

Fast and Accurate Quantitation of Organic Acids Using an Agilent 6546 LC/Q-TOF

Authors

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Abstract

Organic acid analysis is critical for identifying inborn errors of metabolism. Traditional GC/MS workflows require derivatization and long run times, often leading to false positives or negatives. This study demonstrates a high-resolution accurate mass (HRAM) workflow using the Agilent 6546 LC/Q-TOF. The method enables simultaneous qualitative and quantitative analysis of over 60 organic acids in under 20 minutes without derivatization. Enhanced selectivity and resolution allow confident identification of isobaric compounds. The workflow supports life science research applications requiring fast, accurate, and comprehensive organic acid profiling.

Introduction

Organic acids are important biomarkers in life science research, especially for identifying inborn errors of metabolism. Traditionally, gas chromatography/mass spectrometry (GC/MS) has been used for their analysis.¹ However, GC/MS workflows require derivatization and long run times, which can delay results and increase the risk of inaccurate findings.²

The Agilent 6546 LC/Q-TOF offers a faster, more selective alternative. It uses high-resolution accurate mass (HRAM) detection, which improves confidence in compound identification. HRAM enables precise measurement of molecular masses, helping distinguish compounds even in complex mixtures. This LC/Q-TOF can detect small differences in mass that lower-resolution instruments may miss, supporting more reliable identification and quantitation.

Chromatographic separation also plays a critical role in accurate analysis. Selecting the right analytical HPLC column is essential for resolving compounds that may have similar chemical structures or retention times. In this workflow, an Agilent InfinityLab Poroshell 120 Phenyl Hexyl column was chosen for its ability to separate a broad range of organic acids, including challenging isobaric pairs. This ensures cleaner separation, better peak shape, and more dependable quantitation.

Together, HRAM detection and optimized chromatography enable a fast, streamlined LC/MS workflow that supports both qualitative and quantitative analysis of over 60 organic acids.

Experimental

This workflow targets over 60 organic acids relevant to life science research (Table 1). A custom personal compound database and library (PCDL) was created using Agilent MassHunter PCDL Manager to support high-resolution identification. Thirteen isobaric sets were resolved using HRAM and optimized chromatography (Table 2).

Table 1. Organic acid analyte panel.

No.	Organic Acid	No.	Organic Acid	No.	Organic Acid	No.	Organic Acid
1	Lactic acid	16	p-OH-Phenylacetic acid	31	Malonic acid	46	N-Acetyl-leucine
2	Pyruvic acid	17	p-OH-Phenyllactic acid	32	3-Hydroxy-3-methylglutaric acid	47	Erythro-2,3 dihydroxy-methylbutyric acid
3	Succinic acid	18	p-OH-Phenylpyruvic acid	33	Isovalerylglycine	48	Uracil
4	Fumaric acid	19	Succinylacetone	34	Hexanoylglycine	49	3-Methylcrotonylglycine
5	2-Ketoglutaric (2-KG)	20	Glutaric acid	35	Suberylglycine	50	Isobutyrylglycine
6	Methylmalonic acid	21	3-Methylglutaconic acid	36	Orotic acid	51	Homovanillic acid
7	3-OH-Butyric acid	22	3-Hydroxyglutaric acid	37	Hydantoin 5-propionate	52	Vanilmandelic acid
8	Acetoacetic acid (AAA)	23	2-Hydroxyglutaric acid	38	4-OH-Butyric acid	53	Vanilactic acid
9	2-Keto-3-methylvaleric acid (KMV)	24	Pyroglutamic acid	39	Homogentisic acid	54	Propionylglycine
10	2-Ketoisocaproic acid (KIC)	25	3-OH-Propionic acid	40	Mevalonic acid	55	2-Methylbutyrylglycine
11	2-Ketoisovaleric acid (KIV)	26	Methylsuccinic acid	41	N-Acetylaspartic acid	56	4-Phenylbutyric acid
12	Ethylmalonic acid	27	2-OH-Isovaleric acid	42	Glycolic acid	57	Phenylacetic acid
13	Adipic acid	28	3-OH-isovaleric acid	43	Glyceric acid	58	Phenylpropionylglycine
14	Suberic acid	29	3-Methylglutaric acid	44	3-OH-Isobutyric acid	59	Tiglylglycine
15	Sebacic acid	30	2-Methylcitric acid	45	Phenyllactic acid	60	N-Butyrylglycine

Table 2. Isobaric sets of organic acids.

No.	Isobars	No.	Isobars
1	3-Hydroxypropionic acid, lactic acid	8	Adipic acid, 3-methylglutaric acid
2	3-Hydroxybutyric acid, 4-hydroxybutyric acid, 3-hydroxyisobutyric acid	9	2-Hydroxyglutaric acid, 3-hydroxyglutaric acid
3	Succinic acid, methylmalonic acid	10	3-Methylcrotonylglycine, tiglylglycine
4	2-Hydroxyvaleric acid, 3-hydroxyvaleric acid	11	Isovalerylglycine, 3-methylbutyrylglycine
5	2-Keto-3-methylvaleric acid, 2-keto-n-caproic acid	12	Hexanoylglycine, acetyl-leucine
6	Ethylmalonic acid, glutaric acid, methylsuccinic acid	13	Hydroxyphenylacetic acid, homovanillic acid
7	Isobutyrylglycine, n-butyrylglycine		

LC separation was performed on an Agilent 1290 Infinity II LC with an Agilent InfinityLab Poroshell 120 Phenyl Hexyl column. Method conditions are summarized in Table 3. Agilent MassHunter Acquisition software was used to control the LC and MS systems.

Mass spectrometry was conducted on an Agilent 6546 LC/Q-TOF, operating in positive ion mode with an Agilent Jet Stream Technology ion source (AJS). Parameters are listed in Table 4. Compound confirmation was performed using MassHunter Qualitative Analysis, and quantitation was completed using MassHunter Quantitative Analysis.

Results and discussion

The 6546 LC/Q-TOF workflow enabled fast, accurate separation and quantitation of over 60 organic acids. Chromatographic separation using the InfinityLab Poroshell 120 Phenyl Hexyl column produced clean peak shapes and consistent retention times across the full panel. An example chromatogram demonstrating separation of all 60 compounds is shown in Figure 1.

Table 3. LC method parameters.

Parameter	Value												
LC System	Agilent 1290 Infinity II LC												
HPLC Column	Agilent InfinityLab Poroshell 120 Phenyl Hexyl, 2.1 × 150 mm, 2.7 μm (p/n 693775-912)												
Column Temperature	30 °C												
Mobile Phase	A) 0.01% Formic acid in water B) Methanol												
Flow Rate	0.5 mL/min												
Injection Volume	1 μL												
Gradient	<table border="1"> <thead> <tr> <th>Time (min)</th> <th>B (%)</th> </tr> </thead> <tbody> <tr> <td>0.00</td> <td>0</td> </tr> <tr> <td>3.50</td> <td>0</td> </tr> <tr> <td>18.00</td> <td>60</td> </tr> <tr> <td>18.01</td> <td>95</td> </tr> <tr> <td>20.00</td> <td>95</td> </tr> </tbody> </table>	Time (min)	B (%)	0.00	0	3.50	0	18.00	60	18.01	95	20.00	95
Time (min)	B (%)												
0.00	0												
3.50	0												
18.00	60												
18.01	95												
20.00	95												
Post Time	3.0 min												
Stop Time	19 min												

Table 4. MS method parameters.

Parameter	Value
MS System	Agilent 6546 LC/Q-TOF
Gas Temperature	150 °C
Gas Flow	10 L/min
Nebulizer	35 psig
Sheath Gas Temperature	400 °C
Sheath Gas Flow	12 L/min
Capillary Voltage	2,000
Nozzle Voltage	500
Fragmentor	125

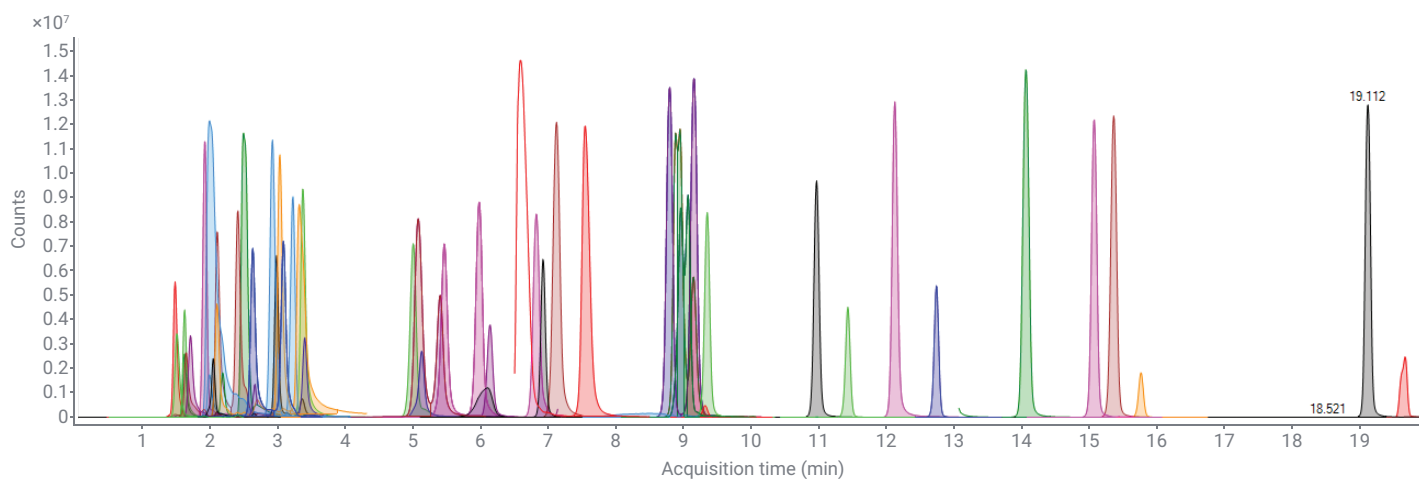


Figure 1. Example chromatogram showing the separation of 60 organic acids.

A key advantage of this method is its ability to resolve isobaric compounds—molecules with identical masses but different structures. Thirteen isobaric sets were successfully separated. Selected examples of these separations are shown in Figure 2.

Quantitative performance was evaluated using four-point calibration curves. Figure 3 shows calibration data for phenyllactic acid, and Figure 4 shows calibration for sebacic acid. Both compounds were chosen for their analytical difficulty and metabolic significance. The method demonstrated consistent linearity and reproducibility across the calibration range.

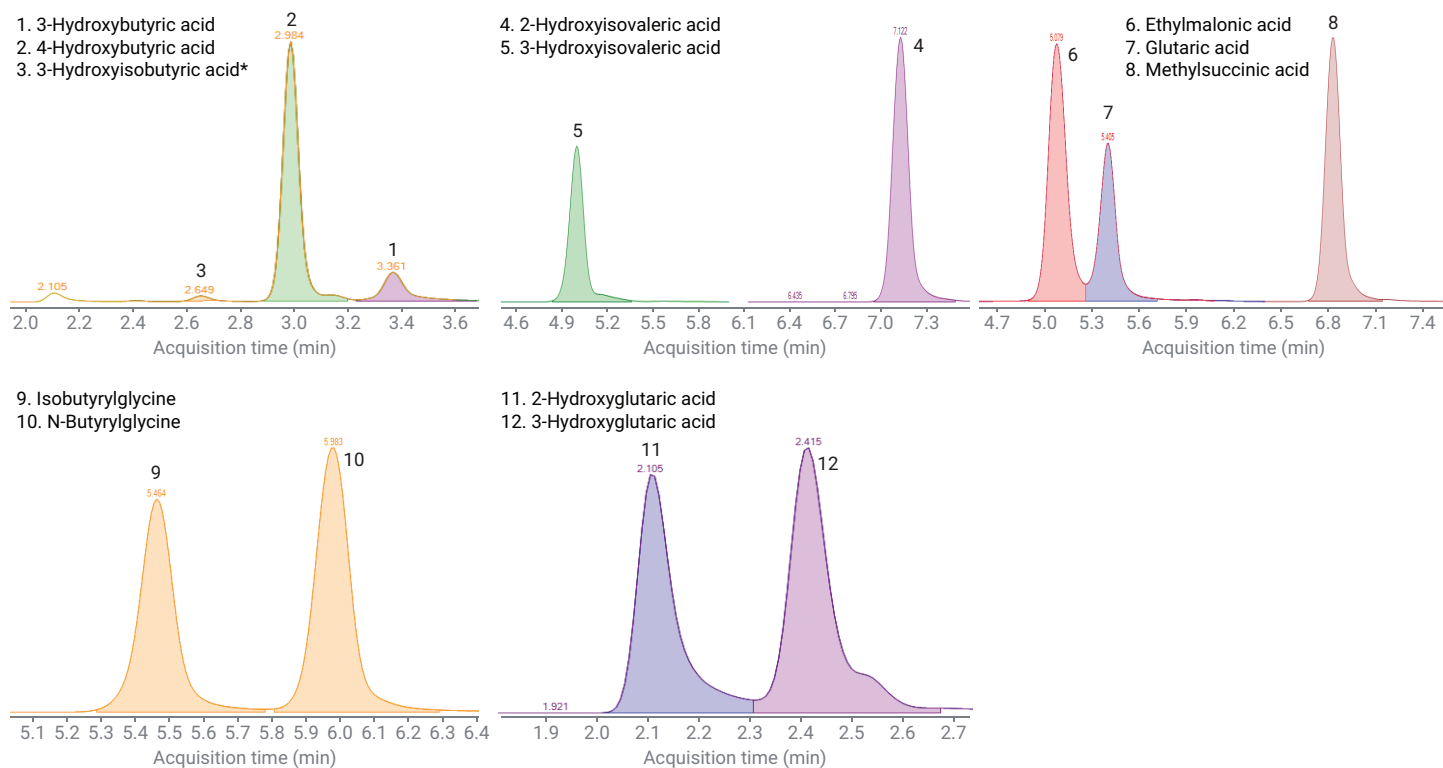


Figure 2. Select separations of isobaric sets of organic acids.

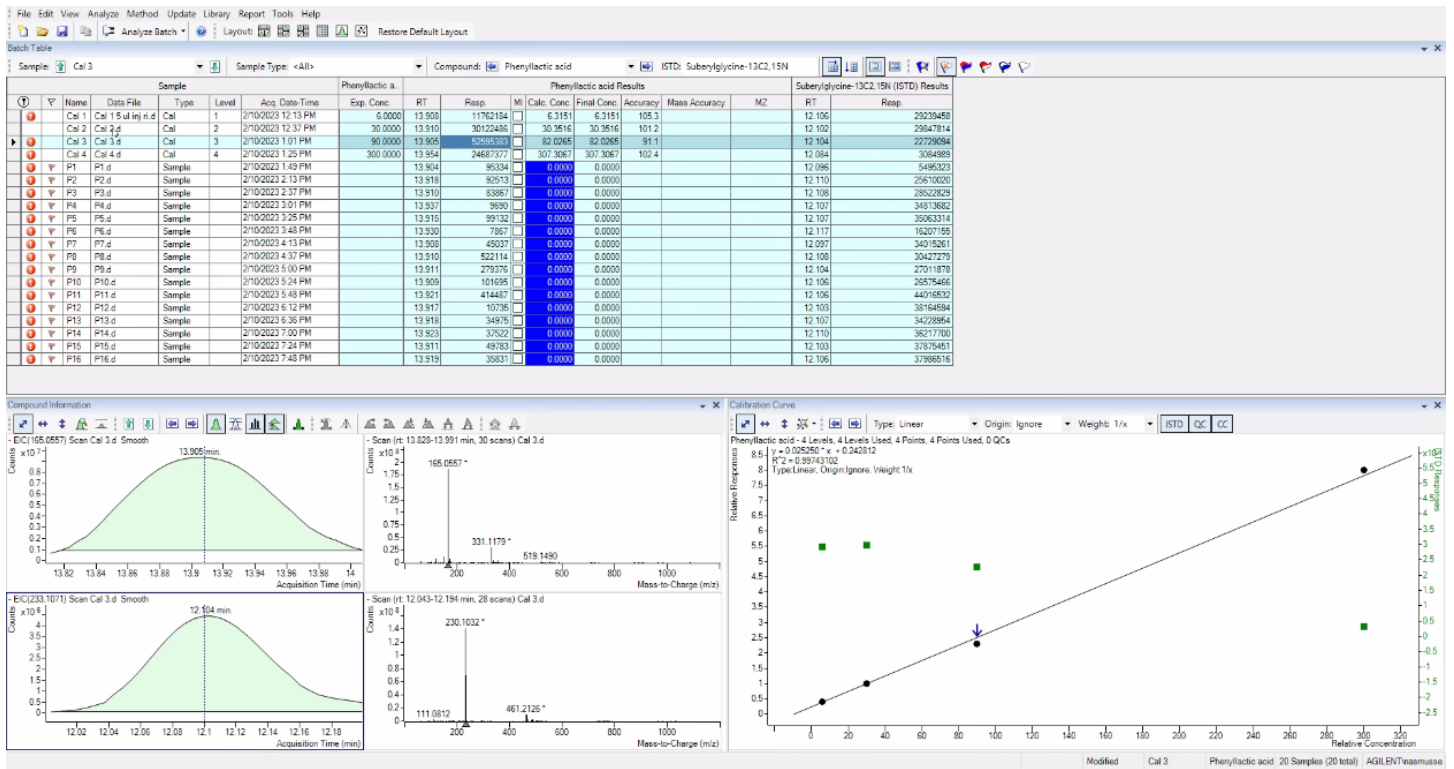


Figure 3. Four-point calibration data for phenylactic acid.

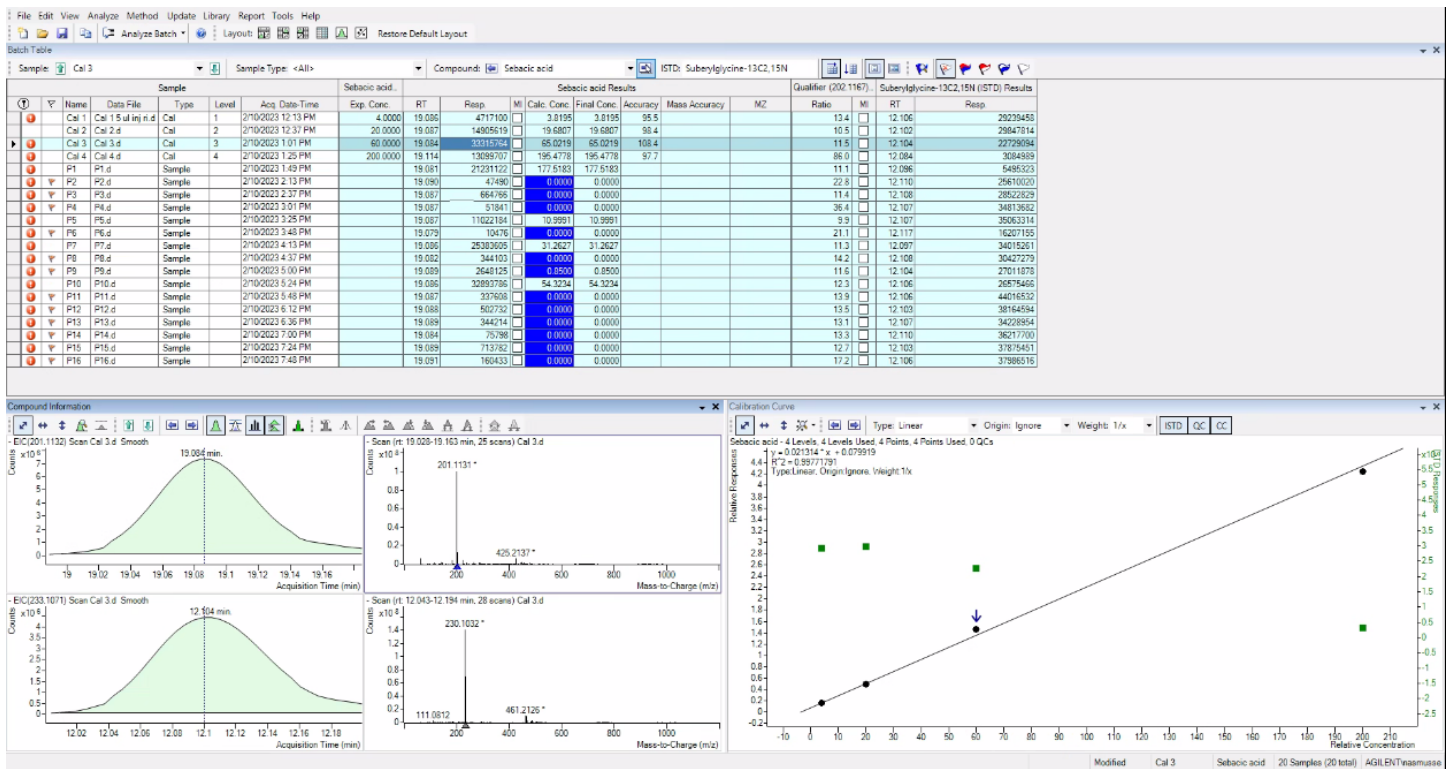


Figure 4. Four-point calibration data for sebacoic acid.

Data acquisition was performed using MassHunter Acquisition software, with compound confirmation completed using MassHunter Qualitative Analysis and the custom PCDL. Quantitative results were processed using MassHunter Quantitative Analysis, which also supported streamlined reporting. An example of reporting output is shown in Figure 5, illustrating calibration and sample data in a clear, review-ready format.

Together, the combination of HRAM detection, optimized chromatography, and integrated software tools supports confident, high-throughput organic acid profiling in under 20 minutes.

Conclusion

This study demonstrates a streamlined LC/MS workflow for the analysis of over 60 organic acids using an Agilent 6546 LC/Q-TOF. By eliminating derivatization and reducing analysis time, the method offers a faster, more efficient alternative to traditional GC/MS approaches. High-resolution accurate mass detection with an optimized chromatographic separation enables confident separation of isobaric compounds with reliable peak shape and retention. Integrated software tools—including Agilent MassHunter Acquisition, Qualitative Analysis, PCDL Manager, and Quantitative Analysis—support a complete workflow from data collection to reporting. Together, these technologies deliver accurate, high-throughput organic acid profiling for life science research applications.

References

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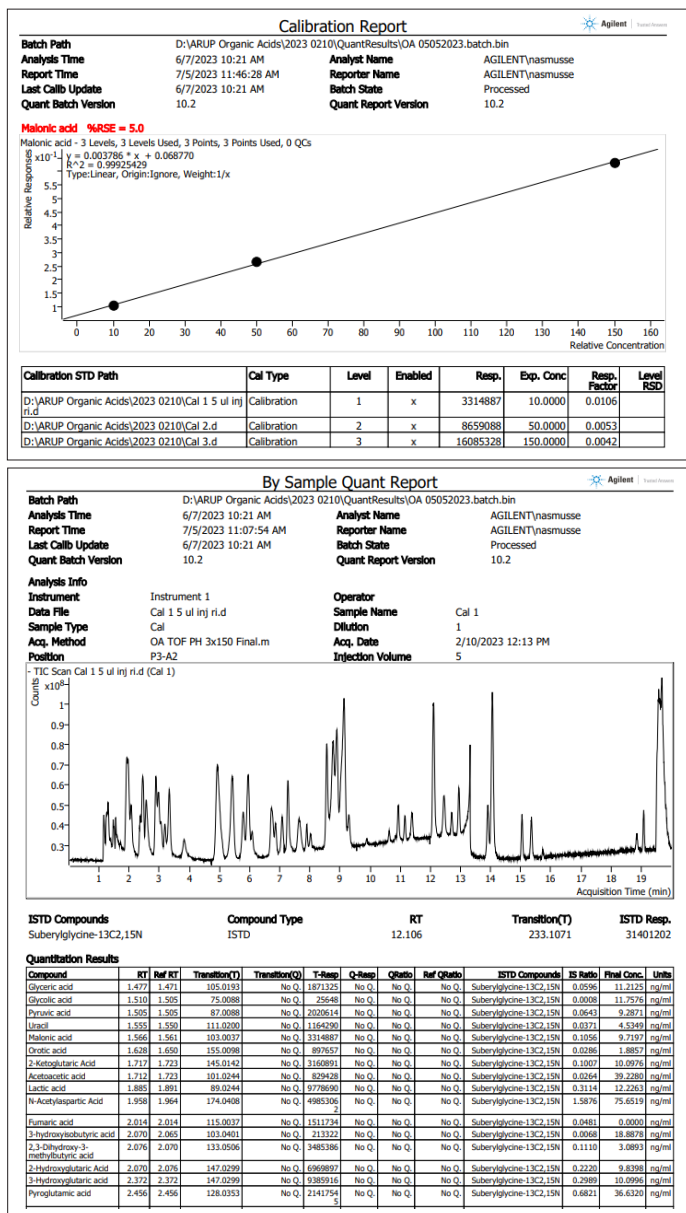


Figure 5. Example reporting results.

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