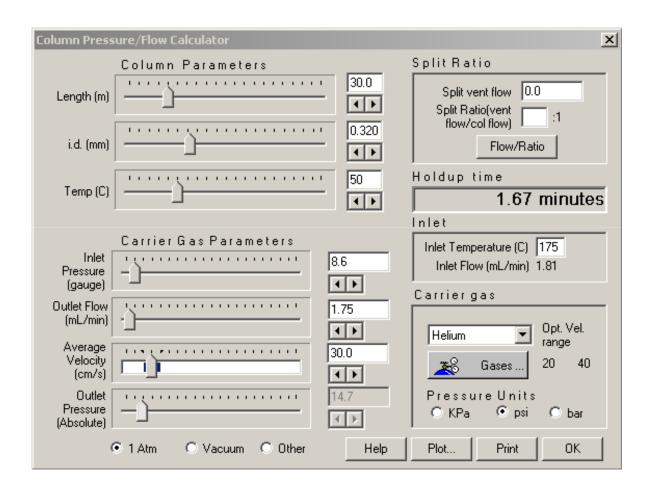
**OPGV:** Optimal practical gas velocity Maximum efficiency per unit time

$$1.5 - 2x \overline{U}_{opt}$$

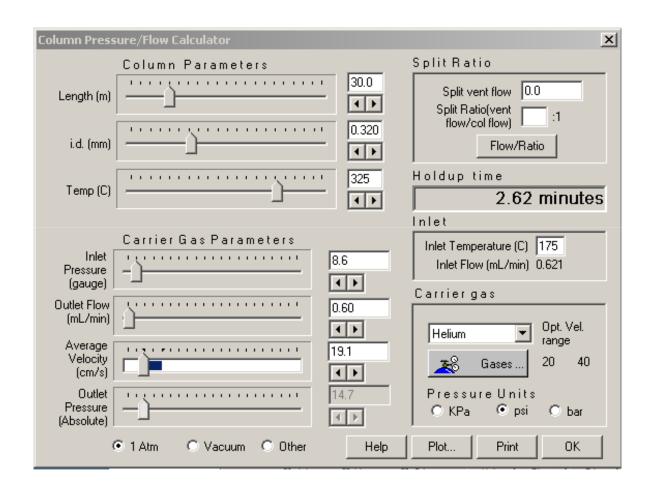
## **VAN DEEMTER CURVES**



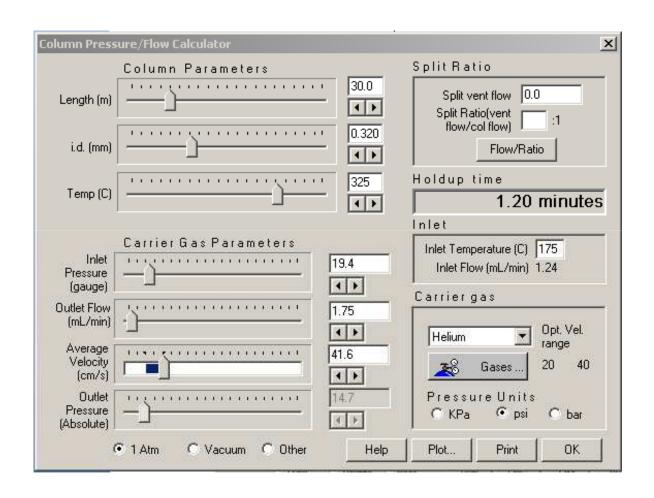
# What Happens to the Flow as Oven Temp Increases?



## **Carrier Gas: Constant Pressure**



## **Carrier Gas: Constant Flow**



## **Detectors**

Detector	Dynamic I	Range	MDL
TCD	10 <sup>5</sup>	Universal	400 pg Tridecane
FID	10 <sup>7</sup>	Responds to C-H bonds	1.8 pg Tridecane
ECD	5x10 <sup>5</sup>	Responds to free electrons	6 fg/mL Lindane
NPD	10 <sup>5</sup>	Specific to N or P	0.4 pgN/s 0.06 pg P /s
FPD	10 <sup>3</sup> S, 10 <sup>4</sup> P	Specific to S or P	60 fg P/s 3.6 pg S/s
SCD	10 <sup>4</sup>	Specific & Selective to S	0.5 pg S/s
NCD	10 <sup>4</sup>	Specific & Selective to N	3 pg N/s
MSD		Universal	S/N 400:1 1 pg/uL OFN

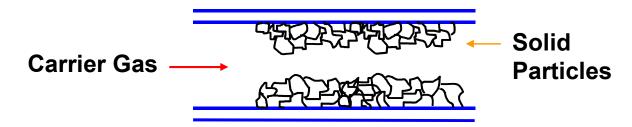
## **Selecting the RIGHT Column**

**Understanding the Stationary Phase** 

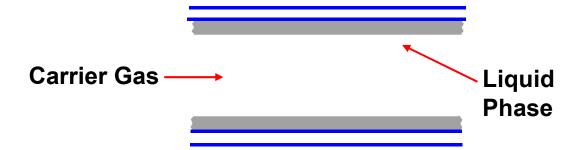
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## CAPILLARY COLUMN TYPES

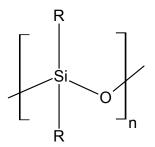
### **Porous Layer Open Tube (PLOT)**



### Wall Coated Open Tube (WCOT)



## STATIONARY PHASE POLYMERS



R= methyl, cyanopropyl, cyanopropylphenyl, trifluoropropyl

#### **Siloxane**

$$HO = \begin{bmatrix} H & H \\ - & - \\ - & - \\ - & - \\ - & - \\ - & - \\ - & - \end{bmatrix}_{n}$$

Polyethylene glycol backbone

#### **└**

## Selectivity Interactions

- Dispersion
- Dipole
- Hydrogen bonding

#### $\overline{}$

# Selectivity Interaction Strengths

Phase	Dispersion	Dipole	H Bonding
Methyl	Strong	None	None
Phenyl	Strong	None	Weak
Cyanopropyl	Strong	Strong	Moderate
Trifluoropropyl	Strong	Moderate	Weak
PEG	Strong	Strong	Moderate

## **Now Let's Apply What We learned**

## **Sample List (drugs)**

1. Cadaverine	$H_2$ N $\longrightarrow$ N $H_2$	11. Phenelzine	Ü.NH2
2. Cyclopentamine	H. C.	12. Phenylpropanolamine	OH NH <sub>2</sub> CH <sub>3</sub>
3. Amphetamine	NH <sub>2</sub>	13. Clortermine	CI NH <sub>2</sub>
4. Phenethylamine	NF <sub>2</sub>	14. Chlorphentermine	CI NH <sub>2</sub>
5. Phentermine	NH <sub>2</sub>	15. Ephedrine	OH CH3
6. Propylhexedrine	HN CH3	16. Pseudoephedrine	HIN CH3
7. Methamphetamine	TZ	17. Phendimetrazine	
8. Methenamine	N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-N-	18. MDA	© T NH₀
9. Amantidine	NH <sub>2</sub>	19. Ecgonine methyl ester	H- W "H
10. Mephentermine		20. diethylpropion	

## **Starting Method Parameters**

Column: DB-5 30m X 0.32mm X 0.25um

S/SI Inlet: Split 50:1 Temp 250°

FID: Temp 350°

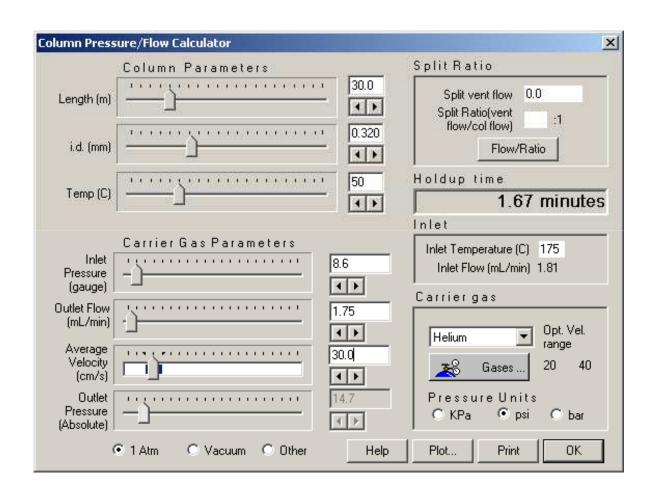
Carrier: He

Constant flow 30cm/sec

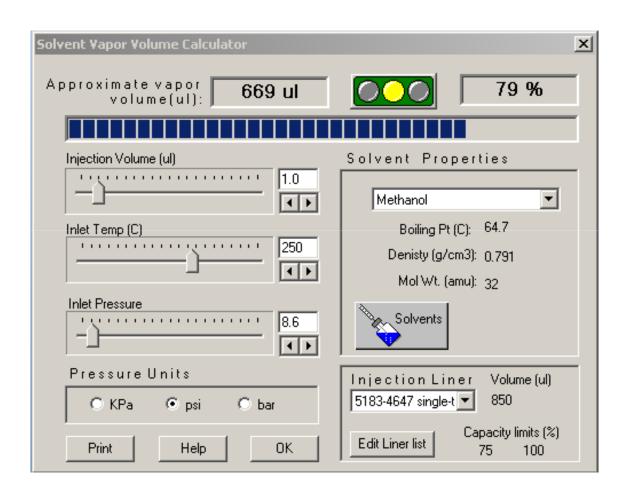
Oven: 50°C Hold for 5 min

10°C/min to 325°C Hold for 5 min

## **Am I Going to Have Backflash?**

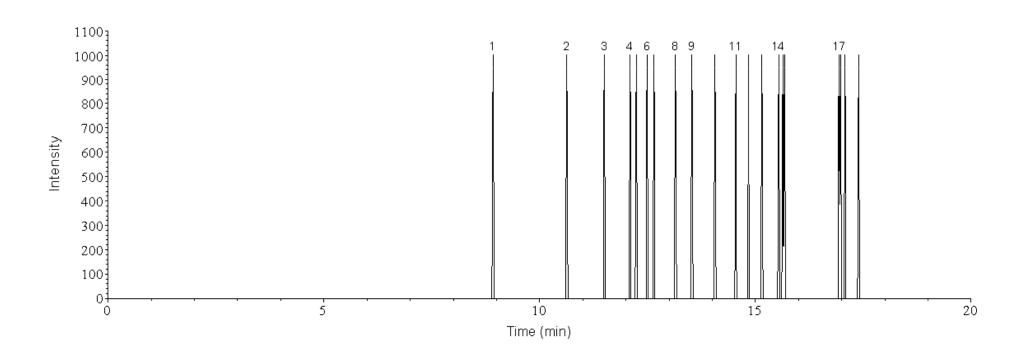


## Injection Volume / Solvent Expansion

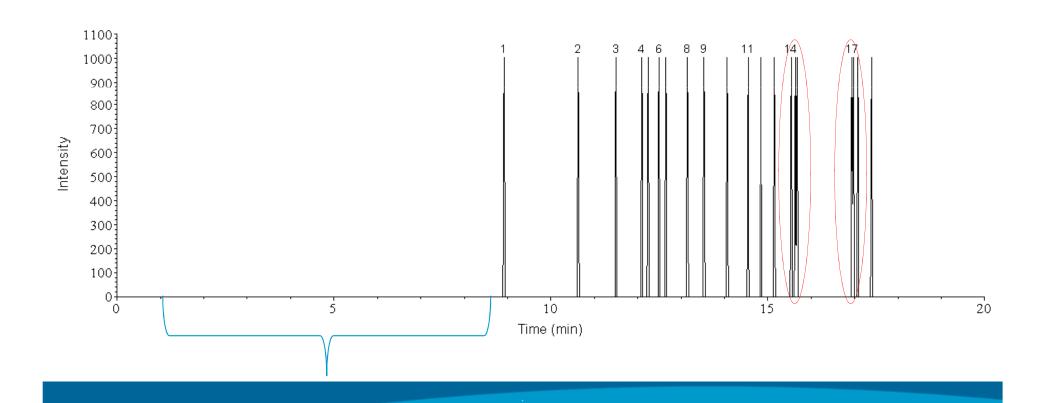


# **Developing Temperature Program Initial Run**

Initial Temp 50°C Hold for 5 min Ramp 10°C/min to 325°C Hold for 5 min



# **Developing Temperature Program**Initial Run - Define Areas for Improvement



**Agilent Technologies** 

## **Next Step...**

When does the first peak come out?

~9 minutes

What temperature does it come out at?

Temp program:

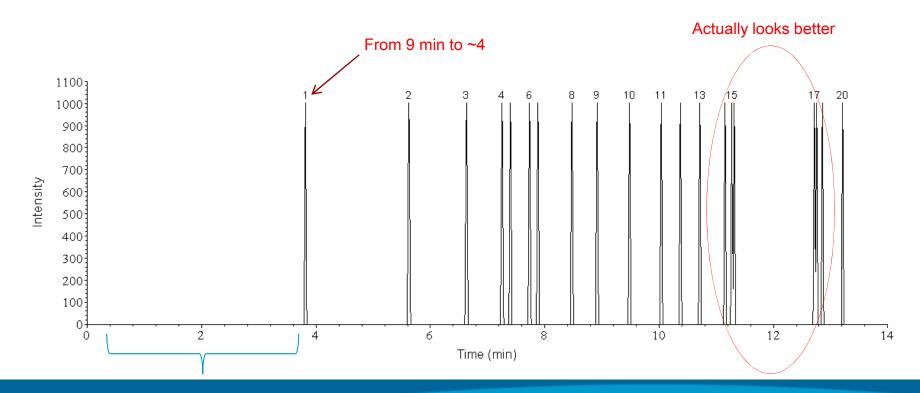
50°C for 5 minutes

10°C to 325°C

1st Peak comes out at 90°C

# Developing Temperature Program 2<sup>nd</sup> Try

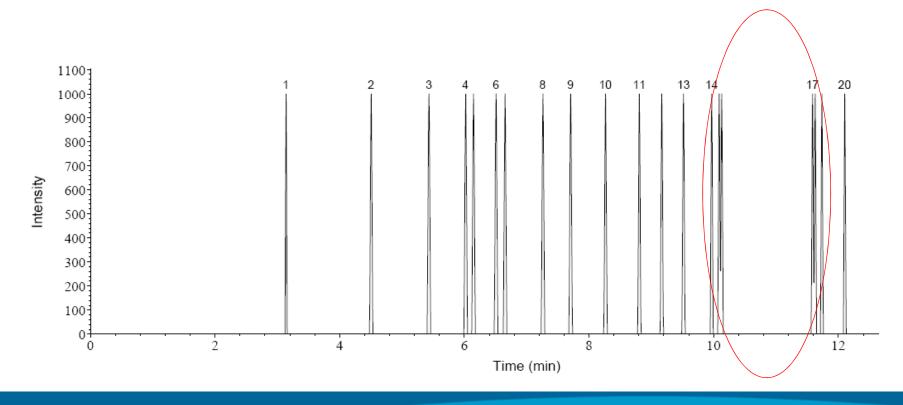
Initial Temp 90°C Hold for 5 min Ramp 10°C/min to 325°C Hold for 5 min



# **Developing Temperature Program 3rd Try**

Initial Temp 100°C Hold for 5 min Ramp 10°C/min to 325°C Hold for 5 min

Time to resolve these peaks



## **Resolve Co-elutions**

Add a hold 20-30° below the elution temperature

Co-elutions occur at 10 minutes

100°C hold for 5 minutes 10°C/min to 325°C

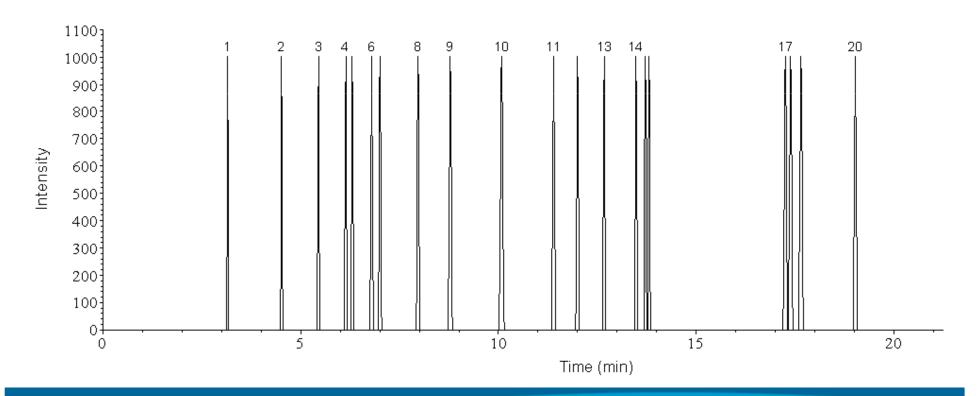
Co-elutions occur at 150°C

Set hold at 130°C

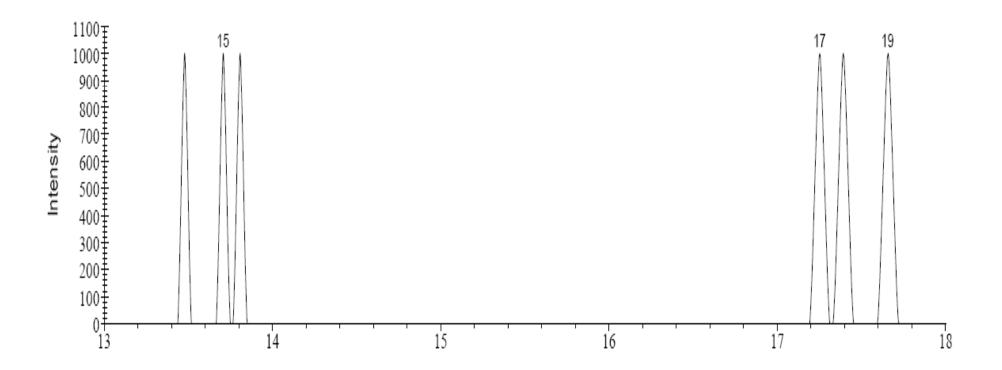
## **Developing a Temperature Program**

Oven: 100°C Hold for 10 minutes 10°C/min to 130°C hold for 5 min

10°C/min to 325°C



## **Developing a Temperature Program**



### **Conclusions:**

Think about the sample first

\*\*Is it chromatographable by GC?

sample composition

sample clean up

level of detection

Use information sources first when choosing a column

Mild oven program to begin with

**Utilize Technical Support** 

## **Agilent J&W Scientific Technical Support**

800-227-9770 (phone: US & Canada)\*

\* Select option 3, then 3, then 1.

866-422-5571 (fax)

GC-Column-support@agilent.com





www.chem.agilent.com