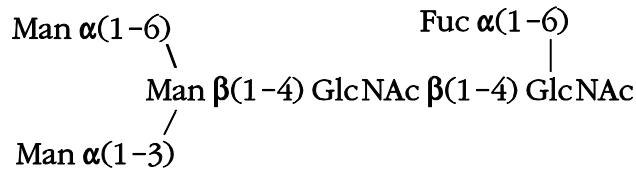


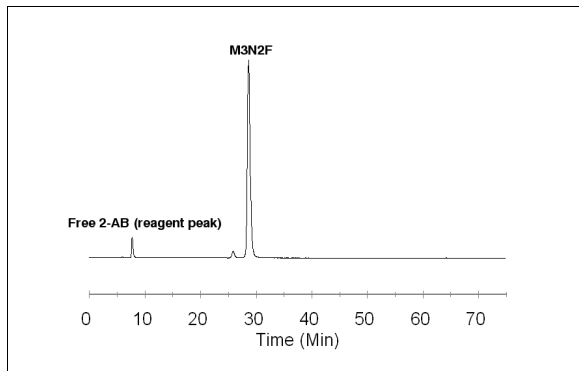
## CERTIFICATE OF ANALYSIS

**PRODUCT NAME:** GLYKO® CONSERVED TRIMANNOSYL CORE, SUBSTITUTED WITH FUCOSE (M3N2F)  
**PRODUCT CODE:** GKR-002301  
**LOT NUMBER:** DP09H1101  
**PACK SIZE:** 10 µg (qualitative standard for glycan identification)  
**PURITY:** ≥90% of glycan by HPLC  
**FORM:** Dry solid  
**STORAGE:** Store at -20°C before and after reconstitution  
**EXPIRATION:** August 2014, may be used for 1 year after reconstitution

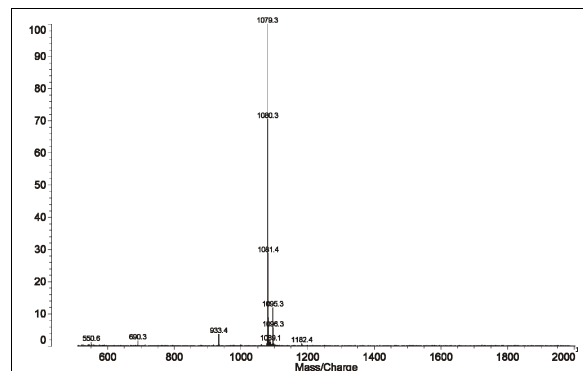
**STRUCTURE:**



**QUALITY CONTROL:**



**Figure 1 - HPLC of 2-AB labeled M3N2F**



**Figure 2 - MALDI-TOF of M3N2F [M + Na]<sup>+</sup>**

**Molecular Weight:** 1057.0 (average)<sup>1</sup>

**Isolation:** M3N2F is a substructure common to most of the N-linked oligosaccharides, which are widely found on glycoproteins. M3N2F N-linked core oligosaccharide is typically released from a glycoprotein using N-Glycanase® or anhydrous hydrazine<sup>8</sup>, separated from peptide material by adsorption chromatography, then purified further using a combination of glycosidase digestion and chromatographic techniques.

**Structural Analysis:** The purity and structural integrity of the glycan is assessed by one or more of the following techniques: HPLC<sup>2</sup>, mass spectrometry<sup>3,4</sup>, FACE<sup>5</sup>, <sup>1</sup>H-NMR<sup>6</sup> and HPAEC-PAD<sup>7</sup>.

### Applications:

- qualitative standard for various analytical procedures
- radio-labeling, fluorescent-labeling or formation of a variety of oligosaccharide derivatives
- substrate for glycosidase and glycosyl transferase assays

**Handling:** The oligosaccharide is shipped as a dried solid. Allow the unopened vial to reach ambient temperature and tap on a solid surface to ensure that most of the material is at the bottom of the vial. Gently remove the cap, add the desired volume of water or buffer, re-cap and mix thoroughly to redissolve all the oligosaccharide. For maximal recovery, ensure that the cap lining is also rinsed, and centrifuge the reconstituted vial briefly before use.

Make sure that any glassware, plasticware, solvents or reagents which come into contact with the glycan are free of glycosidases and carbohydrate contaminants.

Minimize exposure to elevated temperatures or extremes of pH. High pH will cause epimerization of the reducing terminal GlcNAc.

**Reconstitution:** Use HPLC-grade water or an aqueous buffer to dissolve the glycan. Store the reconstituted glycan at -20°C in working aliquots. Avoid multiple freeze/thaw cycles.

## REFERENCES

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<http://us.expasy.org/tools/glycomod/glycanmass.html>
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4. Papac DI, Wong A and Jones AJS. Analysis of acidic oligosaccharides by matrix-assisted laser desorption/ionization time of flight mass spectrometry. *Anal Chem* 1996 Sep 15;68(18):3215-3223.
5. Starr CM, Masada RI, Hague C, Skop E and Klock JC. Fluorophore-assisted carbohydrate electrophoresis in the separation, analysis, and sequencing of carbohydrates. *J Chromatogr A* 1996 Jan 12;720(1-2):295-321.
6. Vliegenthart JFG, Dorland L and van Halbeek H. High-resolution, <sup>1</sup>H-nuclear magnetic resonance spectroscopy as a tool in the structural analysis of carbohydrates related to glycoproteins. *Adv Carb Chem Biochem* 1983 41: 209-374 (1983).
7. Townsend RR, Hardy MR, Hindsgaul O and Lee YC. High-performance anion-exchange chromatography of oligosaccharides using pellicular resins and pulsed amperometric detection. *Anal Biochem* 1988 Nov 1;174(2):459-70.
8. Takasaki S, Mizouchi T and Kobata A. Hydrazinolysis of asparagine-linked sugar chains to produce free oligosaccharides. *Meth Enzymol* 1982; 83:263-8

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Authorized Signature