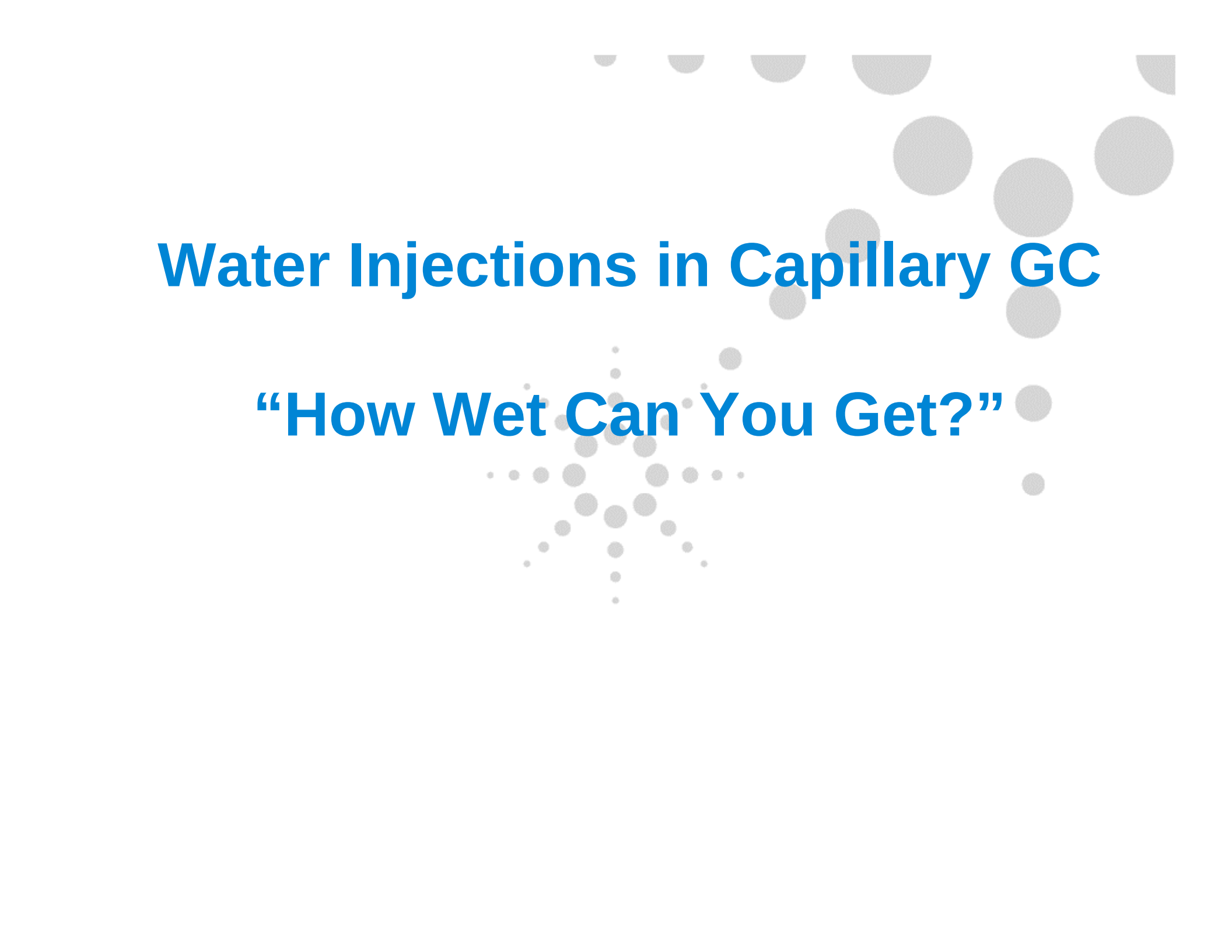




Water Injections in Capillary GC

“How Wet Can You Get?”

Why Inject Water?

- Convenient
 - Aqueous samples (waste water, drinking water, etc.)
 - Biological samples
- Necessary
 - Purge & Trap/Static Headspace



Potential Problems with Water

- Injector Issues
 - Large expansion volume
 - Backflash



Potential Problems with Water

- Detector Issues
 - Extinguish FID flame
 - Decrease sensitivity of ECD
 - “Wet” MSD



Potential Problems with Water

- Solvent - stationary phase mismatch
 - poor wettability of many stationary phases by water
 - “puddles”



Potential Problems with Water

- Damage to stationary phase
 - Change in Retention Times
 - Change in Selectivity
 - Increase in Bleed



What is Normal Column Bleed?

Normal background signal generated by the elution of **normal** degradation products of the column stationary phase



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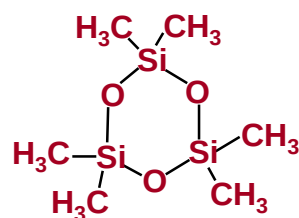
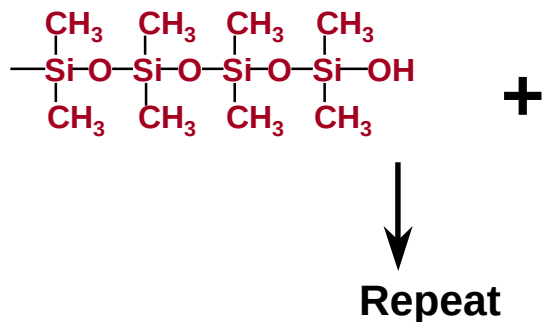
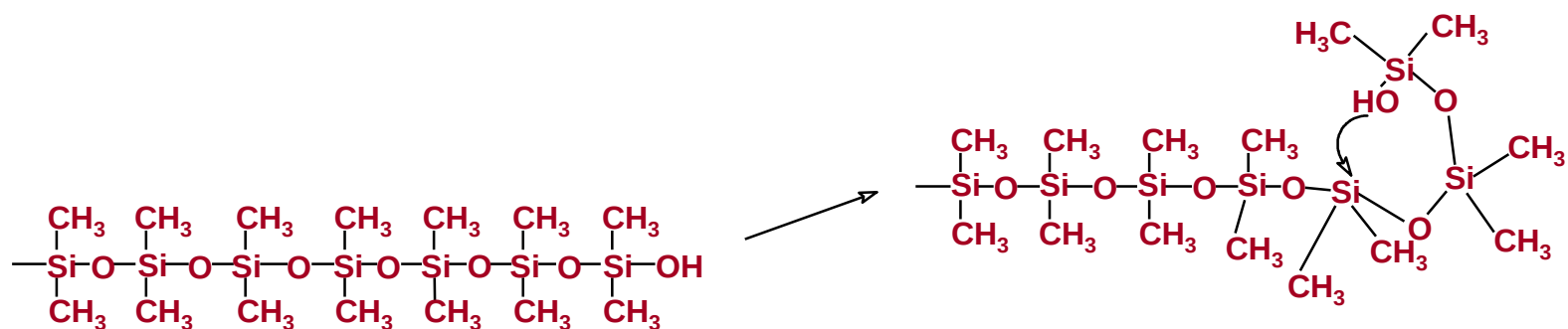
What is Bleed?

- **Thermodynamic equilibrium process occurs to some degree in all columns**
- **Polysiloxane backbone releases low molecular weight, cyclic fragments**
- **Occurs at low level in low temperature, O₂-free, clean system**
- **Increased by increased temperature, oxygen exposure or chemical contamination**



Bleed: Why Does It Happen?

“Back Biting” Mechanism of Product Formation



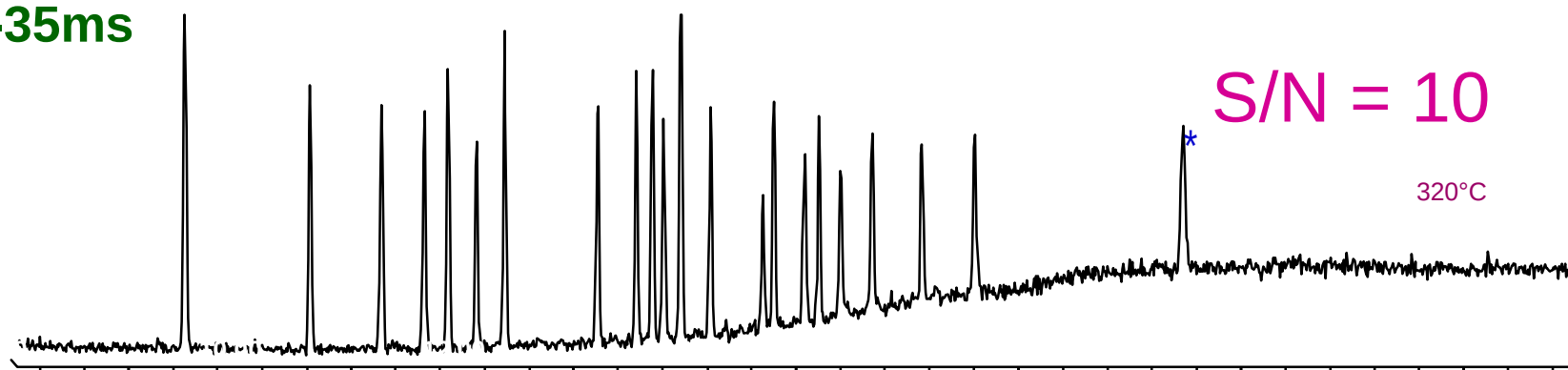
Cyclic products are thermodynamically more stable!



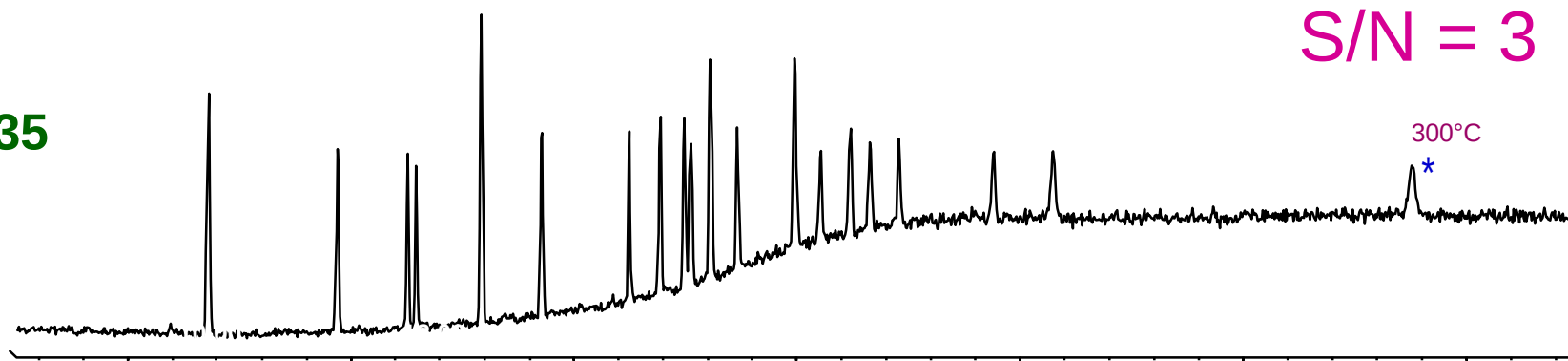
Low Bleed Stationary Phases

DB-35ms vs. DB-35

DB-35ms



DB-35

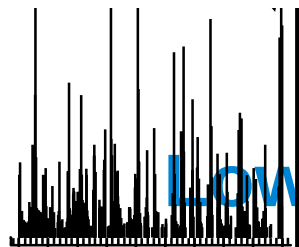


* 5ng Decachlorobiphenyl



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Dial 1-816-650-0621 for e-Seminar Audio



Low Bleed Stationary Phases

Spectral Purity

DB-35ms

Primary Ion

DB-35

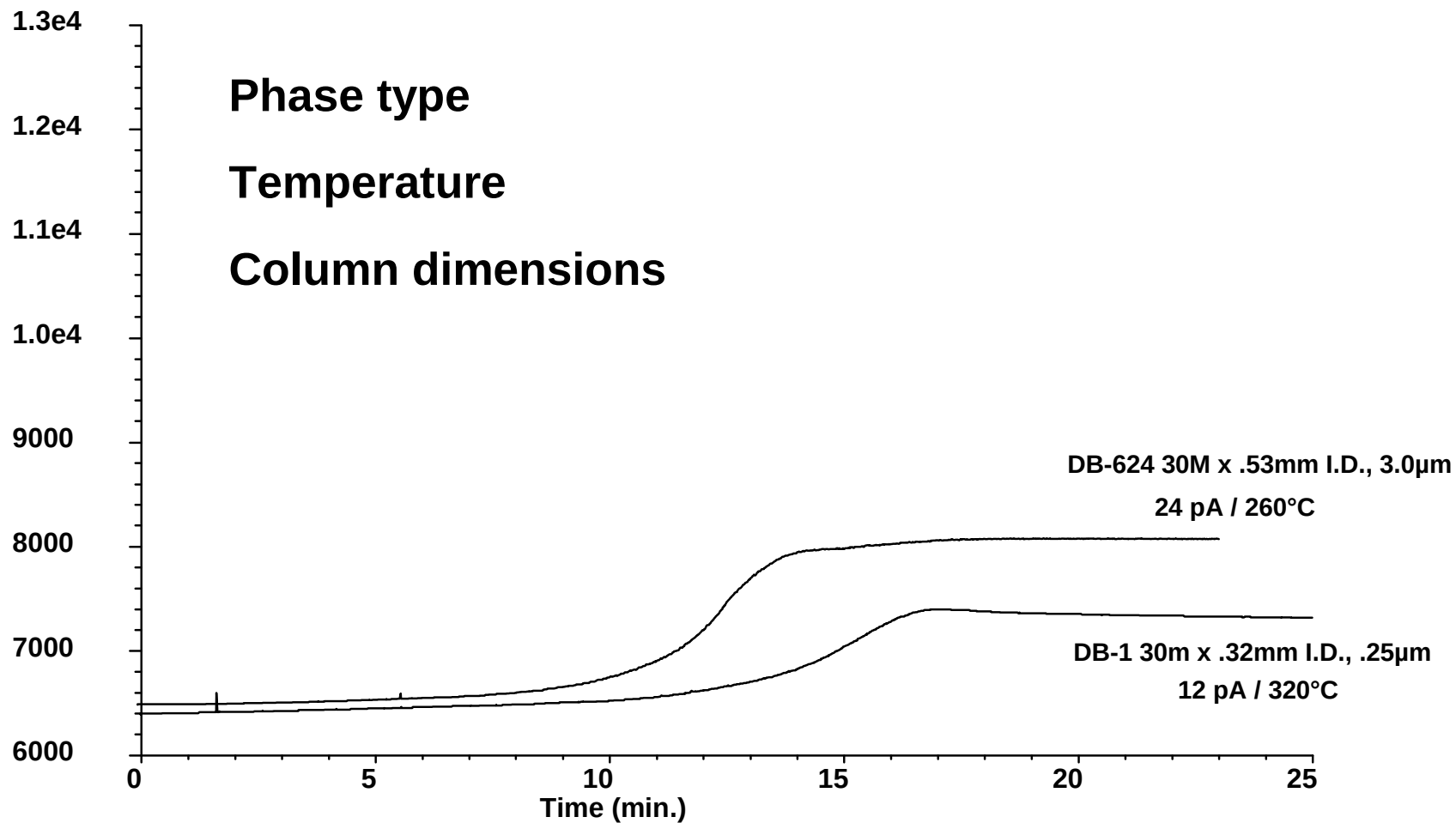
Primary Ion



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Dial 1-816-650-0621 for e-Seminar Audio

Column Bleed is Influenced by:

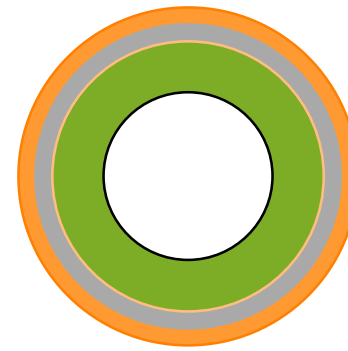
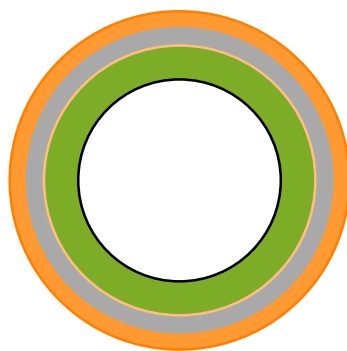
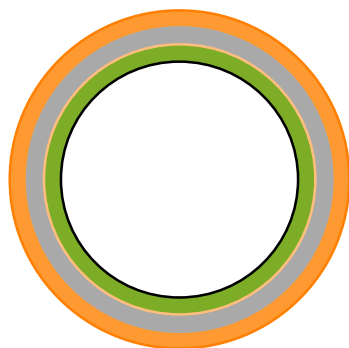


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FILM THICKNESS

Bleed

More stationary phase = More degradation products



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What is a Bleed Problem?

An abnormal elevated baseline at high temperature

IT IS NOT

A high baseline at low temperature

Wandering or drifting baseline at any temperature

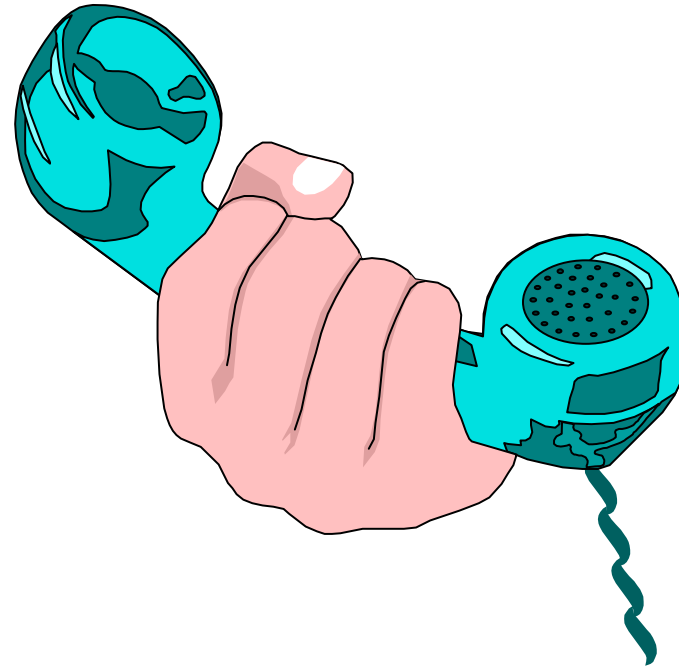
Discrete peaks



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Break

For Questions and Answers
Press 1 on Your Phone to
Ask a Question



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Goals of Water Injection Study

Who cares?

- Frequent Inquiries
- Water does cause problems (but what?)
- Quantitative data
- Establish “guidelines”



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STATIONARY PHASES

- Dimethylpolysiloxane (DB-1)
- Polyethylene glycol (DB-Wax)
- Cyanopropylphenyl (DB-225)
- Cyclodextrin (CycloSil B)
- Divinylbenzene/ethylene glycol dimethacrylate (HP-PLOT U)



Experimental Conditions

- Instrument: Agilent 6890 with Autoinjector
- Injector: 250° C, *1:5 split*
- Injection volume: *1 μ L*
- Detector: FID, 300° C



Experimental Conditions

- Oven: *130° C, 60° C, or 200° C** isothermal
- Column Dimensions: *30 m x 0.53 mm I.D. x 1.0 μm[#]*
- *1,000* water injections at *each* temperature on each column
- Bleed profile after 250, 500, and 1,000 water injections
- Test mix after 250, 500, and 1,000 water injections

*for Cyclosil B and DB-Wax only

[#]CycloSil B: 30 m x 0.32 mm x 0.25 μm



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Test Mixes

Compound List for Chromatograms DB-1

Peak	Compound
1	2-Chlorophenol (skew)
2	Undecane
3	2,4-Dimethylaniline (skew)
4	<u>1-Undecanol (RI 1, skew)</u>
5	Tetradecane
6	<u>Acenaphthalene (RI 2)</u>
7	<i>Pentadecane (k, N)</i>



Test Mixes

Compound List for Chromatograms DB-Wax

Peak	Compound
1	2-Octanone
2	Tetradecane
3	1-Octanol
4	Methyl deconate
5	Methyl undecanoate
6	<u>Naphthalene (RI 1)</u>
7	<u>1-Decanol (RI 2, skew)</u>
8	<i>Methyl dodecanoate (k, N)</i>
9	2,6-Dimethylaniline (skew)
10	2,6-Dimethylphenol (skew)



Test Mixes

Compound List for Chromatograms DB-225

Peak	Compound
1	Tetradecane
2	2-Chlorophenol (skew)
3	Hexadecane
4	<u>Naphthalene (RI 1)</u>
5	2,4-Dimethylaniline (skew)
6	<u>1-Undecanol (RI 2, skew)</u>
7	Octadecane
8	<i>Ethyl dodecanoate (k, N)</i>



Test Mixes

Compound List for Chromatograms CycloSil B

Peak	Compound
1	Dodecane
2	<u>(R)-Linalool (RI 1)</u>
3	<u>(S)-Linalool (RI 2)</u>
4	naphthalene
5	1-nonanol
6	gamma-heptalactone
7	gamma-heptalactone
8	<i>methyl decanoate</i>
9	Tetradecane (k, N)



Test Mixes

Compound List for Chromatograms HP- PLOT U

Peak	Compound
1	Methanol
2	Ethanol
3	Pentane
4	Ethyl ether
5	<u>Acetone (RI 1)</u>
6	Hexane (k, N)
7	<i>Ethyl acetate</i>



Results DB-1

<u>Parameter</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Ret. Factor	14.6	14.5
Ret. Index 1	1349.88	1350.02
Ret. Index 2	1427.77	1428.16
Theor. plates	1448	1474
Bleed (pA)	12.8	11.2

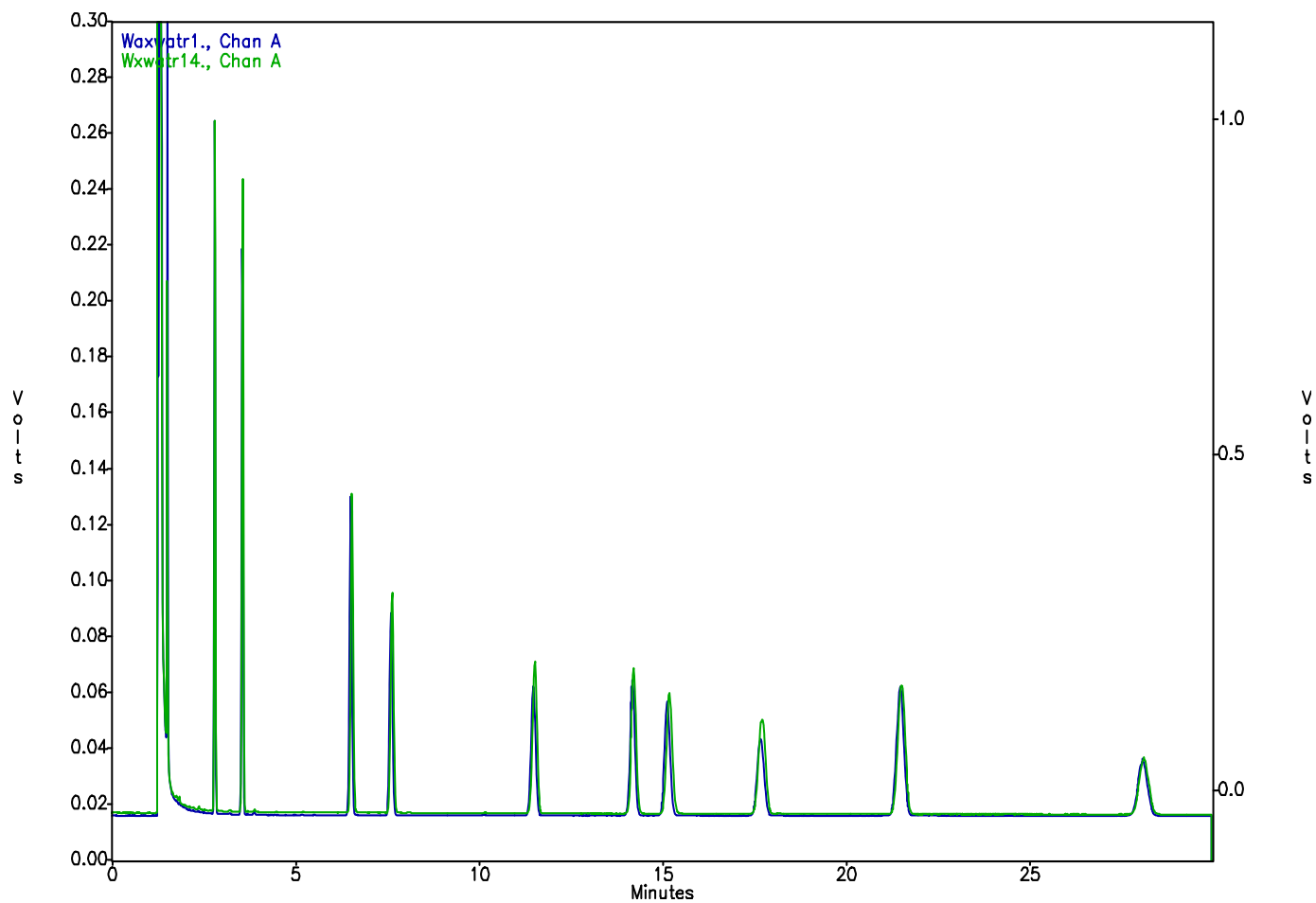


Results DB-Wax

<u>Parameter</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Ret. Factor	12.6	12.6
Ret. Index 1	1149.54	1149.73
Ret. Index 2	1163.44	1163.71
Theor. plates	1277	1261
Bleed (pA)	44.8	32.1



DB-Wax before and **after** 2,000 water injections



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Results DB-225

<u>Parameter</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Ret. Factor	11.5	11.4
Ret. Index 1	1622.30	1621.26
Ret. Index 2	1711.51	1711.03
Theor. plates	1101	1110
Bleed (pA)	34.5	39.3



Results CycloSil B

<u>Parameter</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Ret. Factor	7.8	7.6
Ret. Index	1306.3	1306.0
Resolution	1.9	1.3
Theor. plates	2631	2025
Bleed (pA)	28.4	15.1



Results HP-PLOT U

<u>Parameter</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Ret. Factor	5.2	5.3
Ret. Index	538.0	540.0
Theor. plates	950	982
Bleed (pA)	74.2	35.6



Asymmetry (Skew)

The numbers represent the deviation from an ideal Gaussian, i.e., perfectly symmetrical, peak.

The smaller the number, the better.

Noticeable tailing starts at ~ 0.8 .

Obvious tailing start at ~ 1.2



Asymmetry (Skew)

Skew numbers are based on a modified Gaussian-peak model*. It relies more on the zeroth and first moment calculations, which are less influenced by baseline noise.

*W.W.Yau, Anal. Chem., vol. 49, No. 3 (1977), pp 395-398



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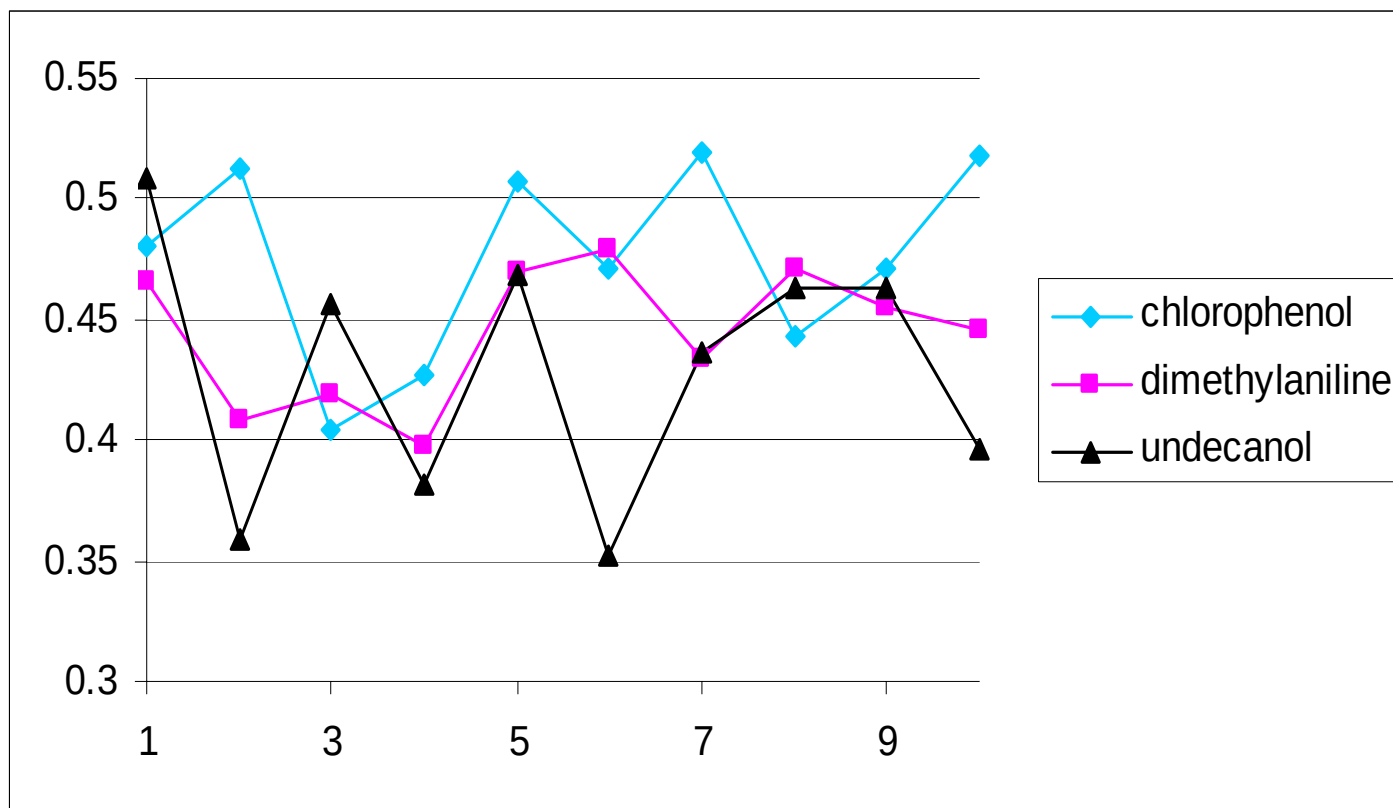
Results DB-1

Asymmetry

<u>Compound</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Chlorophenol	0.48	0.47
Timethylaniline	0.47	0.46
Undecanol	0.51	0.46



Skew Factors DB-1



Results DB-Wax

Asymmetry

<u>Compound</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Decanol	0.38	0.47
Dimethylaniline	0.30	0.32
Dimethylphenol	0.25	0.27



Results DB-225

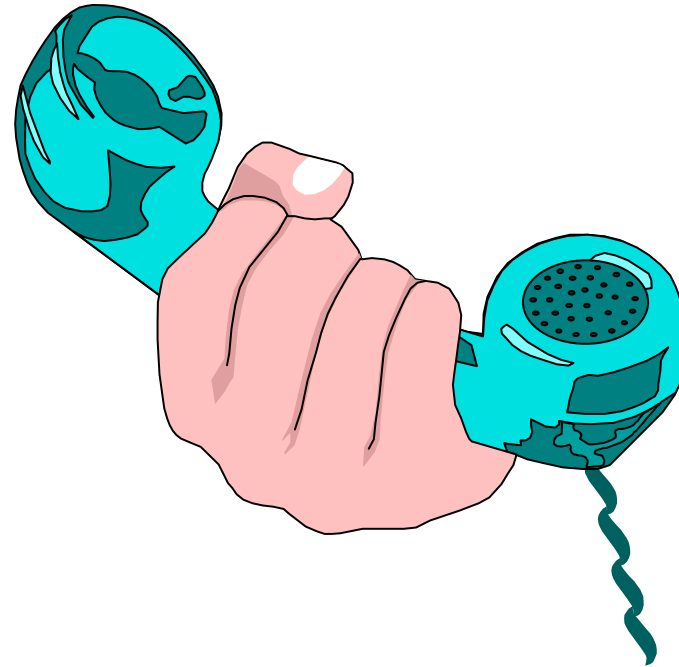
Asymmetry

<u>Compound</u>	<u>Before Inj.</u>	<u>After Inj.</u>
Chlorophenol	0.46	0.47
Dimethylaniline	0.42	0.43
Undecanol	0.63	0.58



Break

For Questions and Answers
Press 1 on Your Phone to
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Summary

For water injections on all columns with bonded stationary phases, no change in:

polarity

selectivity

retention

efficiency

activity

bleed

is observed.



Summary

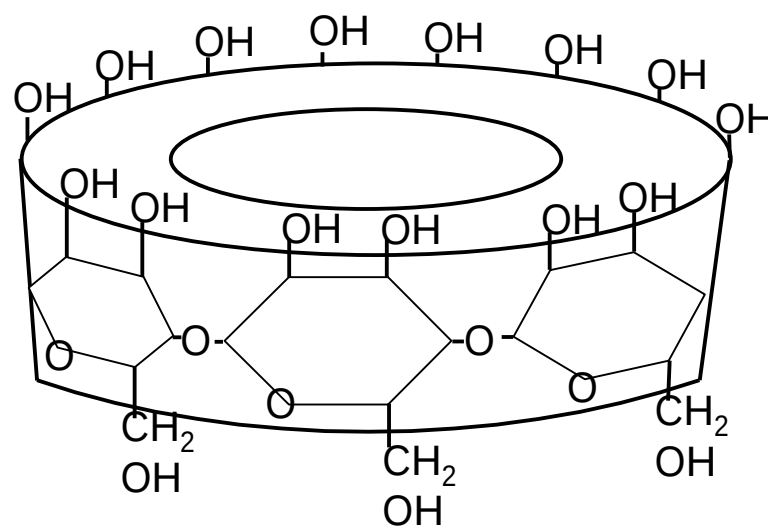
For non-bonded phases, like CycloSil B, water injections can wash out part of the non-bonded stationary phase - loss of resolution, retention, and, possibly, efficiency.



Summary

- Amount of wash-out is dependent on temperature: at 130° C and 200° C it is minimal, at 60° C it is noticeable.
- Solubility of phase material is greater in liquid water.

- 7 Glucose molecules joined through 1,4-glycosidic linkages
- Primary hydroxyl groups partially block the base
- Cavity size is about 6-6.5 Å in diameter
- Hydroxy groups are peralkylated



Summary

Amount of wash-out is gradual!

Most likely dependent on solubility of phase in water (cyclodextrin = high).

No change in selectivity.

No increase in bleed!



Summary

For bonded PLOT columns, no negative effects of injecting water were observed.

Non-bonded PLOT columns (e.g., Alumina and Molesieve), are not suitable for water injections. Observe manufacturers recommendations.



Summary

For all phases, the time to bleed down, or re-condition, a column after injecting water is dependent on the run temperature:

Low-temperature injections will take a lot longer than high-temperature injections.

Periodic bake-out is recommend if injecting water at $<80^{\circ}\text{C}$.



Conclusions

Non-polar columns (bonded and cross-linked) : *safe to inject and rinse with water*

Polar columns (bonded and cross-linked) :
water injections are safe
water-rinsing not recommended

Non-bonded columns: *water injections can wash out stationary phase - use with caution*



Conclusions

Problems associated with water injections are most often caused by injector related phenomena, e.g., backflash.



Solvent Expansion Volumes

Solvent	Vapor volume (μL) of 1 μL liquid
<i>Water</i>	<i>1010</i>
Methanol	450
Carbon disulfide	300
Methylene chloride	285
Acetone	245
n-Hexane	140



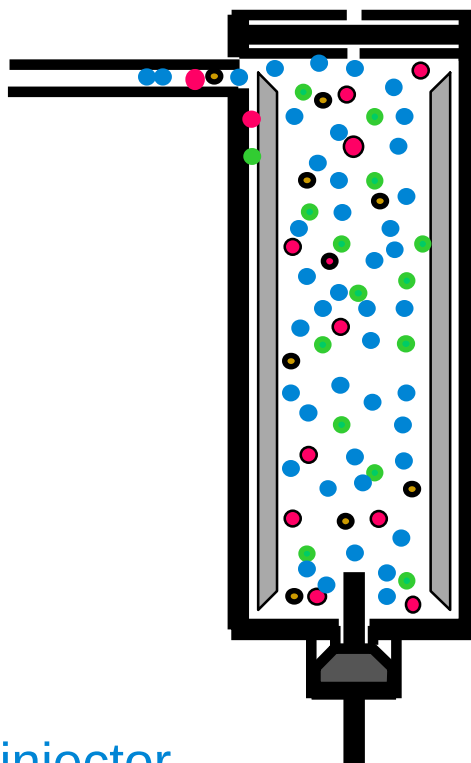
BACKFLASH

Cause

- Vaporized sample expands 100 -1000 X
- Portions may leave the liner
- Occurs when vapor volume > liner volume



BACKFLASH



Sample expands to overfill injector

Some sample condenses on cooler areas (bottom of septum, metal body, etc.)

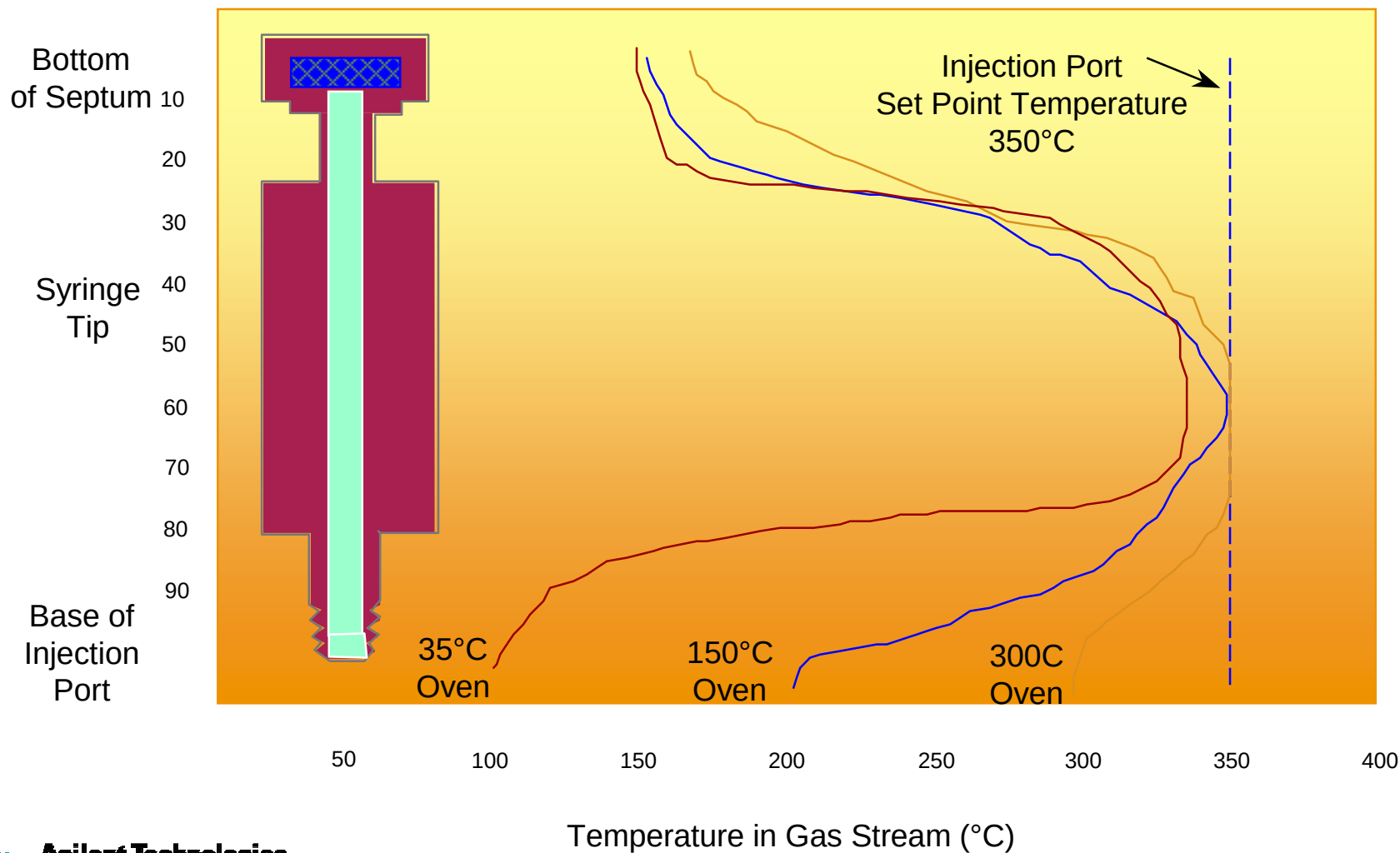
Next injections or carrier gas dislodges condensed sample

Sample then enters the column



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Temperature Profile of a Typical Vaporization Injector vs Oven Temperature



BACKFLASH

Problems

- Loss of sample
- Baseline interferences
- “Ghost” peaks
- Tailing solvent front



BACKFLASH

Minimizing

- Large volume liner
- Small injection volume
- Low expansion solvent
- Low injector temperature
- High carrier gas flow rates
- High head pressures



J&W Scientific GC Columns Technical Support

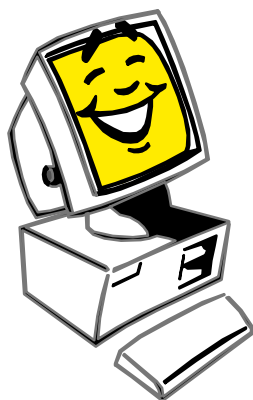
800-227-9770 (phone: US & Canada)*

302-993-5304 (phone)*

** Select option 4, then option 1.*

916-608-1964 (fax)

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