

CERTIFICATE OF ANALYSIS

PRODUCT NAME: GLYKO[®] 6- α -FUCOSYL CHITOBIOSE (N2F)

PRODUCT CODE: GKR-002001

LOT NUMBER: DP13B2001

PACK SIZE: 100 μ g (qualitative standard for glycan identification)

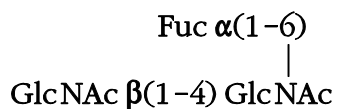
PURITY: $\geq 90\%$ of glycan by HPLC

FORM: Dry solid

STORAGE: Store at -20°C before and after reconstitution

EXPIRATION: March 2018, may be used for 1 year after reconstitution

STRUCTURE:



QUALITY CONTROL:

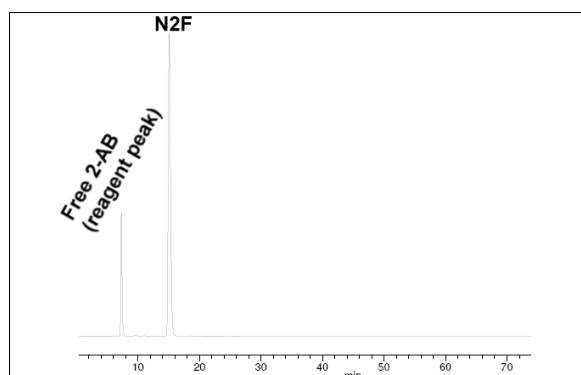


Figure 1 - HPLC of 2-AB labeled N2F

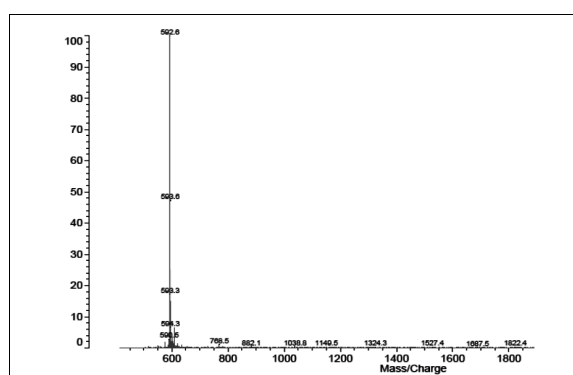


Figure 2 - MALDI-TOF of N2F $[M + Na]^+$

Molecular Weight: 570.5 (average)¹

Isolation: N2F is a core structure common to most of the N-linked oligosaccharides, which are widely found on glycoproteins. N2F N-linked core oligosaccharide is chemically synthesized.

Structural Analysis: The purity and structural integrity of the glycan is assessed by one or more of the following techniques: HPLC², mass spectrometry^{3,4}, FACE^{™5}, ¹H-NMR⁶ and HPAEC-PAD⁷.

Applications:

- qualitative standard for various analytical procedures
- radio-labeling, fluorescent-labeling or formation of a variety of oligosaccharide derivatives
- substrate for glycosidase and glycosyl transferase assays

Handling: The oligosaccharide is shipped as a dried solid. Allow the unopened vial to reach ambient temperature and tap on a solid surface to ensure that most of the material is at the bottom of the vial. Gently remove the cap, add the desired volume of water or buffer, re-cap and mix thoroughly to redissolve all the oligosaccharide. For maximal recovery, ensure that the cap lining is also rinsed, and centrifuge the reconstituted vial briefly before use.

Make sure that any glassware, plasticware, solvents or reagents which come into contact with the glycan are free of glycosidases and carbohydrate contaminants.

Minimize exposure to elevated temperatures or extremes of pH. High pH will cause epimerization of the reducing terminal GlcNAc.

Reconstitution: Use HPLC-grade water or an aqueous buffer to dissolve the glycan. Store the reconstituted glycan at -20°C in working aliquots. Avoid multiple freeze/thaw cycles.

REFERENCES

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5. Starr CM, Masada RI, Hague C, Skop E and Klock JC. Fluorophore-assisted carbohydrate electrophoresis in the separation, analysis, and sequencing of carbohydrates. *J Chromatogr A* 1996 Jan 12;720(1-2):295-321.
6. Vliegenthart JFG, Dorland L and van Halbeek H. High-resolution, ¹H-nuclear magnetic resonance spectroscopy as a tool in the structural analysis of carbohydrates related to glycoproteins. *Adv Carb Chem Biochem* 1983 41: 209-374 (1983).
7. Townsend RR, Hardy MR, Hindsgaul O and Lee YC. High-performance anion-exchange chromatography of oligosaccharides using pellicular resins and pulsed amperometric detection. *Anal Biochem* 1988 Nov 1;174(2):459-70.
8. Takasaki S, Mizouchi T and Kobata A. Hydrazinolysis of asparagine-linked sugar chains to produce free oligosaccharides. *Meth Enzymol* 1982; 83:263-8

Authorized Signature