

A simple and robust CE-Marked diagnostic assay for complete mutation analysis of the *CFTR* gene by Next-Generation Sequencing. Mutations of the *CFTR* gene affect functioning of the chloride ion channels in epithelial cell membranes, leading to cystic fibrosis and other *CFTR*-related diseases.



Application

- For first or second line testing in your CFTR work-up
- For whole blood & dried blood spots derived DNA
- For complete mutational spectrum analysis



Assay characteristics

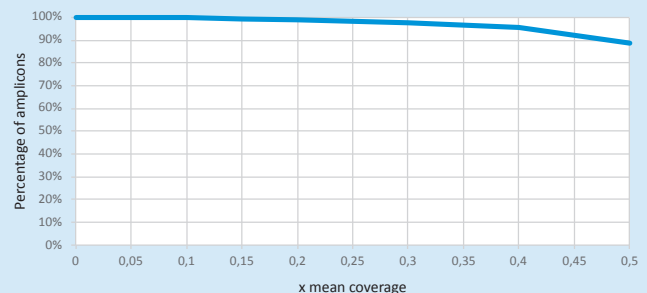
Gene analyzed	<i>CFTR</i>
Genomic region analyzed	8.7 kb All exons ± 30 bp flanking region, selected introns, part of promoter
Number of amplicons	48 including 11 control amplicons
Amplicon length	300-450 bp
Number of plexes	2
DNA amount required	20 ng per multiplex reaction
Validated (CE-IVD) with	MiSeq® System, Illumina
Mutations included	
All exonic SNVs & small indels CNVs*	~30 bp of intronic flanking sequence
HP variants	intron 9 (8): c.1210-34TG(m); c.1210-12T(n)
Selected deep intronic regions:	intron 7 (6b): c.1002-1110_1113delTAAG c.1002-1111A>C
	intron 12 (11): c.1679+1634A>G (1811+1.6kbA->G) c.1680-877G>T (1811+1643G->T) c.1680-883A>G
	intron 22 (19): c.3718-2477C>T (3849+10kbC->T)
Promoter region	up to position c.-108

*For Research Use Only. Not for use in diagnostic procedures

CFTR MASTR Dx on MiSeq, germline DNA

Performance Characteristics

	DNA extracted from blood	DNA extracted from dried blood spots
On target read pair counts	98.92 % [92.70 %-99.51 %]	99.90 % [99.53 %-99.98 %]
Uniformity of amplification (0.2x mean coverage)	9783 %	100 %



Graph representing the read counts for all 48 CFTR MASTR Dx amplicons, showing their uniform representation. To allow comparison between samples, the read counts were normalized.

CFTR MASTR Dx on MiSeq; germline DNA

Performance Characteristics

Performance parameter	Extracted from blood		Extracted from dried blood spots	
	Observed [95 % CI] Using JSI SeqNext	Observed [95 % CI] Using Sophia DDM	Observed Using JSI SeqNext	Observed Using Sophia DDM
Sensitivity	100 % [≥ 99.217 %]	100 % [≥ 98.202 %]	64/64 = 100 %	64/64 = 100 %
Specificity	99.990 % [99.975 % -99.994 %]	100 % [≥ 99.999 %]	1438/1438 = 100 %	1438/1438 = 100 %
Accuracy	99.990 % [99.975 % -99.994 %]	100 % [≥ 99.999 %]		
Repeatability & reproducibility	99.997 % [99.988 % -99.999 %]	100 % [≥ 99.990 %]		

Advised maximum number of samples per run

Workflow

Sequencing System	Illumina MiSeq®		
	Reagent Kit		
Flow cell	Nano v2 2 x 251 cycles	v2 2 x 251 cycles	v3 2 x 276 cycles
SNV variant calling Minimal coverage per allele: 50	52	786	1441
SNV and CNV* variant calling Minimal coverage per amplicon: 200	26	393	720

*For Research Use Only. Not for use in diagnostic procedures



Publications

Supporting publications:

Applicability and Efficiency of NGS in Routine Diagnosis: In-Depth Performance Analysis of a Complete Workflow for CFTR Mutation Analysis. Andrien Pagin et al. (2016)

[PLOS ONE DOI: 10.1371/journal.pone.0149426](https://doi.org/10.1371/journal.pone.0149426), 22 Feb 2016*

*This study was done by a third party and was verified by Multiplicom.

Order information

Cat. No.	Product Name	Reactions	PR Number
MR-2021.024	CFTR MASTR Dx	24	PR7000-1421
MR-2021.048		48	

MID (Molecular Identifiers) kits are necessary to complete the workflow.

5991-8404EN
Printed in Belgium, September 2017